

Patient Name: 김승희
Gender: Female
Sample ID: N25-364

Primary Tumor Site: lung
Collection Date: 2025.12.24

Sample Cancer Type: Lung Cancer

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Relevant Lung Cancer Findings

Gene	Finding	Gene	Finding
ALK	None detected	NTRK1	None detected
BRAF	None detected	NTRK2	None detected
EGFR	None detected	NTRK3	None detected
ERBB2	ERBB2 amplification	RET	None detected
KRAS	None detected	ROS1	None detected
MET	None detected		

Genomic Alteration	Finding
Tumor Mutational Burden	6.64 Mut/Mb measured

Relevant Biomarkers

Tier	Genomic Alteration	Relevant Therapies (In this cancer type)	Relevant Therapies (In other cancer type)	Clinical Trials
IIC	<div>ERBB2 amplification</div> <div>erb-b2 receptor tyrosine kinase 2</div> <div>Locus: chr17:37863255</div>	trastuzumab deruxtecan ¹ / II+	<div>lapatinib + hormone therapy^{1, 2} / I, II+</div> <div>lapatinib + trastuzumab² / I, II+</div> <div>pertuzumab + trastuzumab + chemotherapy^{1, 2} / I, II+</div> <div>trastuzumab + tucatinib¹ / I, II+</div> <div>trastuzumab deruxtecan^{1, 2} / I, II+</div> <div>trastuzumab[†] + chemotherapy^{1, 2} / I, II+</div> <div>trastuzumab[†] + hormone therapy² / I, II+</div> <div>pembrolizumab + trastuzumab + chemotherapy^{1, 2} / I</div> <div>ado-trastuzumab emtansine^{1, 2} / II+</div> <div>lapatinib + chemotherapy^{1, 2} / II+</div> <div>margetuximab + chemotherapy¹ / II+</div> <div>neratinib^{1, 2} / II+</div> <div>neratinib + chemotherapy¹ / II+</div> <div>trastuzumab + tucatinib + chemotherapy^{1, 2} / II+</div> <div>trastuzumab[†]^{1, 2} / II+</div> <div>zanidatamab^{1, 2} / II+</div> <div>pembrolizumab + berahyaluronidase alfa + trastuzumab + chemotherapy¹</div> <div>pertuzumab/trastuzumab/hyaluronidase-zzxf + chemotherapy^{1, 2}</div> <div>trastuzumab and hyaluronidase-oysk¹</div> <div>trastuzumab and hyaluronidase-oysk + chemotherapy¹</div> <div>pertuzumab + trastuzumab^{I, II+}</div> <div>pertuzumab + trastuzumab + hormone therapy^{I, II+}</div> <div>lapatinib + trastuzumab + hormone therapy^I</div> <div>abemaciclib + trastuzumab + hormone therapy^{II+}</div> <div>ado-trastuzumab emtansine + hormone therapy^{II+}</div> <div>hormone therapy^{II+}</div> <div>margetuximab^{II+}</div> <div>pertuzumab + trastuzumab + hormone therapy + chemotherapy^{II+}</div> <div>trastuzumab + hormone therapy + chemotherapy^{II+}</div> <div>ado-trastuzumab emtansine + neratinib</div>	74
IIC	<div>ATM deletion</div> <div>ATM serine/threonine kinase</div> <div>Locus: chr11:108098341</div>	None*	None*	4
IIC	<div>PIK3CA amplification</div> <div>phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha</div> <div>Locus: chr3:178916680</div>	None*	None*	2

* Public data sources included in relevant therapies: FDA¹, NCCN, EMA², ESMO

* Public data sources included in prognostic and diagnostic significance: NCCN, ESMO

† Includes biosimilars/generics

Line of therapy: I: First-line therapy, II+: Other line of therapy

Tier Reference: Li et al. *Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists.* J Mol Diagn. 2017 Jan;19(1):4-23.

Relevant Biomarkers (continued)

Tier	Genomic Alteration	Relevant Therapies (In this cancer type)	Relevant Therapies (In other cancer type)	Clinical Trials
IIC	<i>CHEK1</i> deletion checkpoint kinase 1 Locus: chr11:125496639	None*	None*	1
IIC	<i>FANCM</i> p.(Q1108*) c.3322C>T FA complementation group M Allele Frequency: 74.25% Locus: chr14:45645279 Transcript: NM_020937.4	None*	None*	1

* Public data sources included in relevant therapies: FDA1, NCCN, EMA2, ESMO
* Public data sources included in prognostic and diagnostic significance: NCCN, ESMO
† Includes biosimilars/generics
Line of therapy: I: First-line therapy, II+: Other line of therapy
Tier Reference: Li et al. Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. J Mol Diagn. 2017 Jan;19(1):4-23.

Prevalent cancer biomarkers without relevant evidence based on included data sources

BCL6 amplification, *MSH2* c.645+3A>G, Microsatellite stable, *RB1* p.(L76*) c.227T>A, *TP53* p.(V173L) c.517G>T, *TNFRSF14* deletion, *ENO1* deletion, *PGD* deletion, *MECOM* amplification, *FAT1* p.(S1917Lfs*38) c.5744_5747dup, *HLA-A* p.(L180*) c.539T>A, *MAP3K4* p.(A1195Gfs*16) c.3584_3585delCT, *NQO1* p.(P187S) c.559C>T, Tumor Mutational Burden

Variant Details

DNA Sequence Variants							
Gene	Amino Acid Change	Coding	Variant ID	Locus	Allele Frequency	Transcript	Variant Effect
FANCM	p.(Q1108*)	c.3322C>T	.	chr14:45645279	74.25%	NM_020937.4	nonsense
MSH2	p.(?)	c.645+3A>G	.	chr2:47637514	40.22%	NM_000251.3	unknown
RB1	p.(L76*)	c.227T>A	.	chr13:48881505	59.40%	NM_000321.3	nonsense
TP53	p.(V173L)	c.517G>T	COSM43559	chr17:7578413	58.02%	NM_000546.6	missense
FAT1	p.(S1917Lfs*38)	c.5744_5747dup	.	chr4:187541992	36.77%	NM_005245.4	frameshift Insertion
HLA-A	p.(L180*)	c.539T>A	.	chr6:29911240	98.65%	NM_001242758.1	nonsense
MAP3K4	p.(A1195Gfs*16)	c.3584_3585delCT	.	chr6:161519368	99.77%	NM_005922.4	frameshift Deletion
NQO1	p.(P187S)	c.559C>T	.	chr16:69745145	22.80%	NM_000903.3	missense
FAT1	p.(T4083A)	c.12247A>G	.	chr4:187519136	39.14%	NM_005245.4	missense
CDKN1B	p.(A18G)	c.53C>G	.	chr12:12870826	55.68%	NM_004064.5	missense
RPA1	p.(G434E)	c.1301G>A	.	chr17:1787165	8.00%	NM_002945.5	missense

Copy Number Variations			
Gene	Locus	Copy Number	CNV Ratio
ERBB2	chr17:37863255	25.55	8.66
ATM	chr11:108098341	1	0.74
PIK3CA	chr3:178916680	5.77	2.22

Variant Details (continued)

Copy Number Variations (continued)			
Gene	Locus	Copy Number	CNV Ratio
CHEK1	chr11:125496639	1	0.76
BCL6	chr3:187440209	6.08	2.33
TNFRSF14	chr1:2488070	0.95	0.66
ENO1	chr1:8921399	1.05	0.69
PGD	chr1:10459132	1.05	0.69
MECOM	chr3:168802636	5.69	2.2
RASA2	chr3:141205964	5.4	2.11
ATR	chr3:142168234	5.94	2.28
TP63	chr3:189456442	5.82	2.24
FGFR3	chr4:1801456	0.82	0.62
FAM135B	chr8:139144776	1.05	0.69
KMT2B	chr19:36209128	6.94	2.61

Biomarker Descriptions

ERBB2 amplification

erb-b2 receptor tyrosine kinase 2

Background: The ERBB2 gene encodes the erb-b2 receptor tyrosine kinase 2, a member of the human epidermal growth factor receptor (HER) family¹. Along with ERBB2/HER2, EGFR/ERBB1/HER1, ERBB3/HER3, and ERBB4/HER4 make up the HER protein family¹⁶⁷. All ERBB/HER proteins encode transmembrane receptor tyrosine kinases¹⁶⁸. However, ERBB2/HER2 is an orphan receptor with no known ligand¹⁶⁸. ERBB2 preferentially binds other ligand-bound ERBB/HER family members to form heterodimers resulting in the activation of ERBB2 tyrosine kinase activity and subsequent activation of the PI3K/AKT/MTOR and RAS/RAF/MAPK/ERK signaling pathways which promote cell proliferation, differentiation, and survival¹⁶⁹. Recurrent focal amplification of the ERBB2 gene leads to increased expression in several cancer types¹⁶⁹. ERBB2 overexpression in immortalized cell lines is oncogenic and leads to ERBB2 homo-dimerization and activation without ligand binding^{170,171,172}.

Alterations and prevalence: ERBB2 gene amplification occurs in 10-25% of breast, esophageal, and gastric cancers, 5-10% of bladder, cervical, pancreas, and uterine cancers, and 1-5% of colorectal, lung, and ovarian cancers^{10,11,173,174,175,176,177,178}. ERBB2 gene amplification in pediatric population is observed in 2% of peripheral nervous system cancers (2 in 91 patients) and less than 1% of leukemia (1 in 250 cases)¹¹. Recurrent somatic activating mutations in ERBB2/HER2 occur at low frequencies (<1%) in diverse cancer types^{11,179,180}. In breast, bladder, and colorectal cancers, the most common recurrent ERBB2 activating mutations include kinase domain mutations L755S and V777L and the extracellular domain mutation S310F. In lung cancer, the most common recurrent ERBB2 activating mutations include in-frame exon 20 insertions, particularly Y772_A775dup.

Potential relevance: The discovery of ERBB2/HER2 as an important driver of breast cancer in 1987 led to the development of trastuzumab, a humanized monoclonal antibody with specificity to the extracellular domain of HER2^{181,182}. Trastuzumab¹⁸³ was FDA approved for the treatment of HER2 positive breast cancer in 1998, and subsequently in HER2 positive metastatic gastric and gastroesophageal junction adenocarcinoma in 2010. Additional monoclonal antibody therapies have been approved by the FDA for HER2-positive breast cancer including pertuzumab¹⁸⁴ (2012), a humanized monoclonal antibody that inhibits HER2 dimerization, and ado-trastuzumab emtansine¹⁸⁵ (2013), a conjugate of trastuzumab and a potent antimicrotubule agent. The combination of pertuzumab, trastuzumab, and a taxane is the preferred front-line regimen for HER2-positive metastatic breast cancer¹⁸⁶. In addition to monoclonal antibodies, the small molecule inhibitor lapatinib¹⁸⁷, with specificity for both EGFR and ERBB2, was FDA approved (2007) for the treatment of patients with advanced HER2-positive breast cancer who have received prior therapy including trastuzumab. In 2017, the FDA approved the use of neratinib¹⁸⁸, an irreversible kinase inhibitor of EGFR, ERBB2/HER2, and ERBB4, for the extended adjuvant treatment of adult patients with early stage HER2-positive breast cancer. In 2020, the FDA approved neratinib¹⁸⁸ in combination with capecitabine for HER2-positive advanced or metastatic patients after two or more prior HER2-directed therapies. Also in 2020, the TKI irbinitinib¹⁸⁹ was FDA approved for HER2 overexpressing or amplified breast cancer

Biomarker Descriptions (continued)

in combination with trastuzumab and capecitabine. In 2021, the PD-1 blocking antibody, pembrolizumab, in combination with trastuzumab, fluoropyrimidine- and platinum-based chemotherapy, was approved for HER2 amplified gastric or gastroesophageal (GEJ) adenocarcinoma in the first line⁹⁵. In 2024, a bispecific HER2 antibody, zanidatamab¹⁹⁰, was approved for the treatment of adults with previously treated, unresectable or metastatic ERBB2 overexpressing biliary tract cancer. In 2018 fast track designation was granted to the monoclonal antibody margetuximab¹⁹¹ in patients with ERBB2 positive breast cancer previously treated with an anti-HER2 therapy. Additionally, in 2019, zanidatamab¹⁹², received fast track designation in combination with standard chemotherapy for patients with HER2-overexpressing gastroesophageal adenocarcinoma (GEA). The humanized anti-HER2 antibody drug conjugate disitamab vedotin¹⁹³ (2020) received breakthrough designation for adult patients with HER2-positive urothelial cancer after previous platinum-chemotherapy treatment. In 2021, the antibody-drug conjugate ARX788¹⁹⁴ received fast track designation as a monotherapy for advanced or metastatic HER2-positive breast cancer that have progressed on one or more anti-HER2 regimens. Certain activating mutations have been observed to impart sensitivity to neratinib, afatinib, lapatinib, and trastuzumab, or dacomitinib in early and ongoing clinical studies^{195,196,197,198,199}. ERBB2 kinase domain mutations R896G and V659E both showed response to afatinib in two NSCLC case studies^{200,201}. Additionally, acquired HER2 mutations in estrogen receptor-positive (ER+) breast cancer have been shown to confer resistance to hormone therapy²⁰². However, this was shown to be overcome by neratinib in combination with therapies targeting ER²⁰². Additionally, in 2025, FDA approved the kinase inhibitors zongertinib²⁰³ and sevabertinib²⁰⁴ for the treatment of adult patients with unresectable or metastatic non-squamous non-small cell lung cancer (NSCLC) whose tumors have HER2 tyrosine kinase domain activating mutations. In 2025, a 9 amino acid transmembrane peptide of the HER2/neu protein, GLSI-100 (GP-2)²⁰⁵, received fast track designation for the prevention of breast cancer recurrence following surgery.

ATM deletion

ATM serine/threonine kinase

Background: The ATM gene encodes a serine/threonine kinase that belongs to the phosphatidylinositol-3-kinase related kinases (PIKKs) family of genes that also includes ATR and PRKDC (also known as DNA-PKc)⁶⁹. ATM and ATR act as master regulators of DNA damage response. Specifically, ATM is involved in double-stranded break (DSB) repair while ATR is involved in single-stranded DNA (ssDNA) repair⁷⁰. ATM is recruited to the DNA damage site by the MRE11/RAD50/NBN (MRN) complex that senses DSB^{70,71}. Upon activation, ATM phosphorylates several downstream proteins such as the NBN, MDC1, BRCA1, CHK2 and TP53BP1 proteins⁷². ATM is a tumor suppressor gene and loss of function mutations in ATM are implicated in the BRCAness phenotype, which is characterized by a defect in homologous recombination repair (HRR), mimicking BRCA1 or BRCA2 loss^{6,73}. Germline mutations in ATM often result in Ataxia-telangiectasia, a hereditary disease also referred to as DNA damage response syndrome that is characterized by chromosomal instability⁷⁴.

Alterations and prevalence: Recurrent somatic mutations in ATM are observed in 17% of endometrial carcinoma, 15% of undifferentiated stomach adenocarcinoma, 13% of bladder urothelial carcinoma, 12% of colorectal adenocarcinoma, 9% of melanoma as well as esophagogastric adenocarcinoma and 8% of non-small cell lung cancer^{10,11}.

Potential relevance: The PARP inhibitor, olaparib⁵⁰ is approved (2020) for metastatic castration-resistant prostate cancer (mCRPC) with deleterious or suspected deleterious, germline or somatic mutations in HRR genes that includes ATM. Additionally, talazoparib⁷⁵ in combination with enzalutamide is approved (2023) for metastatic castration-resistant prostate cancer (mCRPC) with mutations in HRR genes that includes ATM. Consistent with other genes associated with the BRCAness phenotype, ATM mutations may aid in selecting patients likely to respond to PARP inhibitors^{73,76,77}. Specifically, in a phase II trial of metastatic, castration-resistant prostate cancer, four of six patients with germline or somatic ATM mutations demonstrated clinical responses to olaparib⁷⁸. However, gene-level analyses from the phase III PROfound trial indicate that ATM-mutated tumors do not experience meaningful radiographic progression-free survival (rPFS) or overall survival (OS) benefit from olaparib, and that the observed survival advantage in the broader HRR-altered population is largely driven by BRCA1/2 alterations rather than ATM^{79,80}. In 2022, the FDA granted fast track designation to the small molecule inhibitor, pidnarulex⁵¹, for BRCA1/2, PALB2, or other homologous recombination deficiency (HRD) mutations in breast and ovarian cancers.

PIK3CA amplification

phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha

Background: The PIK3CA gene encodes the phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha of the class I phosphatidylinositol 3-kinase (PI3K) enzyme¹³⁸. PI3K is a heterodimer that contains a p85 regulatory subunit, which couples one of four p110 catalytic subunits to activated tyrosine protein kinases^{139,140}. The p110 catalytic subunits include p110α, β, δ, γ and are encoded by genes PIK3CA, PIK3CB, PIK3CD, and PIK3CG, respectively¹³⁹. PI3K catalyzes the conversion of phosphatidylinositol (4,5)-bisphosphate (PI(4,5)P2) into phosphatidylinositol (3,4,5)-trisphosphate (PI(3,4,5)P3) while the phosphatase and tensin homolog (PTEN) catalyzes the reverse reaction^{141,142}. The reversible phosphorylation of inositol lipids regulates diverse aspects of cell growth and metabolism^{141,142,143,144}. Recurrent somatic alterations in PIK3CA are frequent in cancer and result in the activation of PI3K/AKT/

Biomarker Descriptions (continued)

MTOR pathway, which can influence several hallmarks of cancer including cell proliferation, apoptosis, cancer cell metabolism and invasion, and genetic instability^{145,146,147}.

Alterations and prevalence: Activating mutations in PIK3CA commonly occur in exons 10 and 21 (previously referred to as exons 9 and 20 due to exon 1 being untranslated)^{148,149}. These mutations typically cluster in the exon 10 helical (codons E542/E545) and exon 21 kinase (codon H1047) domains, each having distinct mechanisms of activation^{150,151,152}. Somatic mutations in PIK3CA are observed in 50% of uterine corpus endometrial carcinoma, 35% of uterine carcinosarcoma, 32% of breast invasive carcinoma, 29% of cervical squamous cell carcinoma, 28% of colorectal adenocarcinoma, 22% of bladder urothelial carcinoma, 17% of head and neck squamous cell carcinoma, 16% of stomach adenocarcinoma, 11% of lung squamous cell carcinoma, 9% of esophageal adenocarcinoma, 8% of brain lower grade glioma, 6% of cholangiocarcinoma, 5% of skin cutaneous melanoma and lung adenocarcinoma, 4% of liver hepatocellular carcinoma, 3% of pancreatic adenocarcinoma and sarcoma, and 2% of mesothelioma, prostate adenocarcinoma, testicular germ cell tumors, and ovarian serous cystadenocarcinoma^{10,11}. PIK3CA is amplified in 38% of lung squamous cell carcinoma, 20% of ovarian serous cystadenocarcinoma, 18% of esophageal adenocarcinoma, 16% of head and neck squamous cell carcinoma, 15% of cervical squamous cell carcinoma, 11% of uterine carcinosarcoma, 7% of uterine corpus endometrial carcinoma, 5% of stomach adenocarcinoma, 4% of bladder urothelial carcinoma, 3% of breast invasive carcinoma and pancreatic adenocarcinoma, and 2% of prostate adenocarcinoma, lung adenocarcinoma, and kidney renal clear cell carcinoma^{10,11}. Alterations in PIK3CA are also observed in pediatric cancers¹¹. Somatic mutations in PIK3CA are observed in 6% of non-Hodgkin Lymphoma (1 in 17 cases), 4% of glioma (11 in 297 cases), 3% of soft tissue sarcoma (1 in 38 patients), 2% of embryonal tumors (6 in 332 cases), 1% of leukemia (5 in 354 cases), and less than 1% of bone cancer (3 in 327 cases), B-lymphoblastic leukemia/lymphoma (2 in 252 cases), and peripheral nervous system tumors (1 in 1158 cases)¹¹.

Potential relevance: The PI3K inhibitor, alpelisib¹⁵³, is FDA-approved (2019) in combination with fulvestrant for the treatment of patients with PIK3CA-mutated, hormone receptor (HR)-positive, human epidermal growth factor receptor 2 (HER2)-negative, advanced or metastatic breast cancer. Specifically, exon 21 H1047R mutations were associated with more durable clinical responses in comparison to exon 10 E545K mutations¹⁵⁴. However, alpelisib did not improve response when administered with letrozole in patients with ER + early breast cancer with PIK3CA mutations¹⁵⁵. The FDA also approved the kinase inhibitor, capivasertib (2023)¹⁵⁶ in combination with fulvestrant for locally advanced or metastatic HR-positive, HER2-negative breast cancer with one or more PIK3CA/AKT1/PTEN-alterations following progression after endocrine treatment. The kinase inhibitor, inavolisib¹⁵⁷, is also FDA-approved (2024) in combination with palbociclib and fulvestrant for the treatment of adults with endocrine-resistant, PIK3CA-mutated, HR-positive, and HER2-negative breast cancer. Case studies with mTOR inhibitors sirolimus and temsirolimus report isolated cases of clinical response in PIK3CA mutated refractory cancers^{158,159}. In colorectal cancers, PIK3CA mutations predict significantly improved survival and reduced disease recurrence with adjuvant aspirin therapy, compared to no benefit in wild-type PIK3CA tumors^{127,133,160,161}. In 2025, the FDA granted fast track designation to the PI3Ka inhibitor and degrader, ETX-636¹⁶², for the treatment of PIK3CA-mutant, HR-positive/HER-negative advanced breast cancer.

CHEK1 deletion

checkpoint kinase 1

Background: The CHEK1 gene encodes the checkpoint kinase 1 protein and belongs to a family of serine/threonine checkpoint kinases, that also includes CHEK2¹. Checkpoint kinases play an important role in S phase and G2/M transition and DNA damage induced cell cycle arrest⁴⁵. CHEK1 is a tumor suppressor and it interacts with proteins involved in transcription regulation, cell-cycle arrest, and DNA repair including homologous recombination repair (HRR)^{46,47}. Upon DNA damage, CHEK1 is phosphorylated and activated by DNA damage repair proteins ATM and ATR⁴⁶. Activated CHEK1 subsequently phosphorylates and negatively regulates downstream proteins such as CDC25A thereby slowing or stalling DNA replication^{46,48}.

Alterations and prevalence: Recurrent somatic alterations of CHEK1 include mutations and copy number loss. Somatic mutations of CHEK1 are observed in 3% of endometrial carcinoma, 2% of non-small cell lung cancer and 1% of cervical squamous carcinoma cases^{10,49}. CHEK1 copy number loss occurs in 10% of seminoma, 8% of non-seminomatous germ cell tumor, 5% of ocular melanoma, and 3% of melanoma cases^{10,49}.

Potential relevance: The PARP inhibitor, olaparib⁵⁰ is approved (2020) for metastatic castration-resistant prostate cancer (mCRPC) with deleterious or suspected deleterious, germline or somatic mutations in HRR genes that includes CHEK1. In 2022, the FDA granted fast track designation to the small molecule inhibitor, pidnarulex⁵¹, for BRCA1/2, PALB2, or other homologous recombination deficiency (HRD) mutations in breast and ovarian cancers.

FANCM p.(Q1108*) c.3322C>T

FA complementation group M

Background: The FANCM gene encodes the FA complementation group M protein, a member of the Fanconi Anemia (FA) family, which also includes FANCA, FANCB, FANCC, FANCD1 (BRCA2), FANCD2, FANCE, FANCF, FANCG, FANCI, FANCI (BRIP1), FANCL, and

Biomarker Descriptions (continued)

FANCN (PALB2)¹. FA genes are tumor suppressors that are responsible for the maintenance of replication fork stability, DNA damage repair through the removal of interstrand cross-links (ICL), and subsequent initiation of the homologous recombination repair (HRR) pathway^{2,3}. In response to DNA damage, FANCA, FANCB, FANCC, FANCE, FANCF, FANCG, FANCL, and FANCM assemble to form the FA core complex which is responsible for the monoubiquitination of the FANCI-FANCD2 (ID2) complex². Monoubiquitination of the ID2 complex promotes co-localization with BRCA1/2, which is critical in BRCA mediated DNA repair^{4,5}. Loss of function mutations in the FA family and HRR pathway can result in the BRCAness phenotype, characterized by a defect in the HRR pathway, mimicking BRCA1 or BRCA2 loss^{6,7}. Germline mutations in FA genes lead to Fanconi Anemia, a condition characterized by chromosomal instability and congenital abnormalities, including bone marrow failure and cancer predisposition^{8,9}.

Alterations and prevalence: Somatic mutations in FANCM are observed in 11% of uterine corpus endometrial carcinoma, 8% of skin cutaneous melanoma, 7% of lung adenocarcinoma, 6% of stomach adenocarcinoma, 5% colorectal adenocarcinoma, uterine carcinosarcoma, and bladder urothelial carcinoma^{10,11}.

Potential relevance: Currently, no therapies are approved for FANCM aberrations. Consistent with other genes that contribute to the BRCAness phenotype, mutations in FANCM are shown to confer enhanced sensitivity in vitro to PARP inhibitors such as olaparib¹².

BCL6 amplification

B-cell CLL/lymphoma 6

Background: The BCL6 gene encodes the B-cell lymphoma 6 (BCL6) transcription repressor, a protein that is responsible for inhibiting the expression of several genes including those involved in the DNA damage response, cell cycle checkpoints, and modulating BCL2 expression^{52,53,54}. BCL6 is most commonly expressed in germinal center B-cells and is required for germinal cell formation and affinity maturation during T-cell dependent antibody responses⁵³. BCL6 is observed to competitively bind DNA motifs recognized by the oncogenic transcription factor STAT6, thereby repressing STAT6 mediated gene transcription^{55,56}. Aberrations in BCL6 often lead to altered target gene transcription, including those involved in cell cycle arrest, differentiation, and apoptosis^{52,53}.

Alterations and prevalence: BCL6 rearrangement most commonly occurs with immunoglobulin H (IGH) partners and results in the truncation or removal the BCL6 promoter region and juxtaposition of BCL6 downstream of the partner gene promoter⁵⁷. Replacement of the BCL6 promoter resulting from such translocations has been observed to lead to aberrant BCL6 expression⁵⁸. BCL6 rearrangement is a common event in lymphoma and has been observed in up to 40% of diffuse large B-cell lymphoma (DLBCL) and 15% of follicle center lymphomas^{53,57}. Somatic mutations in BCL6 are observed in 7% of uterine corpus endometrial carcinoma, 4% of skin cutaneous melanoma, and 3% of stomach adenocarcinoma and colorectal adenocarcinoma, and 2% of uterine carcinosarcoma, lung adenocarcinoma, and sarcoma^{10,11}. Mutations in the 5' regulatory sequences of BCL6 are observed in 30-40% of germinal center B-cells and are believed to disrupt BCL6 negative autoregulation⁵³. Amplifications are observed in 31% of lung squamous cell carcinoma, 16% of esophageal adenocarcinoma and ovarian serous cystadenocarcinoma, and 14% of head and neck and cervical squamous cell carcinoma, 9% of uterine carcinosarcoma, 6% of uterine corpus endometrial carcinoma, and 2-4% of stomach adenocarcinoma, diffuse large B-cell lymphoma, bladder urothelial carcinoma, breast invasive carcinoma, testicular germ cell tumors, liver hepatocellular carcinoma, and pancreatic adenocarcinoma^{10,11}. Alterations in BCL6 are rare in pediatric cancers^{10,11}. Somatic mutations in BCL6 are observed in 3% of soft tissue sarcoma, and less than 1% of bone cancer (3 in 327 cases), embryonal tumors (2 in 332 cases), and glioma (1 in 297 cases)^{10,11}. Amplification of BCL6 is observed in 1% or less of Wilms tumor (2 in 136 cases) and B-lymphoblastic leukemia/lymphoma (1 in 731 cases)^{10,11}.

Potential relevance: B-cell lymphoma with BCL6 translocations that co-occur with MYC are referred to as double-hit lymphoma (DHL), while co-occurrence with MYC and BCL2 rearrangements is referred to as triple-hit lymphoma⁵⁹. Such concomitant rearrangements are recognized by the World Health Organization (WHO) as diagnostic entity of diffuse large B-cell lymphoma/high grade B-cell lymphoma (HGBL) with MYC and BCL2 rearrangements⁶⁰. DHL expressing BCL6 rearrangements are most often aggressive with poor prognosis, involve extra nodal sites, and have a germinal center phenotype^{61,62}.

MSH2 c.645+3A>G

mutS homolog 2

Background: The MSH2 gene encodes the mutS homolog 2 protein¹. MSH2 is a tumor suppressor gene that heterodimerizes with MSH6 to form the MutSa complex or with MSH3 to form the MutSβ complex⁸¹. Both MutS complexes function in DNA damage recognition of base-base mismatches or insertion/deletion (indels) mispairs⁸¹. Specifically, the MutSa complex recognizes 1-2 nucleotide indels while MutSβ recognizes longer indel mispairs⁸¹. DNA damage recognition initiates the mismatch repair (MMR) process that repairs mismatch errors which typically occur during DNA replication⁸¹. Mutations in MSH2 result in the degradation of MSH6⁸². Loss of MSH2 protein expression correlates with mutations in the genes and are used to pre-screen colorectal cancer or endometrial hyperplasia⁸³. MSH2, along with MLH1, MSH6, and PMS2, form the core components of the MMR pathway⁸⁴. Deficiency in MMR (dMMR) is characterized by mutations and loss of expression in these genes⁸⁵. dMMR is associated with microsatellite instability (MSI), which is defined as a change in the length of a microsatellite in a tumor as compared to normal tissue^{84,86,87}. MSI-high (MSI-

Biomarker Descriptions (continued)

H) is a hallmark of Lynch Syndrome (LS), also known as hereditary non-polyposis colorectal cancer, which is caused by germline mutations in MMR genes^{84,88}. LS is associated with an increased risk of developing colorectal cancer, as well as other cancers, including endometrial and stomach cancer.^{86,88,89,90} Specifically, MSH2 mutations are associated with an increased risk of ovarian and pancreatic cancer^{91,92,93,94}.

Alterations and prevalence: Somatic mutations in MSH2 are observed in 8% of uterine corpus endometrial carcinoma, as well as 2-3% of bladder urothelial carcinoma, melanoma, and colorectal adenocarcinoma^{10,11}. Alterations in MSH2 are observed in pediatric cancers^{10,11}. Somatic mutations are observed in 3% of soft tissue sarcoma, 1% of embryonal tumor, and less than 1% of B-lymphoblastic leukemia/lymphoma (2 in 252 cases), glioma (2 in 297 cases), leukemia (2 in 311 cases), bone cancer (2 in 327 cases), and peripheral nervous system tumors (1 in 1158 cases)^{10,11}.

Potential relevance: Pembrolizumab (2014) is an anti-PD-1 immune checkpoint inhibitor that is approved for patients with dMMR solid tumors that have progressed on prior therapies⁹⁵. Nivolumab (2015), an anti-PD-1 immune checkpoint inhibitor, is approved alone or in combination with the cytotoxic T-lymphocyte antigen 4 (CTLA-4) blocking antibody, ipilimumab (2011), for patients with dMMR colorectal cancer that have progressed on prior treatment^{96,97}. MSH2 mutations are consistent with high grade in pediatric diffuse gliomas^{98,99}.

Microsatellite stable

Background: Microsatellites are short tandem repeats (STR) of 1 to 6 bases of DNA between 5 to 50 repeat units in length. There are approximately 0.5 million STRs that occupy 3% of the human genome¹²⁴. Microsatellite instability (MSI) is defined as a change in the length of a microsatellite in a tumor as compared to normal tissue^{86,88}. MSI is closely tied to the status of the mismatch repair (MMR) genes. In humans, the core MMR genes include MLH1, MSH2, MSH6, and PMS2⁸⁷. Mutations and loss of expression in MMR genes, known as defective MMR (dMMR), lead to MSI. In contrast, when MMR genes lack alterations, they are referred to as MMR proficient (pMMR). Consensus criteria were first described in 1998 and defined MSI-high (MSI-H) as instability in two or more of the following five markers: BAT25, BAT26, D5S346, D2S123, and D17S250¹²⁵. Tumors with instability in one of the five markers were defined as MSI-low (MSI-L) whereas, those with instability in zero markers were defined as MS-stable (MSS)¹²⁵. Tumors classified as MSI-L are often phenotypically indistinguishable from MSS tumors and tend to be grouped with MSS^{89,126,127,128,129}. MSI-H is a hallmark of Lynch syndrome (LS), also known as hereditary non-polyposis colorectal cancer, which is caused by germline mutations in the MMR genes⁸⁸. LS is associated with an increased risk of developing colorectal cancer, as well as other cancers, including endometrial and stomach cancer^{86,88,89,90}.

Alterations and prevalence: The MSI-H phenotype is observed in 30% of uterine corpus endometrial carcinoma, 20% of stomach adenocarcinoma, 15-20% of colon adenocarcinoma, and 5-10% of rectal adenocarcinoma^{86,88,130,131}. MSI-H is also observed in 5% of adrenal cortical carcinoma and at lower frequencies in other cancers such as esophageal, liver, and ovarian cancers^{130,131}.

Potential relevance: Anti-PD-1 immune checkpoint inhibitors including pembrolizumab⁹⁵ (2014) and nivolumab⁹⁶ (2015) are approved for patients with MSI-H or dMMR colorectal cancer who have progressed following chemotherapy. Pembrolizumab⁹⁵ is also approved as a single agent, for the treatment of patients with advanced endometrial carcinoma that is MSI-H or dMMR with disease progression on prior therapy who are not candidates for surgery or radiation. Importantly, pembrolizumab is approved for the treatment of MSI-H or dMMR solid tumors that have progressed following treatment, with no alternative option and is the first anti-PD-1 inhibitor to be approved with a tumor agnostic indication⁹⁵. Dostarlimab¹³² (2021) is also approved for dMMR recurrent or advanced endometrial carcinoma or solid tumors that have progressed on prior treatment and is recommended as a subsequent therapy option in dMMR/MSI-H advanced or metastatic colon or rectal cancer^{127,133}. The cytotoxic T-lymphocyte antigen 4 (CTLA-4) blocking antibody, ipilimumab⁹⁷ (2011), is approved alone or in combination with nivolumab in MSI-H or dMMR colorectal cancer that has progressed following treatment with chemotherapy. MSI-H may confer a favorable prognosis in colorectal cancer although outcomes vary depending on stage and tumor location^{127,134,135}. Specifically, MSI-H is a strong prognostic indicator of better overall survival (OS) and relapse free survival (RFS) in stage II as compared to stage III colorectal cancer patients¹³⁵. The majority of patients with tumors classified as either MSS or pMMR do not benefit from treatment with single-agent immune checkpoint inhibitors as compared to those with MSI-H tumors^{136,137}. However, checkpoint blockade with the addition of chemotherapy or targeted therapies have demonstrated response in MSS or pMMR cancers^{136,137}.

RB1 p.(L76*) c.227T>A

RB transcriptional corepressor 1

Background: The RB1 gene encodes the retinoblastoma protein (pRB), and is an early molecular hallmark of cancer³⁷. RB1 belongs to the family of pocket proteins that also includes p107 and p130, which play a crucial role in the cell proliferation, apoptosis, and differentiation^{37,38}. RB1 is well characterized as a tumor suppressor gene that restrains cell cycle progression from G1 phase to S phase³⁹. Specifically, RB1 binds and represses the E2F family of transcription factors that regulate the expression of genes involved in

Biomarker Descriptions (continued)

the G1/S cell cycle regulation^{37,38,40}. Germline mutations in RB1 are associated with retinoblastoma (a rare childhood tumor) as well as other cancer types such as osteosarcoma, soft tissue sarcoma, and melanoma⁴¹.

Alterations and prevalence: Recurrent somatic alterations in RB1, including mutations and biallelic loss, lead to the inactivation of the RB1 protein. RB1 mutations are observed in 20% of bladder urothelial carcinoma, 13% of uterine corpus endometrial carcinoma, and 10% of sarcoma and glioblastoma multiforme^{10,11}. Biallelic loss of RB1 is also observed in several cancers including 15% of sarcoma, 10% of prostate adenocarcinoma, 9% of uterine carcinosarcoma, ovarian serous cystadenocarcinoma, and bladder urothelial carcinoma, 5% of liver hepatocellular carcinoma and adrenocortical carcinoma, and 4% of esophageal adenocarcinoma, diffuse large B-cell lymphoma, and breast invasive carcinoma^{10,11}. Biallelic loss of the RB1 gene is also linked to the activation of chemotherapy-induced acute myeloid leukemia (AML) and acute lymphoblastic leukemia (ALL)^{42,43,44}. Alterations in RB1 are also observed in pediatric cancers¹¹. Somatic mutations in RB1 are observed in 52% of retinoblastoma (16 in 31 cases), 3% of bone cancer (10 in 327 cases), and less than 1% of B-lymphoblastic leukemia/lymphoma (2 in 252 cases), glioma (2 in 297 cases), and leukemia (2 in 311 cases)¹¹. Biallelic deletion of RB1 is observed in 5% of bone cancer (2 in 42 cases), 4% of B-lymphoblastic leukemia/lymphoma (28 in 731 cases), 3% of leukemia (7 in 250 cases), and less than 1% of Wilms tumor (1 in 136 cases)¹¹. Structural variants in RB1 are observed in 3% of bone cancer (5 in 150 cases)¹¹.

Potential relevance: Currently, there are no therapies approved for RB1 aberrations.

TP53 p.(V173L) c.517G>T

tumor protein p53

Background: The TP53 gene encodes the tumor suppressor protein p53, which binds to DNA and activates transcription in response to diverse cellular stresses to induce cell cycle arrest, apoptosis, or DNA repair¹. In unstressed cells, TP53 is kept inactive by targeted degradation via MDM2, a substrate recognition factor for ubiquitin-dependent proteolysis¹⁰⁵. Alterations in TP53 are required for oncogenesis as they result in loss of protein function and gain of transforming potential¹⁰⁶. Germline mutations in TP53 are the underlying cause of Li-Fraumeni syndrome, a complex hereditary cancer predisposition disorder associated with early-onset cancers^{107,108}.

Alterations and prevalence: TP53 is the most frequently mutated gene in the cancer genome with approximately half of all cancers experiencing TP53 mutations. Ovarian, head and neck, esophageal, and lung squamous cancers have particularly high TP53 mutation rates (60-90%)^{10,11,109,110,111,112}. Approximately two-thirds of TP53 mutations are missense mutations and several recurrent missense mutations are common, including substitutions at codons R158, R175, Y220, R248, R273, and R282^{10,11}. Invariably, recurrent missense mutations in TP53 inactivate its ability to bind DNA and activate transcription of target genes^{113,114,115,116}. Alterations in TP53 are also observed in pediatric cancers^{10,11}. Somatic mutations are observed in 53% of non-Hodgkin lymphoma, 24% of soft tissue sarcoma, 19% of glioma, 13% of bone cancer, 9% of B-lymphoblastic leukemia/lymphoma, 4% of embryonal tumors, 3% of Wilms tumor and leukemia, 2% of T-lymphoblastic leukemia/lymphoma, and less than 1% of peripheral nervous system cancers (5 in 1158 cases)^{10,11}. Biallelic loss of TP53 is observed in 10% of bone cancer, 2% of Wilms tumor, and less than 1% of B-lymphoblastic leukemia/lymphoma (2 in 731 cases) and leukemia (1 in 250 cases)^{10,11}.

Potential relevance: The small molecule p53 reactivator, PC14586¹¹⁷ (2020), received a fast track designation by the FDA for advanced tumors harboring a TP53 Y220C mutation. In addition to investigational therapies aimed at restoring wild-type TP53 activity, compounds that induce synthetic lethality are also under clinical evaluation^{118,119}. TP53 mutations are a diagnostic marker of SHH-activated, TP53-mutant medulloblastoma¹²⁰. TP53 mutations confer poor prognosis and poor risk in multiple blood cancers including AML, MDS, myeloproliferative neoplasms (MPN), and chronic lymphocytic leukemia (CLL), and acute lymphoblastic leukemia (ALL)^{21,22,25,121,122}. In mantle cell lymphoma, TP53 mutations are associated with poor prognosis when treated with conventional therapy including hematopoietic cell transplant¹⁰². Mono- and bi-allelic mutations in TP53 confer unique characteristics in MDS, with multi-hit patients also experiencing associations with complex karyotype, few co-occurring mutations, and high-risk disease presentation as well as predicted death and leukemic transformation independent of the IPSS-R staging system¹²³.

TNFRSF14 deletion

TNF receptor superfamily member 14

Background: The TNFRSF14 gene encodes TNF receptor superfamily member 14¹. TNFRSF14, also known as HVEM, belongs to the tumor necrosis factor superfamily of cell surface receptors (TNFRSF), which interact with the tumor necrosis factor superfamily (TNFSF) of cytokines¹⁰⁰. TNFSF-TNFRSF interactions regulate several signaling pathways, including those involved in immune cell differentiation, survival, and death¹⁰⁰. TNFRSF14 can be stimulated by several ligands, including the TNFSF14 ligand (also known as LIGHT), BTLA, and CD160^{100,101}. Following ligand binding to TNFRSF in T-cells, TNFRSF proteins aggregate at the cell membrane and initiate co-signaling cascades which promotes activation, differentiation, and survival¹⁰⁰. In lymphoma, binding of TNFRSF14 by TNFSF14 has been observed to enhance Fas-induced apoptosis, suggesting a tumor suppressor role¹⁰¹.

Biomarker Descriptions (continued)

Alterations and prevalence: Somatic mutations in TNFRSF14 are observed in 5% of diffuse large B-cell lymphoma (DLBCL), and 2% of skin cutaneous melanoma^{10,11}. Biallelic loss of TNFRSF14 occurs in 8% of DLBCL and uveal melanoma, 3% of cholangiocarcinoma, and 2% of adrenocortical carcinoma and liver hepatocellular carcinoma^{10,11}.

Potential relevance: Currently, no therapies are approved for TNFRSF14 aberrations. Somatic mutations in TNFRSF14 are diagnostic for follicular lymphoma¹⁰². In addition, TNFRSF14 mutations are associated with poor prognosis in follicular lymphoma^{103,104}.

ENO1 deletion

enolase 1

Background: The ENO1 gene encodes enolase 1 and its alternatively spliced protein isoform, c-MYC promoter binding protein 1 (MBP1)^{1,163}. ENO1 is a glycolytic enzyme that catalyzes the dehydration of 2-phosphoglyceric acid to phosphoenolpyruvic acid during glycolysis¹⁶³. In addition to its role in glycolysis, ENO1 acts as a cell surface plasminogen receptor and is involved in cytoskeleton reorganization, stabilization of the mitochondrial membrane, and modulation of several oncogenic pathways, including PI3K/AKT, AMPK/mTOR and Wnt/ β -catenin^{163,164,165}. ENO1 has been found to be overexpressed in various cancers contributing to upregulation of glycolysis, cancer cell survival and proliferation, chemoresistance, extracellular matrix degradation, migration, invasion, and metastases^{163,164,166}. In contrast, MBP1 is known to repress c-MYC transcription under cellular stress and low glucose conditions, leading to suppression of cellular proliferation, migration, and invasion^{163,164}.

Alterations and prevalence: Somatic mutations in ENO1 are observed in 3% uterine corpus endometrial carcinoma and kidney chromophobe, and 2% of diffuse large B-cell lymphoma, skin cutaneous melanoma, and cervical squamous cell carcinoma^{10,11}. Amplification of ENO1 is observed in 2% of adrenocortical carcinoma, pancreatic adenocarcinoma, esophageal adenocarcinoma, ovarian serous cystadenocarcinoma, and sarcoma^{10,11}. Biallelic loss of ENO1 is observed in 6% of cholangiocarcinoma, 4% of adrenocortical carcinoma, and 2% of pheochromocytoma and paraganglioma, liver hepatocellular carcinoma, and diffuse large B-cell lymphoma^{10,11}.

Potential relevance: Currently, no therapies are approved for ENO1 aberrations.

PGD deletion

phosphogluconate dehydrogenase

Background: The PGD gene encodes phosphogluconate dehydrogenase, an essential enzyme of the pentose phosphate pathway (PPP) that catalyzes oxidative decarboxylation of 6-phosphogluconate to ribulose-5-phosphate and reduction of NADP⁺ to NADPH^{1,33}. PPP mediated generation of pentose phosphates and NADPH is essential for nucleic acid synthesis and fatty acid synthesis, respectively, making it a crucial metabolic pathway for cancer cell survival and proliferation^{34,35}. Although biallelic deletion appears to be more common than amplification across cancer types, post-translational modifications and overexpression of PGD in cancer have also been observed to result in elevated PPP activity, which is associated with cancer cell proliferation^{33,36}.

Alterations and prevalence: Somatic mutations in PGD have been observed in 4% of skin cutaneous melanoma, 3% of uterine corpus endometrial carcinoma, 2% of diffuse large B-cell lymphoma, stomach adenocarcinoma, and bladder urothelial carcinoma^{10,11}. Biallelic loss of PGD has been observed in 4% of adrenocortical carcinoma, 3% of cholangiocarcinoma, and 2% of pheochromocytoma and paraganglioma and diffuse large B-cell lymphoma^{10,11}. Amplification of PGD has been observed in 2% of esophageal adenocarcinoma, ovarian serous cystadenocarcinoma, stomach adenocarcinoma, and sarcoma^{10,11}.

Potential relevance: Currently, no therapies are approved for PGD aberrations.

MECOM amplification

MDS1 and EVI1 complex locus

Background: The MECOM gene encodes the MDS1 and EVI1 complex locus (MECOM), a zinc-finger transcriptional factor that regulates hematopoietic cell differentiation¹⁴. The MECOM locus encodes multiple alternative splice variants that result in MDS1-EVI1, MDS1, and EVI1 protein isoforms¹⁵. The EVI1 isoform is the most abundant and oncogenic form of MECOM that is expressed in various cancers including acute myeloid leukemia (AML)^{15,16}. MECOM is a frequent target of chromosomal translocation which can lead to MECOM overexpression and leukemogenesis¹⁷.

Alterations and prevalence: Somatic mutations MECOM are observed in up to 22% of malignant melanoma; 75% of these mutations are missense and the remaining 25% are truncating mutations^{10,11,18}. MECOM amplifications are observed in up to 35% of lung squamous cell carcinoma, 30% of ovarian serous cystadenocarcinoma, and 20% of esophageal adenocarcinoma, uterine carcinosarcoma, and cervical squamous cell carcinoma^{10,11}. MECOM rearrangements occur with various partner genes including ETV6, RUNX1, and H2AFY¹⁹. The t(3;21)(q26;q22) translocation that results in the MECOM::RUNX1 fusion is most commonly observed in chronic myeloid

Biomarker Descriptions (continued)

leukemia (CML) in blast crisis. The t(3;3)(q21.3;q26.2)/ inv(3)(q21.3;q26.3) translocation, also referred to as inv(3)/t(3;3), results in a GATA2::MECOM fusion and is observed in AML, primary myelofibrosis (PMF), and myelodysplastic syndrome (MDS)^{20,21,22}. The inv(3)/t(3;3) translocation repositions the distal GATA enhancer element and activates MECOM expression while simultaneously causing GATA2 haploinsufficiency²³.

Potential relevance: AML with MECOM rearrangement is considered a distinct molecular subtype of AML as defined by the World Health Organization (WHO)²⁴. MECOM rearrangements, including GATA2::MECOM fusions, are associated with poor/adverse risk in AML^{20,25}. Inv(3) is associated with poor cytogenetic risk in MDS as defined by the revised international prognostic scoring system (IPSS-R) scoring system²². In PMF, inv(3) is considered an unfavorable karyotype associated with intermediate risk as defined by the dynamic international prognostic scoring system (DIPSS)-Plus scoring system²¹. MECOM overexpression is observed in 10% of de novo AML associated with poor prognosis, and is commonly found in MLL-rearranged cases^{26,27}. Amplification of MECOM is associated with favorable prognosis in ovarian cancer²⁸.

FAT1 p.(S1917Lfs*38) c.5744_5747dup

FAT atypical cadherin 1

Background: FAT1 encodes the FAT atypical cadherin 1 protein, a member of the cadherin superfamily characterized by the presence of cadherin-type repeats^{1,13}. FAT cadherins, which also include FAT2, FAT3, and FAT4, are transmembrane proteins containing a cytoplasmic domain and a number of extracellular laminin G-like motifs and EGF-like motifs, which contributes to their individual functions¹³. The cytoplasmic tail of FAT1 is known to interact with a number of protein targets involved in cell adhesion, proliferation, migration, and invasion¹³. FAT1 has been observed to influence the regulation of several oncogenic pathways, including the WNT/β-catenin, Hippo, and MAPK/ERK signaling pathways, as well as epithelial to mesenchymal transition¹³. Alterations of FAT1 lead to down-regulation or loss of function, supporting a tumor suppressor role for FAT1¹³.

Alterations and prevalence: Somatic mutations in FAT1 are predominantly truncating although, the R1627Q mutation has been identified as a recurrent hotspot^{10,11}. Mutations in FAT1 are observed in 22% of head and neck squamous cell carcinoma, 20% of uterine corpus endometrial carcinoma, 14% of lung squamous cell carcinoma and skin cutaneous melanoma, and 12% diffuse large b-cell lymphoma and bladder urothelial carcinoma^{10,11}. Biallelic loss of FAT1 is observed in 7% of head and neck squamous cell carcinoma, 6% of lung squamous cell carcinoma, 5% of esophageal adenocarcinoma, and 4% of diffuse large b-cell lymphoma, stomach adenocarcinoma and uterine carcinosarcoma^{10,11}.

Potential relevance: Currently, no therapies are approved for FAT1 aberrations.

HLA-A p.(L180*) c.539T>A

major histocompatibility complex, class I, A

Background: The HLA-A gene encodes the major histocompatibility complex, class I, A¹. MHC (major histocompatibility complex) class I molecules are located on the cell surface of nucleated cells and present antigens from within the cell for recognition by cytotoxic T cells⁶³. MHC class I molecules are heterodimers composed of two polypeptide chains, α and B2M⁶⁴. The classical MHC class I genes include HLA-A, HLA-B, and HLA-C and encode the α polypeptide chains, which present short polypeptide chains, of 7 to 11 amino acids, to the immune system to distinguish self from non-self^{65,66,67}. Downregulation of MHC class I promotes tumor evasion of the immune system, suggesting a tumor suppressor role for HLA-A⁶⁸.

Alterations and prevalence: Somatic mutations in HLA-A are observed in 7% of diffuse large B-cell lymphoma (DLBCL), 4% of cervical squamous cell carcinoma and head and neck squamous cell carcinoma, 3% of colorectal adenocarcinoma, and 2% of uterine corpus endometrial carcinoma and stomach adenocarcinoma^{10,11}. Biallelic loss of HLA-A is observed in 4% of DLBCL^{10,11}.

Potential relevance: Currently, no therapies are approved for HLA-A aberrations.

MAP3K4 p.(A1195Gfs*16) c.3584_3585delCT

mitogen-activated protein kinase kinase kinase 4

Background: The MAP3K4 gene encodes the mitogen-activated protein kinase kinase kinase 4, also known as MEKK4¹. MAP3K4 is involved in the JNK signaling pathway along with MAP3K12, MAP2K4, MAP2K7, MAPK8, MAPK9, and MAPK10²⁹. Activation of MAPK proteins occurs through a kinase signaling cascade^{29,30,31}. Specifically, MAP3Ks are responsible for phosphorylation of MAP2K family members^{29,30,31}. Once activated, MAP2Ks are responsible for the phosphorylation of various MAPK proteins whose signaling is involved in several cellular processes including cell proliferation, differentiation, and inflammation^{29,30,31}. In intrahepatic cholangiocarcinoma, mutations leading to lack of MAP3K4 activity result in vascular invasion and poor survival, supporting a tumor suppressor role for MAP3K4³².

Biomarker Descriptions (continued)

Alterations and prevalence: Somatic mutations in MAP3K4 are observed in 10% of uterine corpus endometrial carcinoma, 9% of skin cutaneous melanoma, 7% of uterine carcinosarcoma, and 6% of colorectal adenocarcinoma^{10,11}. Biallelic deletions are observed in 6% of uveal melanoma, 3% of ovarian serous cystadenocarcinoma, and 2% of diffuse large B-cell lymphoma (DLBCL)^{10,11}.

Potential relevance: Currently, no therapies are approved for MAP3K4 aberrations.

Alerts Informed By Public Data Sources

Current FDA Information

 Contraindicated  Not recommended  Resistance  Breakthrough  Fast Track

FDA information is current as of 2025-11-25. For the most up-to-date information, search www.fda.gov.

ERBB2 amplification

trastuzumab pamirtecan

Cancer type: Endometrial Carcinoma

Variant class: ERBB2 overexpression

Supporting Statement:

The FDA has granted Breakthrough Therapy designation to antibody-drug conjugate, trastuzumab pamirtecan (DB-1303), for the treatment of patients with HER2-expressing advanced endometrial cancer.

Reference:

<https://investors.biontech.de//news-releases/news-release-details/biontech-and-dualitybio-receive-fda-breakthrough-therapy>

disitamab vedotinaide

Cancer type: Bladder Urothelial Carcinoma

Variant class: ERBB2 positive

Supporting Statement:

The FDA has granted Breakthrough Therapy designation to the humanized anti-HER2 antibody drug conjugate (ADC), disitamab vedotin, for the second-line treatment of HER2 positive locally advanced or metastatic urothelial cancer (UC) after previous platinum-containing chemotherapy treatment.

Reference:

<https://www.prnewswire.com/news-releases/remegen-announces-us-fda-has-granted-breakthrough-therapy-designation-for-disitamab-vedotin-rc48-in-urothelial-cancer-301138315.html>

zanidatamab + chemotherapy

Cancer type: Gastroesophageal Junction Adenocarcinoma

Variant class: ERBB2 overexpression

Supporting Statement:

The FDA has granted Fast Track designation to the HER2 targeted bispecific antibody, zanidatamab, for HER2-overexpressing gastroesophageal adenocarcinoma (GEA) to be used in combination with standard-of-care chemotherapy.

Reference:

<https://www.targetedonc.com/view/her2targeted-antibody-zw25-earns-fda-fast-track-designation-in-gea>

anvtabart opadotin

Cancer type: Breast Cancer

Variant class: ERBB2 positive

Supporting Statement:

The FDA has granted Fast Track designation to the HER2-targeting antibody drug conjugate, anvtabart opadotin (ARX-788), for HER2-positive metastatic breast cancer.

Reference:

<https://ir.ambrx.com/news/news-details/2023/ACE-Breast-02-Pivotal-Phase-3-Study-of-Ambrxs-ARX788-for-the-Treatment-of-HER2-Positive-Metastatic-Breast-Cancer-Achieves-Positive-Results/default.aspx>

ERBB2 amplification (continued)

evorpacept

Cancer type: Gastric Cancer,
Gastroesophageal Junction Adenocarcinoma

Variant class: ERBB2 positive

Supporting Statement:

The FDA has granted Fast Track designation to the CD47 checkpoint inhibitor, ALX148, for the second-line treatment of patients with HER2-positive gastric or gastroesophageal junction carcinoma.

Reference:

<https://www.targetedonc.com/view/two-fda-fast-track-designations-granted-to-alx148-for-hnsc-and-gastric-adenocarcinomas>

GLSI-100

Cancer type: Breast Cancer

Variant class: ERBB2 positive


Supporting Statement:

The FDA has granted Fast Track designation to the immunotherapy, GLSI-100, for the treatment of patients with HLA-A*02 genotype and HER2-positive breast cancer who have completed treatment with standard of care HER2/neu targeted therapy to improve invasive breast cancer free survival.

Reference:


<https://investor.greenwichlifesciences.com/news-events/press-releases/detail/102/us-fda-fast-track-designation>


Current ESMO Information

 Contraindicated

 Not recommended

 Resistance

 Breakthrough

 Fast Track

ESMO information is current as of 2025-11-03. For the most up-to-date information, search www.esmo.org.

ERBB2 amplification

trastuzumab

Cancer type: Gastric Cancer

Variant class: ERBB2 overexpression

Summary:

ESMO Clinical Practice Guidelines include the following supporting statement:

- "Treatment with trastuzumab is not recommended after first-line therapy in HER2-positive advanced gastric cancer [I, D]."

Reference: ESMO Clinical Practice Guidelines - ESMO-Gastric Cancer [Ann Oncol (2022), doi: <https://doi.org/10.1016/j.annonc.2022.07.004>]

ERBB2 amplification (continued)

— hormone therapy

Cancer type: Breast Cancer

Variant class: ERBB2 positive

Other criteria: Hormone receptor positive

ESMO Level of Evidence/Grade of Recommendation: III / C

Summary:

ESMO™ Clinical Practice Guidelines include the following supporting statement:

- "The use of single-agent ET without a HER2-targeted therapy is not routinely recommended unless cardiac disease precludes the safe use of HER2-directed therapies [III, C]"

Reference: ESMO Clinical Practice Guidelines - ESMO-Metastatic Breast Cancer [Ann Oncol (2021) VOLUME 32, ISSUE 12, P1475-1495, DECEMBER 01, 2021; DOI:<https://doi.org/10.1016/j.annonc.2021.09.019>]

Genes Assayed

Genes Assayed for the Detection of DNA Sequence Variants

ABL1, ABL2, ACVR1, AKT1, AKT2, AKT3, ALK, AR, ARAF, ATP1A1, AURKA, AURKB, AURKC, AXL, BCL2, BCL2L12, BCL6, BCR, BMP5, BRAF, BTK, CACNA1D, CARD11, CBL, CCND1, CCND2, CCND3, CCNE1, CD79B, CDK4, CDK6, CHD4, CSF1R, CTNNB1, CUL1, CYSLTR2, DDR2, DGCR8, DROSHA, E2F1, EGFR, EIF1AX, EPAS1, ERBB2, ERBB3, ERBB4, ESR1, EZH2, FAM135B, FGF7, FGFR1, FGFR2, FGFR3, FGFR4, FLT3, FLT4, FOXA1, FOXL2, FOXO1, GATA2, GLI1, GNA11, GNAQ, GNAS, HIF1A, HRAS, IDH1, IDH2, IKBKB, IL6ST, IL7R, IRF4, IRS4, KCNJ5, KDR, KIT, KLF4, KLF5, KNSTRN, KRAS, MAGOH, MAP2K1, MAP2K2, MAPK1, MAX, MDM4, MECOM, MED12, MEF2B, MET, MITF, MPL, MTOR, MYC, MYCN, MYD88, MYOD1, NFE2L2, NRAS, NSD2, NT5C2, NTRK1, NTRK2, NTRK3, NUP93, PAX5, PCBP1, PDGFRA, PDGFRB, PIK3C2B, PIK3CA, PIK3CB, PIK3CD, PIK3CG, PIK3R2, PIM1, PLCG1, PPP2R1A, PPP6C, PRKACA, PTPN11, PTPRD, PXDNL, RAC1, RAF1, RARA, RET, RGS7, RHEB, RHOA, RICTOR, RIT1, ROS1, RPL10, SETBP1, SF3B1, SIX1, SIX2, SLC01B3, SMC1A, SMO, SNCAIP, SOS1, SOX2, SPOP, SRC, SRSF2, STAT3, STAT5B, STAT6, TAF1, TERT, TGFB1, TOP1, TOP2A, TPMT, TRRAP, TSHR, U2AF1, USP8, WAS, XPO1, ZNF217, ZNF429

Genes Assayed for the Detection of Copy Number Variations

ABCB1, ABL1, ABL2, ABRAXAS1, ACVR1B, ACVR2A, ADAMTS12, ADAMTS2, AKT1, AKT2, AKT3, ALK, AMER1, APC, AR, ARAF, ARHGAP35, ARID1A, ARID1B, ARID2, ARID5B, ASXL1, ASXL2, ATM, ATR, ATRX, AURKA, AURKC, AXIN1, AXIN2, AXL, B2M, BAP1, BARD1, BCL2, BCL2L12, BCL6, BCOR, BLM, BMPR2, BRAF, BRCA1, BRCA2, BRIP1, CARD11, CASP8, CBFB, CBL, CCND1, CCND2, CCND3, CCNE1, CD274, CD276, CDC73, CDH1, CDH10, CDK12, CDK4, CDK6, CDKN1A, CDKN1B, CDKN2A, CDKN2B, CDKN2C, CHD4, CHEK1, CHEK2, CIC, CREBBP, CSMD3, CTCF, CTLA4, CTNND2, CUL3, CUL4A, CUL4B, CYLD, CYP2C9, DAXX, DDR1, DDR2, DDX3X, DICER1, DNMT3A, DOCK3, DPYD, DSC1, DSC3, EGFR, EIF1AX, ELF3, EMSY, ENO1, EP300, EPCAM, EPHA2, ERAP1, ERAP2, ERBB2, ERBB3, ERBB4, ERCC2, ERCC4, ERRFI1, ESR1, ETV6, EZH2, FAM135B, FANCA, FANCC, FANCD2, FANCE, FANCF, FANCG, FANCI, FANCL, FANCM, FAT1, FBXW7, FGF19, FGF23, FGF3, FGF4, FGF9, FGFR1, FGFR2, FGFR3, FGFR4, FLT3, FLT4, FOXA1, FUBP1, FYN, GATA2, GATA3, GLI3, GNA13, GNAS, GPS2, HDAC2, HDAC9, HLA-A, HLA-B, HNF1A, IDH2, IGF1R, IKBKB, IL7R, INPP4B, JAK1, JAK2, JAK3, KDM5C, KDM6A, KDR, KEAP1, KIT, KLF5, KMT2A, KMT2B, KMT2C, KMT2D, KRAS, LARP4B, LATS1, LATS2, MAGOH, MAP2K1, MAP2K4, MAP2K7, MAP3K1, MAP3K4, MAPK1, MAPK8, MAX, MCL1, MDM2, MDM4, MECOM, MEF2B, MEN1, MET, MGA, MITF, MLH1, MLH3, MPL, MRE11, MSH2, MSH3, MSH6, MTAP, MTOR, MUTHYH, MYC, MYCL, MYCN, MYD88, NBN, NCOR1, NF1, NF2, NFE2L2, NOTCH1, NOTCH2, NOTCH3, NOTCH4, NRAS, NTRK1, NTRK3, PALB2, PARP1, PARP2, PARP3, PARP4, PBRM1, PCBP1, PDCD1, PDCD1LG2, PDGFRA, PDGFRB, PDIA3, PGD, PHF6, PIK3C2B, PIK3CA, PIK3CB, PIK3R1, PIK3R2, PIM1, PLCG1, PMS1, PMS2, POLD1, POLE, POT1, PPM1D, PPP2R1A, PPP2R2A, PPP6C, PRDM1, PRDM9, PRKACA, PRKAR1A, PTCH1, PTEN, PTPN11, PTPRT, PXDNL, RAC1, RAD50, RAD51, RAD51B, RAD51C, RAD51D, RAD52, RAD54L, RAF1, RARA, RASA1, RASA2, RB1, RBM10, RECQL4, RET, RHEB, RICTOR, RIT1, RNASEH2A, RNASEH2B, RNF43, ROS1, RPA1, RPS6KB1, RPTOR, RUNX1, SDHA, SDHB, SDHD, SETBP1, SETD2, SF3B1, SLC01B3, SLX4, SMAD2, SMAD4, SMARCA4, SMARCB1, SMC1A, SMO, SOX9, SPEN, SPOP, SRC, STAG2, STAT3, STAT6, STK11, SUFU, TAP1, TAP2, TBX3, TCF7L2, TERT, TET2, TGFB1, TNFAIP3, TNFRSF14, TOP1, TP53, TP63, TPMT, TPP2, TSC1, TSC2, U2AF1, USP8, USP9X, VHL, WT1, XPO1, XRCC2, XRCC3, YAP1, YES1, ZFH3, ZMYM3, ZNF217, ZNF429, ZRSR2

Genes Assayed (continued)

Genes Assayed for the Detection of Fusions

AKT2, ALK, AR, AXL, BRAF, BRCA1, BRCA2, CDKN2A, EGFR, ERBB2, ERBB4, ERG, ESR1, ETV1, ETV4, ETV5, FGFR1, FGFR2, FGFR3, FGR, FLT3, JAK2, KRAS, MDM4, MET, MYB, MYBL1, NF1, NOTCH1, NOTCH4, NRG1, NTRK1, NTRK2, NTRK3, NUTM1, PDGFRA, PDGFRB, PIK3CA, PPARG, PRKACA, PRKACB, PTEN, RAD51B, RAF1, RB1, RELA, RET, ROS1, RSP02, RSP03, TERT

Genes Assayed with Full Exon Coverage

ABRAXAS1, ACVR1B, ACVR2A, ADAMTS12, ADAMTS2, AMER1, APC, ARHGAP35, ARID1A, ARID1B, ARID2, ARID5B, ASXL1, ASXL2, ATM, ATR, ATRX, AXIN1, AXIN2, B2M, BAP1, BARD1, BCOR, BLM, BMPR2, BRCA1, BRCA2, BRIP1, CALR, CASP8, CBFB, CD274, CD276, CDC73, CDH1, CDH10, CDK12, CDKN1A, CDKN1B, CDKN2A, CDKN2B, CDKN2C, CHEK1, CHEK2, CIC, CIITA, CREBBP, CSMD3, CTCF, CTLA4, CUL3, CUL4A, CUL4B, CYLD, CYP2C9, CYP2D6, DAXX, DDX3X, DICER1, DNMT3A, DOCK3, DPYD, DSC1, DSC3, ELF3, ENO1, EP300, EPCAM, EPHA2, ERAP1, ERAP2, ERCC2, ERCC4, ERCC5, ERRF1, ETV6, FANCA, FANCC, FANCD2, FANCE, FANCF, FANCG, FANCI, FANCL, FANCM, FAS, FAT1, FBXW7, FUBP1, GATA3, GNA13, GPS2, HDAC2, HDAC9, HLA-A, HLA-B, HNF1A, ID3, INPP4B, JAK1, JAK2, JAK3, KDM5C, KDM6A, KEAP1, KLHL13, KMT2A, KMT2B, KMT2C, KMT2D, LARP4B, LATS1, LATS2, MAP2K4, MAP2K7, MAP3K1, MAP3K4, MAPK8, MEN1, MGA, MLH1, MLH3, MRE11, MSH2, MSH3, MSH6, MTAP, MTUS2, MUTYH, NBN, NCOR1, NF1, NF2, NOTCH1, NOTCH2, NOTCH3, NOTCH4, PALB2, PARP1, PARP2, PARP3, PARP4, PBRM1, PDCD1, PDCD1LG2, PDIA3, PGD, PHF6, PIK3R1, PMS1, PMS2, POLD1, POLE, POT1, PPM1D, PPP2R2A, PRDM1, PRDM9, PRKAR1A, PSMB10, PSMB8, PSMB9, PTCH1, PTEN, PTPRT, RAD50, RAD51, RAD51B, RAD51C, RAD51D, RAD52, RAD54L, RASA1, RASA2, RB1, RBM10, RECQL4, RNASEH2A, RNASEH2B, RNASEH2C, RNF43, RPA1, RPL22, RPL5, RUNX1, RUNX1T1, SDHA, SDHB, SDHC, SDHD, SETD2, SLX4, SMAD2, SMAD4, SMARCA4, SMARCB1, SOCS1, SOX9, SPEN, STAG2, STAT1, STK11, SUFU, TAP1, TAP2, TBX3, TCF7L2, TET2, TGFB2, TMEM132D, TNFAIP3, TNFRSF14, TP53, TP63, TPP2, TSC1, TSC2, UGT1A1, USP9X, VHL, WT1, XRCC2, XRCC3, ZBTB20, ZFH3, ZMYM3, ZRSR2

Relevant Therapy Summary

☒ In this cancer type ☐ In other cancer type ☒ In this cancer type and other cancer types ☒ No evidence

ERBB2 amplification

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
trastuzumab deruxtecan	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/> (II)
zanidatamab	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/> (II)
ado-trastuzumab emtansine	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
lapatinib + capecitabine	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
neratinib	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
pertuzumab + trastuzumab + chemotherapy	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
pertuzumab + trastuzumab + docetaxel	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
trastuzumab + docetaxel	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
trastuzumab + paclitaxel	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
trastuzumab + tucatinib + capecitabine	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>
trastuzumab	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>	<input checked="" type="radio"/> (II)
trastuzumab + capecitabine + cisplatin	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>	<input checked="" type="radio"/>
trastuzumab + carboplatin + docetaxel	<input type="radio"/>	<input type="radio"/>	<input type="radio"/>	<input checked="" type="radio"/>	<input checked="" type="radio"/>

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

● In this cancer type
 ○ In other cancer type
 ⓘ In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
trastuzumab + cisplatin + fluorouracil	○	○	○	✕	✕
neratinib + capecitabine	○	○	✕	✕	✕
trastuzumab + tucatinib	○	○	✕	✕	✕
lapatinib + letrozole	○	✕	○	✕	✕
pembrolizumab + trastuzumab + chemotherapy + fluoropyrimidine	○	✕	○	✕	✕
pertuzumab/trastuzumab/hyaluronidase-zzxf + cyclophosphamide + doxorubicin	○	✕	○	✕	✕
pertuzumab/trastuzumab/hyaluronidase-zzxf + docetaxel	○	✕	○	✕	✕
trastuzumab (Biocon)	○	✕	○	✕	✕
trastuzumab (Biocon) + capecitabine + cisplatin	○	✕	○	✕	✕
trastuzumab (Biocon) + carboplatin + docetaxel	○	✕	○	✕	✕
trastuzumab (Biocon) + cisplatin + fluorouracil	○	✕	○	✕	✕
trastuzumab (Biocon) + docetaxel	○	✕	○	✕	✕
trastuzumab (Biocon) + paclitaxel	○	✕	○	✕	✕
trastuzumab (Celltrion)	○	✕	○	✕	✕
trastuzumab (Celltrion) + capecitabine + cisplatin	○	✕	○	✕	✕
trastuzumab (Celltrion) + carboplatin + docetaxel	○	✕	○	✕	✕
trastuzumab (Celltrion) + cisplatin + fluorouracil	○	✕	○	✕	✕
trastuzumab (Celltrion) + docetaxel	○	✕	○	✕	✕
trastuzumab (Celltrion) + paclitaxel	○	✕	○	✕	✕
trastuzumab (Henlius)	○	✕	○	✕	✕
trastuzumab (Pfizer)	○	✕	○	✕	✕
trastuzumab (Pfizer) + capecitabine + cisplatin	○	✕	○	✕	✕
trastuzumab (Pfizer) + carboplatin + docetaxel	○	✕	○	✕	✕
trastuzumab (Pfizer) + cisplatin + fluorouracil	○	✕	○	✕	✕
trastuzumab (Pfizer) + docetaxel	○	✕	○	✕	✕
trastuzumab (Pfizer) + paclitaxel	○	✕	○	✕	✕
trastuzumab (Samsung Bioepis)	○	✕	○	✕	✕

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

● In this cancer type
 ○ In other cancer type
 ① In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
trastuzumab (Samsung Bioepis) + capecitabine + cisplatin	○	✕	○	✕	✕
trastuzumab (Samsung Bioepis) + carboplatin + docetaxel	○	✕	○	✕	✕
trastuzumab (Samsung Bioepis) + cisplatin + fluorouracil	○	✕	○	✕	✕
trastuzumab (Samsung Bioepis) + docetaxel	○	✕	○	✕	✕
trastuzumab (Samsung Bioepis) + paclitaxel	○	✕	○	✕	✕
trastuzumab (Synthon)	○	✕	○	✕	✕
trastuzumab (Synthon) + capecitabine + cisplatin	○	✕	○	✕	✕
trastuzumab (Synthon) + carboplatin + docetaxel	○	✕	○	✕	✕
trastuzumab (Synthon) + cisplatin + fluorouracil	○	✕	○	✕	✕
trastuzumab (Synthon) + docetaxel	○	✕	○	✕	✕
trastuzumab (Synthon) + paclitaxel	○	✕	○	✕	✕
margetuximab + chemotherapy	○	✕	✕	○	✕
pembrolizumab + berahyaluronidase alfa + trastuzumab + chemotherapy	○	✕	✕	✕	✕
trastuzumab and hyaluronidase-oysk	○	✕	✕	✕	✕
trastuzumab and hyaluronidase-oysk + carboplatin + docetaxel	○	✕	✕	✕	✕
trastuzumab and hyaluronidase-oysk + docetaxel	○	✕	✕	✕	✕
trastuzumab and hyaluronidase-oysk + paclitaxel	○	✕	✕	✕	✕
lapatinib + trastuzumab	✕	○	○	○	✕
pertuzumab + trastuzumab	✕	○	✕	○	● (II/III)
pertuzumab + trastuzumab + hormone therapy	✕	○	✕	○	✕
pertuzumab + trastuzumab + paclitaxel	✕	○	✕	○	✕
trastuzumab + chemotherapy	✕	○	✕	○	✕
trastuzumab + hormone therapy	✕	○	✕	○	✕
abemaciclib + trastuzumab + fulvestrant	✕	○	✕	✕	✕
ado-trastuzumab emtansine + neratinib	✕	○	✕	✕	✕
aromatase inhibitor	✕	○	✕	✕	✕

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

☒ In this cancer type
 ☐ In other cancer type
 ☒ In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
fulvestrant	✕	○	✕	✕	✕
hormone therapy	✕	○	✕	✕	✕
lapatinib + aromatase inhibitor	✕	○	✕	✕	✕
lapatinib + trastuzumab + aromatase inhibitor	✕	○	✕	✕	✕
margetuximab + capecitabine	✕	○	✕	✕	✕
margetuximab + eribulin	✕	○	✕	✕	✕
margetuximab + gemcitabine	✕	○	✕	✕	✕
margetuximab + vinorelbine	✕	○	✕	✕	✕
neratinib + paclitaxel	✕	○	✕	✕	✕
pembrolizumab + trastuzumab + capecitabine + cisplatin	✕	○	✕	✕	✕
pembrolizumab + trastuzumab + capecitabine + oxaliplatin	✕	○	✕	✕	✕
pembrolizumab + trastuzumab + cisplatin + fluorouracil	✕	○	✕	✕	✕
pembrolizumab + trastuzumab + fluorouracil + oxaliplatin	✕	○	✕	✕	✕
pertuzumab + trastuzumab + carboplatin + docetaxel	✕	○	✕	✕	✕
pertuzumab + trastuzumab + carboplatin + paclitaxel	✕	○	✕	✕	✕
pertuzumab + trastuzumab + hormone therapy + chemotherapy	✕	○	✕	✕	✕
tamoxifen	✕	○	✕	✕	✕
trastuzumab + aromatase inhibitor	✕	○	✕	✕	✕
trastuzumab + capecitabine	✕	○	✕	✕	✕
trastuzumab + capecitabine + oxaliplatin	✕	○	✕	✕	✕
trastuzumab + carboplatin + paclitaxel	✕	○	✕	✕	✕
trastuzumab + chemotherapy (non-anthracycline)	✕	○	✕	✕	✕
trastuzumab + cisplatin + docetaxel	✕	○	✕	✕	✕
trastuzumab + cisplatin + docetaxel + fluorouracil	✕	○	✕	✕	✕
trastuzumab + cisplatin + paclitaxel	✕	○	✕	✕	✕
trastuzumab + cyclophosphamide + docetaxel	✕	○	✕	✕	✕

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

● In this cancer type
 ○ In other cancer type
 ⓘ In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
trastuzumab + docetaxel + fluorouracil + oxaliplatin	✕	○	✕	✕	✕
trastuzumab + fluorouracil	✕	○	✕	✕	✕
trastuzumab + fluorouracil + irinotecan	✕	○	✕	✕	✕
trastuzumab + fluorouracil + oxaliplatin	✕	○	✕	✕	✕
trastuzumab + fulvestrant	✕	○	✕	✕	✕
trastuzumab + hormone therapy + chemotherapy	✕	○	✕	✕	✕
trastuzumab + tamoxifen	✕	○	✕	✕	✕
trastuzumab + vinorelbine	✕	○	✕	✕	✕
pertuzumab/trastuzumab/hyaluronidase-zzxf + carboplatin + docetaxel	✕	✕	○	✕	✕
pertuzumab/trastuzumab/hyaluronidase-zzxf + cyclophosphamide + doxorubicin + fluorouracil	✕	✕	○	✕	✕
pertuzumab/trastuzumab/hyaluronidase-zzxf + cyclophosphamide + epirubicin	✕	✕	○	✕	✕
pertuzumab/trastuzumab/hyaluronidase-zzxf + paclitaxel	✕	✕	○	✕	✕
trastuzumab (Biocon) + anastrozole	✕	✕	○	✕	✕
trastuzumab (Celltrion) + anastrozole	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics)	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics) + anastrozole	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics) + capecitabine + cisplatin	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics) + carboplatin + docetaxel	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics) + cisplatin + fluorouracil	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics) + docetaxel	✕	✕	○	✕	✕
trastuzumab (CuraTeQ Biologics) + paclitaxel	✕	✕	○	✕	✕
trastuzumab (EirGenix)	✕	✕	○	✕	✕
trastuzumab (EirGenix) + anastrozole	✕	✕	○	✕	✕
trastuzumab (EirGenix) + capecitabine + cisplatin	✕	✕	○	✕	✕
trastuzumab (EirGenix) + carboplatin + docetaxel	✕	✕	○	✕	✕

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

● In this cancer type
 ○ In other cancer type
 ① In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
trastuzumab (EirGenix) + cisplatin + fluorouracil	✕	✕	○	✕	✕
trastuzumab (EirGenix) + docetaxel	✕	✕	○	✕	✕
trastuzumab (EirGenix) + paclitaxel	✕	✕	○	✕	✕
trastuzumab (Henlius) + anastrozole	✕	✕	○	✕	✕
trastuzumab (Henlius) + capecitabine + cisplatin	✕	✕	○	✕	✕
trastuzumab (Henlius) + carboplatin + docetaxel	✕	✕	○	✕	✕
trastuzumab (Henlius) + cisplatin + fluorouracil	✕	✕	○	✕	✕
trastuzumab (Henlius) + docetaxel	✕	✕	○	✕	✕
trastuzumab (Henlius) + paclitaxel	✕	✕	○	✕	✕
trastuzumab (Pfizer) + anastrozole	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma)	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma) + anastrozole	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma) + capecitabine + cisplatin	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma) + carboplatin + docetaxel	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma) + cisplatin + fluorouracil	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma) + docetaxel	✕	✕	○	✕	✕
trastuzumab (Prestige BioPharma) + paclitaxel	✕	✕	○	✕	✕
trastuzumab (Samsung Bioepis) + anastrozole	✕	✕	○	✕	✕
trastuzumab (Synthon) + anastrozole	✕	✕	○	✕	✕
trastuzumab + anastrozole	✕	✕	○	✕	✕
ado-trastuzumab emtansine + hormone therapy	✕	✕	✕	○	✕
lapatinib + hormone therapy	✕	✕	✕	○	✕
lapatinib + trastuzumab + hormone therapy	✕	✕	✕	○	✕
margetuximab	✕	✕	✕	○	✕
neratinib + chemotherapy	✕	✕	✕	○	✕
pertuzumab + trastuzumab + nab-paclitaxel	✕	✕	✕	○	✕
pyrotinib	✕	✕	✕	✕	● (IV)
IAH-0968, chemotherapy	✕	✕	✕	✕	● (III)

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

☒ In this cancer type
 ☐ In other cancer type
 ☒ In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
trastuzumab deruxtecan, pembrolizumab, chemotherapy	✕	✕	✕	✕	● (III)
allitinib	✕	✕	✕	✕	● (II)
CART-HER2, chemotherapy	✕	✕	✕	✕	● (II)
disitamab vedotinaide, tislelizumab, bevacizumab	✕	✕	✕	✕	● (II)
FDA022-BB05	✕	✕	✕	✕	● (II)
neratinib, neratinib + palbociclib	✕	✕	✕	✕	● (II)
pertuzumab + trastuzumab, atezolizumab + pertuzumab/trastuzumab/hyaluronidase-zzxf, trastuzumab + tucatinib	✕	✕	✕	✕	● (II)
pyrotinib, chemotherapy	✕	✕	✕	✕	● (II)
trastuzumab (Samsung Bioepis), chemotherapy	✕	✕	✕	✕	● (II)
tucatinib, ado-trastuzumab emtansine	✕	✕	✕	✕	● (II)
tucatinib, trastuzumab	✕	✕	✕	✕	● (II)
zongertinib	✕	✕	✕	✕	● (II)
AP-402	✕	✕	✕	✕	● (I/II)
AZD-9574, trastuzumab deruxtecan	✕	✕	✕	✕	● (I/II)
BAT-8010, BAT-1006	✕	✕	✕	✕	● (I/II)
BL-M07D1	✕	✕	✕	✕	● (I/II)
DF-1001, nivolumab	✕	✕	✕	✕	● (I/II)
disitamab vedotinaide, catequentinib	✕	✕	✕	✕	● (I/II)
E01001	✕	✕	✕	✕	● (I/II)
HypoSti.CART-HER2, chemotherapy	✕	✕	✕	✕	● (I/II)
IAH-0968	✕	✕	✕	✕	● (I/II)
IBI-354	✕	✕	✕	✕	● (I/II)
JIN-A-04	✕	✕	✕	✕	● (I/II)
ST-1703	✕	✕	✕	✕	● (I/II)
trastuzumab deruxtecan, neratinib	✕	✕	✕	✕	● (I/II)
trastuzumab pamirtecan, pertuzumab	✕	✕	✕	✕	● (I/II)
YH32367	✕	✕	✕	✕	● (I/II)
ZV-0203	✕	✕	✕	✕	● (I/II)

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

● In this cancer type
 ○ In other cancer type
 ● In this cancer type and other cancer types
 ✕ No evidence

ERBB2 amplification (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
177Lu-RAD202	✕	✕	✕	✕	● (I)
ado-trastuzumab emtansine (Shanghai Fosun Pharma)	✕	✕	✕	✕	● (I)
anti-HER-2 MAb (Anke Biotechnology)	✕	✕	✕	✕	● (I)
BC004	✕	✕	✕	✕	● (I)
BL-M17D1	✕	✕	✕	✕	● (I)
BM-230	✕	✕	✕	✕	● (I)
CART-HER2	✕	✕	✕	✕	● (I)
CART-HER2/PD-L1	✕	✕	✕	✕	● (I)
ceralasertib, trastuzumab deruxtecan	✕	✕	✕	✕	● (I)
D3L-001	✕	✕	✕	✕	● (I)
doxorubicin (Hangzhou HighField Biopharma)	✕	✕	✕	✕	● (I)
DX126-262	✕	✕	✕	✕	● (I)
ENT-H-1, trastuzumab	✕	✕	✕	✕	● (I)
GQ-1005	✕	✕	✕	✕	● (I)
GQ1001	✕	✕	✕	✕	● (I)
HF-50	✕	✕	✕	✕	● (I)
KJ-015	✕	✕	✕	✕	● (I)
MBS301	✕	✕	✕	✕	● (I)
NC-18	✕	✕	✕	✕	● (I)
NVL-330	✕	✕	✕	✕	● (I)
SPH5030	✕	✕	✕	✕	● (I)
TAS0728	✕	✕	✕	✕	● (I)
TL-938	✕	✕	✕	✕	● (I)
trastuzumab deruxtecan, azenosertib	✕	✕	✕	✕	● (I)
trastuzumab deruxtecan, durvalumab, chemotherapy, volrustomig	✕	✕	✕	✕	● (I)
VRN-10	✕	✕	✕	✕	● (I)
VVD-159642	✕	✕	✕	✕	● (I)
XMT-2056	✕	✕	✕	✕	● (I)

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

Relevant Therapy Summary (continued)

In this cancer type

In other cancer type

In this cancer type and other cancer types

No evidence

ATM deletion

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
olaparib	×	×	×	×	● (II)
pamiparib, tislelizumab	×	×	×	×	● (II)
senaparib, IMP-9064	×	×	×	×	● (I/II)

PIK3CA amplification

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
palbociclib, gedatolisib	×	×	×	×	● (I)
TOS-358	×	×	×	×	● (I)

CHEK1 deletion

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
pamiparib, tislelizumab	×	×	×	×	● (II)

FANCM p.(Q1108*) c.3322C>T

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
talazoparib	×	×	×	×	● (II)

* Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

HRR Details

Gene/Genomic Alteration	Finding
LOH percentage	39.54%
BRCA2	LOH, 13q13.1(32890491-32972932)x3
ATM	CNV, CN:1.0
ATM	LOH, 11q22.3(108098341-108236285)x1
BARD1	LOH, 2q35(215593375-215674382)x2
CHEK1	CNV, CN:1.0
CHEK1	LOH, 11q24.2(125496639-125525271)x1
RAD51B	LOH, 14q24.1(68290164-69061406)x4

Homologous recombination repair (HRR) genes were defined from published evidence in relevant therapies, clinical guidelines, as well as clinical trials, and include - BRCA1, BRCA2, ATM, BARD1, BRIP1, CDK12, CHEK1, CHEK2, FANCL, PALB2, RAD51B, RAD51C, RAD51D, and RAD54L.

Thermo Fisher Scientific's Ion Torrent OncoPrint Reporter software was used in generation of this report. Software was developed and designed internally by Thermo Fisher Scientific. The analysis was based on OncoPrint Reporter (6.2.4 data version 2025.12(007)). The data presented here are from a curated knowledge base of publicly available information, but may not be exhaustive. FDA information was sourced from www.fda.gov and is current as of 2025-11-25. NCCN information was sourced from www.nccn.org and is current as of 2025-11-03. EMA information was sourced from www.ema.europa.eu and is current as of 2025-11-25. ESMO information was sourced from www.esmo.org and is current as of 2025-11-03. Clinical Trials information is current as of 2025-11-03. For the most up-to-date information regarding a particular trial, search www.clinicaltrials.gov by NCT ID or search local clinical trials authority website by local identifier listed in 'Other identifiers.' Variants are reported according to HGVS nomenclature and classified following AMP/ASCO/CAP guidelines (Li et al. 2017). Based on the data sources selected, variants, therapies, and trials listed in this report are listed in order of potential clinical significance but not for predicted efficacy of the therapies.

References

1. O'Leary et al. Reference sequence (RefSeq) database at NCBI: current status, taxonomic expansion, and functional annotation. *Nucleic Acids Res.* 2016 Jan 4;44(D1):D733-45. PMID: 26553804
2. Niraj et al. The Fanconi Anemia Pathway in Cancer. *Annu Rev Cancer Biol.* 2019 Mar;3:457-478. PMID: 30882047
3. Rodríguez et al. Fanconi anemia pathway. *Curr Biol.* 2017 Sep 25;27(18):R986-R988. PMID: 28950089
4. Garcia-Higuera et al. Interaction of the Fanconi anemia proteins and BRCA1 in a common pathway. *Mol. Cell.* 2001 Feb;7(2):249-62. PMID: 11239454
5. Hussain et al. Direct interaction of FANCD2 with BRCA2 in DNA damage response pathways. *Hum. Mol. Genet.* 2004 Jun 15;13(12):1241-8. PMID: 15115758
6. Lord et al. BRCAness revisited. *Nat. Rev. Cancer.* 2016 Feb;16(2):110-20. PMID: 26775620
7. Byrum et al. Defining and Modulating 'BRCAness'. *Trends Cell Biol.* 2019 Sep;29(9):740-751. PMID: 31362850
8. Michl et al. Interplay between Fanconi anemia and homologous recombination pathways in genome integrity. *EMBO J.* 2016 May 2;35(9):909-23. PMID: 27037238
9. Abbasi et al. A rare FANCA gene variation as a breast cancer susceptibility allele in an Iranian population. *Mol Med Rep.* 2017 Jun;15(6):3983-3988. PMID: 28440412
10. Weinstein et al. The Cancer Genome Atlas Pan-Cancer analysis project. *Nat. Genet.* 2013 Oct;45(10):1113-20. PMID: 24071849
11. Cerami et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. *Cancer Discov.* 2012 May;2(5):401-4. PMID: 22588877
12. Stoepker et al. DNA helicases FANCM and DDX11 are determinants of PARP inhibitor sensitivity. *DNA Repair (Amst).* 2015 Feb;26:54-64. PMID: 25583207
13. Peng et al. Role of FAT1 in health and disease. *Oncol Lett.* 2021 May;21(5):398. PMID: 33777221
14. Hinai et al. Review: Aberrant EVI1 expression in acute myeloid leukaemia. *Br. J. Haematol.* 2016 Mar;172(6):870-8. PMID: 26729571
15. Bard-Chapeau et al. EVI1 oncoprotein interacts with a large and complex network of proteins and integrates signals through protein phosphorylation. *Proc. Natl. Acad. Sci. U.S.A.* 2013 Jul 30;110(31):E2885-94. PMID: 23858473
16. Ogawa et al. Abnormal expression of Evi-1 gene in human leukemias. *Hum. Cell.* 1996 Dec;9(4):323-32. PMID: 9183665
17. Choi et al. Intratumoral Heterogeneity of Frameshift Mutations in MECOM Gene is Frequent in Colorectal Cancers with High Microsatellite Instability. *Pathol. Oncol. Res.* 2017 Jan;23(1):145-149. PMID: 27620344
18. Lee et al. Targeted next-generation sequencing reveals high frequency of mutations in epigenetic regulators across treatment-naïve patient melanomas. 2015 Jun 9;7:59. PMID: 26221190
19. Han et al. H2AFY is a novel fusion partner of MECOM in acute myeloid leukemia. *Cancer Genet.* 2018 Apr;222-223:9-12. PMID: 29666008
20. NCCN Guidelines® - NCCN-Acute Myeloid Leukemia [Version 2.2026]
21. NCCN Guidelines® - NCCN-Myeloproliferative Neoplasms [Version 2.2025]
22. NCCN Guidelines® - NCCN-Myelodysplastic Syndromes [Version 1.2026]
23. Gröschel et al. A single oncogenic enhancer rearrangement causes concomitant EVI1 and GATA2 deregulation in leukemia. *Cell.* 2014 Apr 10;157(2):369-381. PMID: 24703711
24. Khoury et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Myeloid and Histiocytic/Dendritic Neoplasms. *Leukemia.* 2022 Jul;36(7):1703-1719. PMID: 35732831
25. Döhner et al. Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood.* 2022 Sep 22;140(12):1345-1377. PMID: 35797463
26. Barjesteh van Waalwijk van Doorn-Khosrovani et al. High EVI1 expression predicts poor survival in acute myeloid leukemia: a study of 319 de novo AML patients. *Blood.* 2003 Feb 1;101(3):837-45. PMID: 12393383
27. Stevens et al. EVI1 expression in childhood acute lymphoblastic leukaemia is not restricted to MLL and BCR/ABL rearrangements and is influenced by age. *Blood Cancer J.* 2014 Jan 24;4:e179. PMID: 24464103
28. Nanjundan et al. Amplification of MDS1/EVI1 and EVI1, located in the 3q26.2 amplicon, is associated with favorable patient prognosis in ovarian cancer. *Cancer Res.* 2007 Apr 1;67(7):3074-84. PMID: 17409414
29. Pritchard et al. Molecular pathways: mitogen-activated protein kinase pathway mutations and drug resistance. *Clin. Cancer Res.* 2013 May 1;19(9):2301-9. PMID: 23406774
30. Lee et al. Targeting MAPK Signaling in Cancer: Mechanisms of Drug Resistance and Sensitivity. *Int J Mol Sci.* 2020 Feb 7;21(3). PMID: 32046099

References (continued)

31. Bubici et al. JNK signalling in cancer: in need of new, smarter therapeutic targets. *Br J Pharmacol*. 2014 Jan;171(1):24-37. PMID: 24117156
32. Yang et al. Mitogen-activated protein kinase kinase 4 deficiency in intrahepatic cholangiocarcinoma leads to invasive growth and epithelial-mesenchymal transition. *Hepatology*. 2015 Dec;62(6):1804-16. PMID: 26340507
33. Liu et al. Tyrosine phosphorylation activates 6-phosphogluconate dehydrogenase and promotes tumor growth and radiation resistance. *Nat Commun*. 2019 Mar 1;10(1):991. PMID: 30824700
34. Patra et al. The pentose phosphate pathway and cancer. *Trends Biochem Sci*. 2014 Aug;39(8):347-54. PMID: 25037503
35. Kowalik et al. Emerging Role of the Pentose Phosphate Pathway in Hepatocellular Carcinoma. *Front Oncol*. 2017;7:87. PMID: 28553614
36. Rao et al. O-GlcNAcylation of G6PD promotes the pentose phosphate pathway and tumor growth. *Nat Commun*. 2015 Sep 24;6:8468. PMID: 26399441
37. Korenjak et al. E2F-Rb complexes regulating transcription of genes important for differentiation and development. *Curr Opin Genet Dev*. 2005 Oct;15(5):520-7. doi: 10.1016/j.gde.2005.07.001. PMID: 16081278
38. Sachdeva et al. Understanding pRb: toward the necessary development of targeted treatments for retinoblastoma. *J. Clin. Invest*. 2012 Feb;122(2):425-34. PMID: 22293180
39. Dyson. RB1: a prototype tumor suppressor and an enigma. *Genes Dev*. 2016 Jul 1;30(13):1492-502. PMID: 27401552
40. Cobrinik. Pocket proteins and cell cycle control. *Oncogene*. 2005 Apr 18;24(17):2796-809. PMID: 15838516
41. Dommering et al. RB1 mutations and second primary malignancies after hereditary retinoblastoma. *Fam. Cancer*. 2012 Jun;11(2):225-33. PMID: 22205104
42. Anasua et al. Acute lymphoblastic leukemia as second primary tumor in a patient with retinoblastoma. *Oman J Ophthalmol*. May-Aug 2016;9(2):116-8. PMID: 27433042
43. Tanaka et al. Frequent allelic loss of the RB, D13S319 and D13S25 locus in myeloid malignancies with deletion/translocation at 13q14 of chromosome 13, but not in lymphoid malignancies. *Leukemia*. 1999 Sep;13(9):1367-73. PMID: 10482987
44. Gombos et al. Secondary acute myelogenous leukemia in patients with retinoblastoma: is chemotherapy a factor?. *Ophthalmology*. 2007 Jul;114(7):1378-83. PMID: 17613328
45. Patil et al. Checkpoint kinase 1 in DNA damage response and cell cycle regulation. *Cell. Mol. Life Sci*. 2013 Nov;70(21):4009-21. PMID: 23508805
46. Bartek et al. Chk1 and Chk2 kinases in checkpoint control and cancer. *Cancer Cell*. 2003 May;3(5):421-9. PMID: 12781359
47. Huang et al. Chk1 and Chk2 are differentially involved in homologous recombination repair and cell cycle arrest in response to DNA double-strand breaks induced by camptothecins. *Mol. Cancer Ther*. 2008 Jun;7(6):1440-9. PMID: 18566216
48. Zhang et al. Roles of Chk1 in cell biology and cancer therapy. *Int. J. Cancer*. 2014 Mar 1;134(5):1013-23. PMID: 23613359
49. Sen et al. CHK1 Inhibition in Small-Cell Lung Cancer Produces Single-Agent Activity in Biomarker-Defined Disease Subsets and Combination Activity with Cisplatin or Olaparib. *Cancer Res*. 2017 Jul 15;77(14):3870-3884. PMID: 28490518
50. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/208558s031lbl.pdf
51. <https://www.senhwabio.com/en/news/20220125>
52. Cardenas et al. The Expanding Role of the BCL6 Oncoprotein as a Cancer Therapeutic Target. *Clin Cancer Res*. 2017 Feb 15;23(4):885-893. PMID: 27881582
53. Pasqualucci et al. Mutations of the BCL6 proto-oncogene disrupt its negative autoregulation in diffuse large B-cell lymphoma. *Blood*. 2003 Apr 15;101(8):2914-23. PMID: 12515714
54. Liongue et al. B Cell Lymphoma 6 (BCL6): A Conserved Regulator of Immunity and Beyond. *Int J Mol Sci*. 2024 Oct 11;25(20). PMID: 39456751
55. Harris et al. Transcriptional repression of Stat6-dependent interleukin-4-induced genes by BCL-6: specific regulation of I epsilon transcription and immunoglobulin E switching. *Mol Cell Biol*. 1999 Oct;19(10):7264-75. PMID: 10490661
56. Delgado-Ramirez et al. Signal transducer and activator of transcription 6 as a target in colon cancer therapy. *Oncol Lett*. 2020 Jul;20(1):455-464. PMID: 32565970
57. Lossos et al. The BCL6 gene in B-cell lymphomas with 3q27 translocations is expressed mainly from the rearranged allele irrespective of the partner gene. *Leukemia*. 2003 Jul;17(7):1390-7. PMID: 12835729
58. Ye et al. Chromosomal translocations cause deregulated BCL6 expression by promoter substitution in B cell lymphoma. *EMBO J*. 1995 Dec 15;14(24):6209-17. PMID: 8557040
59. Beham-Schmid. Aggressive lymphoma 2016: revision of the WHO classification. *Memo*. 2017;10(4):248-254. PMID: 29250206

References (continued)

60. Alaggio et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Lymphoid Neoplasms. *Leukemia*. 2022 Jul;36(7):1720-1748. PMID: 35732829
61. Raju K et al. Double-hit B-cell lymphomas with BCL6 and MYC translocations are aggressive, frequently extranodal lymphomas distinct from BCL2 double-hit B-cell lymphomas. *Am J Sure Pathol*. 2013 Mar;37(3):323-32. PMID: 23348205
62. Li et al. MYC/BCL6 double-hit lymphoma (DHL): a tumour associated with an aggressive clinical course and poor prognosis. *Histopathology*. 2016 Jun;68(7):1090-8. PMID: 26426741
63. Hulpke et al. The MHC I loading complex: a multitasking machinery in adaptive immunity. *Trends Biochem Sci*. PMID: 23849087
64. Adams et al. The adaptable major histocompatibility complex (MHC) fold: structure and function of nonclassical and MHC class I-like molecules. *Annu Rev Immunol*. 2013;31:529-61. PMID: 23298204
65. Rossjohn et al. T cell antigen receptor recognition of antigen-presenting molecules. *Annu Rev Immunol*. 2015;33:169-200. PMID: 25493333
66. Parham. MHC class I molecules and KIRs in human history, health and survival. *Nat Rev Immunol*. 2005 Mar;5(3):201-14. PMID: 15719024
67. Sidney et al. HLA class I supertypes: a revised and updated classification. *BMC Immunol*. 2008 Jan 22;9:1. PMID: 18211710
68. Cornel et al. MHC Class I Downregulation in Cancer: Underlying Mechanisms and Potential Targets for Cancer Immunotherapy. *Cancers (Basel)*. 2020 Jul 2;12(7). PMID: 32630675
69. Maréchal et al. DNA damage sensing by the ATM and ATR kinases. *Cold Spring Harb Perspect Biol*. 2013 Sep 1;5(9). PMID: 24003211
70. Matsuoka et al. ATM and ATR substrate analysis reveals extensive protein networks responsive to DNA damage. *Science*. 2007 May 25;316(5828):1160-6. PMID: 17525332
71. Ditch et al. The ATM protein kinase and cellular redox signaling: beyond the DNA damage response. *Trends Biochem. Sci*. 2012 Jan;37(1):15-22. PMID: 22079189
72. Kozlov et al. Autophosphorylation and ATM activation: additional sites add to the complexity. *J. Biol. Chem*. 2011 Mar 18;286(11):9107-19. PMID: 21149446
73. Lim et al. Evaluation of the methods to identify patients who may benefit from PARP inhibitor use. *Endocr. Relat. Cancer*. 2016 Jun;23(6):R267-85. PMID: 27226207
74. Cynthia et al. Ataxia telangiectasia: a review. *Orphanet J Rare Dis*. 2016 Nov 25;11(1):159. PMID: 27884168
75. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/217439s003lbl.pdf
76. Gilardini Montani et al. ATM-depletion in breast cancer cells confers sensitivity to PARP inhibition. *CR*. PMID: 24252502
77. Pennington et al. Germline and somatic mutations in homologous recombination genes predict platinum response and survival in ovarian, fallopian tube, and peritoneal carcinomas. *Clin. Cancer Res*. 2014 Feb 1;20(3):764-75. PMID: 24240112
78. Mateo et al. DNA-Repair Defects and Olaparib in Metastatic Prostate Cancer. *N. Engl. J. Med*. 2015 Oct 29;373(18):1697-708. PMID: 26510020
79. Naqvi et al. Heterogeneity of the Treatment Effect with PARP Inhibitors in Metastatic Castration-resistant Prostate Cancer: A Living Interactive Systematic Review and Meta-analysis. *Eur Urol*. 2025 Jun;87(6):626-640. PMID: 39848867
80. Evans et al. Exploring the Impact of Treatment Switching on Overall Survival from the PROfound Study in Homologous Recombination Repair (HRR)-Mutated Metastatic Castration-Resistant Prostate Cancer (mCRPC). *Target Oncol*. 2021 Sep;16(5):613-623. PMID: 34478046
81. Li. Mechanisms and functions of DNA mismatch repair. *Cell Res*. 2008 Jan;18(1):85-98. PMID: 18157157
82. Zhao et al. Mismatch Repair Deficiency/Microsatellite Instability-High as a Predictor for anti-PD-1/PD-L1 Immunotherapy Efficacy. *J Hematol Oncol*. 12(1),54. PMID: 31151482
83. Berends et al. MLH1 and MSH2 protein expression as a pre-screening marker in hereditary and non-hereditary endometrial hyperplasia and cancer. *Int. J. Cancer*. 2001 May 1;92(3):398-403. PMID: 11291077
84. Lynch et al. Review of the Lynch syndrome: history, molecular genetics, screening, differential diagnosis, and medicolegal ramifications. *Clin. Genet*. 2009 Jul;76(1):1-18. PMID: 19659756
85. Martin et al. Therapeutic targeting of the DNA mismatch repair pathway. *Clin Cancer Res*. 2010 Nov 1;16(21):5107-13. PMID: 20823149
86. Baudrin et al. Molecular and Computational Methods for the Detection of Microsatellite Instability in Cancer. *Front Oncol*. 2018 Dec 12;8:621. doi: 10.3389/fonc.2018.00621. eCollection 2018. PMID: 30631754
87. Saeed et al. Microsatellites in Pursuit of Microbial Genome Evolution. *Front Microbiol*. 2016 Jan 5;6:1462. doi: 10.3389/fmicb.2015.01462. eCollection 2015. PMID: 26779133

References (continued)

88. Nojadeh et al. Microsatellite instability in colorectal cancer. *EXCLI J.* 2018;17:159-168. PMID: 29743854
89. Imai et al. Carcinogenesis and microsatellite instability: the interrelationship between genetics and epigenetics. *Carcinogenesis.* 2008 Apr;29(4):673-80. PMID: 17942460
90. Latham et al. Microsatellite Instability Is Associated With the Presence of Lynch Syndrome Pan-Cancer. *J. Clin. Oncol.* 2019 Feb 1;37(4):286-295. PMID: 30376427
91. Bonadona et al. Cancer risks associated with germline mutations in MLH1, MSH2, and MSH6 genes in Lynch syndrome. *JAMA.* 2011 Jun 8;305(22):2304-10. PMID: 21642682
92. Engel et al. Risks of less common cancers in proven mutation carriers with lynch syndrome. *J Clin Oncol.* 2012 Dec 10;30(35):4409-15. PMID: 23091106
93. Grant et al. Prevalence of germline mutations in cancer predisposition genes in patients with pancreatic cancer. *Gastroenterology.* 2015 Mar;148(3):556-64. PMID: 25479140
94. Hu et al. Association Between Inherited Germline Mutations in Cancer Predisposition Genes and Risk of Pancreatic Cancer. *JAMA.* 2018 Jun 19;319(23):2401-2409. PMID: 29922827
95. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/125514s178lbl.pdf
96. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/125554s131lbl.pdf
97. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/125377s136lbl.pdf
98. Buccoliero et al. Pediatric High Grade Glioma Classification Criteria and Molecular Features of a Case Series. *Genes (Basel).* 2022 Mar 31;13(4). PMID: 35456430
99. Friker et al. MSH2, MSH6, MLH1, and PMS2 immunohistochemistry as highly sensitive screening method for DNA mismatch repair deficiency syndromes in pediatric high-grade glioma. *Acta Neuropathol.* 2025 Feb 2;149(1):11. PMID: 39894875
100. So et al. The TNF-TNFR Family of Co-signal Molecules. *Adv Exp Med Biol.* 2019;1189:53-84. PMID: 31758531
101. Costello et al. Stimulation of non-Hodgkin's lymphoma via HVEM: an alternate and safe way to increase Fas-induced apoptosis and improve tumor immunogenicity. *Leukemia.* 2003 Dec;17(12):2500-7. PMID: 14562115
102. NCCN Guidelines® - NCCN-B-Cell Lymphomas [Version 3.2025]
103. Launay et al. High rate of TNFRSF14 gene alterations related to 1p36 region in de novo follicular lymphoma and impact on prognosis. *Leukemia.* 2012 Mar;26(3):559-62. PMID: 21941365
104. Cheung et al. Acquired TNFRSF14 mutations in follicular lymphoma are associated with worse prognosis. *Cancer Res.* 2010 Nov 15;70(22):9166-74. PMID: 20884631
105. Nag et al. The MDM2-p53 pathway revisited. *J Biomed Res.* 2013 Jul;27(4):254-71. PMID: 23885265
106. Muller et al. Mutant p53 in cancer: new functions and therapeutic opportunities. *Cancer Cell.* 2014 Mar 17;25(3):304-17. PMID: 24651012
107. Olivier et al. TP53 mutations in human cancers: origins, consequences, and clinical use. *Cold Spring Harb Perspect Biol.* 2010 Jan;2(1):a001008. PMID: 20182602
108. Guha et al. Inherited TP53 Mutations and the Li-Fraumeni Syndrome. *Cold Spring Harb Perspect Med.* 2017 Apr 3;7(4). PMID: 28270529
109. Peter S et al. Comprehensive genomic characterization of squamous cell lung cancers. *Nature.* 2012 Sep 27;489(7417):519-25. PMID: 22960745
110. Cancer Genome Atlas Network. Comprehensive genomic characterization of head and neck squamous cell carcinomas. *Nature.* 2015 Jan 29;517(7536):576-82. PMID: 25631445
111. Campbell et al. Distinct patterns of somatic genome alterations in lung adenocarcinomas and squamous cell carcinomas. *Nat. Genet.* 2016 Jun;48(6):607-16. PMID: 27158780
112. Cancer Genome Atlas Research Network. Integrated genomic characterization of oesophageal carcinoma. *Nature.* 2017 Jan 12;541(7636):169-175. doi: 10.1038/nature20805. Epub 2017 Jan 4. PMID: 28052061
113. Olivier et al. The IARC TP53 database: new online mutation analysis and recommendations to users. *Hum. Mutat.* 2002 Jun;19(6):607-14. PMID: 12007217
114. Rivlin et al. Mutations in the p53 Tumor Suppressor Gene: Important Milestones at the Various Steps of Tumorigenesis. *Genes Cancer.* 2011 Apr;2(4):466-74. PMID: 21779514
115. Petitjean et al. TP53 mutations in human cancers: functional selection and impact on cancer prognosis and outcomes. *Oncogene.* 2007 Apr 2;26(15):2157-65. PMID: 17401424
116. Soussi et al. Recommendations for analyzing and reporting TP53 gene variants in the high-throughput sequencing era. *Hum. Mutat.* 2014 Jun;35(6):766-78. PMID: 24729566

References (continued)

117. <https://www.globenewswire.com/news-release/2020/10/13/2107498/0/en/PMV-Pharma-Granted-FDA-Fast-Track-Designation-of-PC14586-for-the-Treatment-of-Advanced-Cancer-Patients-that-have-Tumors-with-a-p53-Y220C-Mutation.html>
118. Parrales et al. Targeting Oncogenic Mutant p53 for Cancer Therapy. *Front Oncol.* 2015 Dec 21;5:288. doi: 10.3389/fonc.2015.00288. eCollection 2015. PMID: 26732534
119. Zhao et al. Molecularly targeted therapies for p53-mutant cancers. *Cell. Mol. Life Sci.* 2017 Nov;74(22):4171-4187. PMID: 28643165
120. Louis et al. The 2021 WHO Classification of Tumors of the Central Nervous System: a summary. *Neuro Oncol.* 2021 Aug 2;23(8):1231-1251. PMID: 34185076
121. NCCN Guidelines® - NCCN-Chronic Lymphocytic Leukemia/Small Lymphocytic Lymphoma [Version 1.2026]
122. NCCN Guidelines® - NCCN-Acute Lymphoblastic Leukemia [Version 2.2025]
123. Bernard et al. Implications of TP53 allelic state for genome stability, clinical presentation and outcomes in myelodysplastic syndromes. *Nat. Med.* 2020 Aug 3. PMID: 32747829
124. Lander et al. Initial sequencing and analysis of the human genome. *Nature.* 2001 Feb 15;409(6822):860-921. PMID: 11237011
125. Boland et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res.* 1998 Nov 15;58(22):5248-57. PMID: 9823339
126. Halford et al. Low-level microsatellite instability occurs in most colorectal cancers and is a nonrandomly distributed quantitative trait. *Cancer Res.* 2002 Jan 1;62(1):53-7. PMID: 11782358
127. NCCN Guidelines® - NCCN-Colon Cancer [Version 5.2025]
128. Pawlik et al. Colorectal carcinogenesis: MSI-H versus MSI-L. *Dis. Markers.* 2004;20(4-5):199-206. PMID: 15528785
129. Lee et al. Low-Level Microsatellite Instability as a Potential Prognostic Factor in Sporadic Colorectal Cancer. *Medicine (Baltimore).* 2015 Dec;94(50):e2260. PMID: 26683947
130. Cortes-Ciriano et al. A molecular portrait of microsatellite instability across multiple cancers. *Nat Commun.* 2017 Jun 6;8:15180. doi: 10.1038/ncomms15180. PMID: 28585546
131. Bonneville et al. Landscape of Microsatellite Instability Across 39 Cancer Types. *JCO Precis Oncol.* 2017;2017. PMID: 29850653
132. https://www.accessdata.fda.gov/drugsatfda_docs/label/2024/761174s009lbl.pdf
133. NCCN Guidelines® - NCCN-Rectal Cancer [Version 4.2025]
134. Ribic et al. Tumor microsatellite-instability status as a predictor of benefit from fluorouracil-based adjuvant chemotherapy for colon cancer. *N. Engl. J. Med.* 2003 Jul 17;349(3):247-57. PMID: 12867608
135. Klingbiel et al. Prognosis of stage II and III colon cancer treated with adjuvant 5-fluorouracil or FOLFIRI in relation to microsatellite status: results of the PETACC-3 trial. *Ann. Oncol.* 2015 Jan;26(1):126-32. PMID: 25361982
136. Hermel et al. The Emerging Role of Checkpoint Inhibition in Microsatellite Stable Colorectal Cancer. *J Pers Med.* 2019 Jan 16;9(1). PMID: 30654522
137. Ciardiello et al. Immunotherapy of colorectal cancer: Challenges for therapeutic efficacy. *Cancer Treat. Rev.* 2019 Jun;76:22-32. PMID: 31079031
138. Volinia et al. Molecular cloning, cDNA sequence, and chromosomal localization of the human phosphatidylinositol 3-kinase p110 alpha (PIK3CA) gene. *Genomics.* 1994 Dec;24(3):472-7. PMID: 7713498
139. Whale et al. Functional characterization of a novel somatic oncogenic mutation of PIK3CB. *Signal Transduct Target Ther.* 2017;2:17063. PMID: 29279775
140. Osaki et al. PI3K-Akt pathway: its functions and alterations in human cancer. *Apoptosis.* 2004 Nov;9(6):667-76. PMID: 15505410
141. Cantley. The phosphoinositide 3-kinase pathway. *Science.* 2002 May 31;296(5573):1655-7. PMID: 12040186
142. Fruman et al. The PI3K Pathway in Human Disease. *Cell.* 2017 Aug 10;170(4):605-635. PMID: 28802037
143. Engelman et al. The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat. Rev. Genet.* 2006 Aug;7(8):606-19. PMID: 16847462
144. Vanhaesebroeck et al. PI3K signalling: the path to discovery and understanding. *Nat. Rev. Mol. Cell Biol.* 2012 Feb 23;13(3):195-203. PMID: 22358332
145. Yuan et al. PI3K pathway alterations in cancer: variations on a theme. *Oncogene.* 2008 Sep 18;27(41):5497-510. PMID: 18794884
146. Liu et al. Targeting the phosphoinositide 3-kinase pathway in cancer. *Nat Rev Drug Discov.* 2009 Aug;8(8):627-44. PMID: 19644473
147. Hanahan et al. Hallmarks of cancer: the next generation. *Cell.* 2011 Mar 4;144(5):646-74. PMID: 21376230

References (continued)

148. Brito et al. PIK3CA Mutations in Diffuse Gliomas: An Update on Molecular Stratification, Prognosis, Recurrence, and Aggressiveness. *Clin Med Insights Oncol.* 2022;16:11795549211068804. PMID: 35023985
149. Huret et al. Atlas of genetics and cytogenetics in oncology and haematology in 2013. *Nucleic Acids Res.* 2013 Jan;41(Database issue):D920-4. PMID: 23161685
150. Miled et al. Mechanism of two classes of cancer mutations in the phosphoinositide 3-kinase catalytic subunit. *Science.* 2007 Jul 13;317(5835):239-42. PMID: 17626883
151. Burke et al. Synergy in activating class I PI3Ks. *Trends Biochem. Sci.* 2015 Feb;40(2):88-100. PMID: 25573003
152. Burke et al. Oncogenic mutations mimic and enhance dynamic events in the natural activation of phosphoinositide 3-kinase p110 α (PIK3CA). *Proc. Natl. Acad. Sci. U.S.A.* 2012 Sep 18;109(38):15259-64. PMID: 22949682
153. https://www.accessdata.fda.gov/drugsatfda_docs/label/2024/212526s009lbl.pdf
154. Mayer et al. A Phase Ib Study of Alpelisib (BYL719), a PI3K α -Specific Inhibitor, with Letrozole in ER+/HER2- Metastatic Breast Cancer. *Clin. Cancer Res.* 2017 Jan 1;23(1):26-34. PMID: 27126994
155. Mayer et al. A Phase II Randomized Study of Neoadjuvant Letrozole Plus Alpelisib for Hormone Receptor-Positive, Human Epidermal Growth Factor Receptor 2-Negative Breast Cancer (NEO-ORB). *Clin. Cancer Res.* 2019 Feb 5. PMID: 30723140
156. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/218197s002lbl.pdf
157. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/219249s002lbl.pdf
158. Jung et al. Pilot study of sirolimus in patients with PIK3CA mutant/amplified refractory solid cancer. *Mol Clin Oncol.* 2017 Jul;7(1):27-31. PMID: 28685070
159. Janku et al. PIK3CA mutations in patients with advanced cancers treated with PI3K/AKT/mTOR axis inhibitors. *Mol. Cancer Ther.* 2011 Mar;10(3):558-65. PMID: 21216929
160. Liao et al. Aspirin use, tumor PIK3CA mutation, and colorectal-cancer survival. *N Engl J Med.* 2012 Oct 25;367(17):1596-606. PMID: 23094721
161. Domingo et al. Evaluation of PIK3CA mutation as a predictor of benefit from nonsteroidal anti-inflammatory drug therapy in colorectal cancer. *J Clin Oncol.* 2013 Dec 1;31(34):4297-305. PMID: 24062397
162. <https://www.cancernetwork.com/view/fda-grants-fast-track-designation-to-novel-pik3-inhibitor-in-breast-cancer>
163. Huang et al. ENO1 and Cancer. *Mol Ther Oncolytics.* 2022 Mar 17;24:288-298. PMID: 35434271
164. Almaguel et al. Alpha-Enolase: Emerging Tumor-Associated Antigen, Cancer Biomarker, and Oncotherapeutic Target. *Front Genet.* 2020;11:614726. PMID: 33584813
165. Qiao et al. Enolase 1, a Moonlighting Protein, as a Potential Target for Cancer Treatment. *Int J Biol Sci.* 2021;17(14):3981-3992. PMID: 34671213
166. Xu et al. Enolase 1 Correlated With Cancer Progression and Immune-Infiltrating in Multiple Cancer Types: A Pan-Cancer Analysis. *Front Oncol.* 2020;10:593706. PMID: 33643901
167. King et al. Amplification of a novel v-erbB-related gene in a human mammary carcinoma. *Science.* 1985 Sep 6;229(4717):974-6. PMID: 2992089
168. Hsu et al. The role of HER2, EGFR, and other receptor tyrosine kinases in breast cancer. *Cancer Metastasis Rev.* 2016 Dec;35(4):575-588. PMID: 27913999
169. Gutierrez et al. HER2: biology, detection, and clinical implications. *Arch. Pathol. Lab. Med.* 2011 Jan;135(1):55-62. PMID: 21204711
170. Di Fiore et al. erbB-2 is a potent oncogene when overexpressed in NIH/3T3 cells. *Science.* 1987 Jul 10;237(4811):178-82. PMID: 2885917
171. Hudziak et al. Increased expression of the putative growth factor receptor p185HER2 causes transformation and tumorigenesis of NIH 3T3 cells. *Proc. Natl. Acad. Sci. U.S.A.* 1987 Oct;84(20):7159-63. PMID: 2890160
172. Lonardo et al. The normal erbB-2 product is an atypical receptor-like tyrosine kinase with constitutive activity in the absence of ligand. *New Biol.* 1990 Nov;2(11):992-1003. PMID: 1983208
173. Ciriello et al. Comprehensive Molecular Portraits of Invasive Lobular Breast Cancer. *Cell.* 2015 Oct 8;163(2):506-19. PMID: 26451490
174. Cancer Genome Atlas Research Network. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature.* 2014 Sep 11;513(7517):202-9. doi: 10.1038/nature13480. Epub 2014 Jul 23. PMID: 25079317
175. Cancer Genome Atlas Research Network. Comprehensive molecular characterization of urothelial bladder carcinoma. *Nature.* 2014 Mar 20;507(7492):315-22. doi: 10.1038/nature12965. Epub 2014 Jan 29. PMID: 24476821

References (continued)

176. Donna M et al. Comprehensive molecular characterization of human colon and rectal cancer. *Nature*. 2012 Jul 18;487(7407):330-7. PMID: 22810696
177. Cancer Genome Atlas Research Network. Comprehensive molecular profiling of lung adenocarcinoma. *Nature*. 2014 Jul 31;511(7511):543-50. doi: 10.1038/nature13385. Epub 2014 Jul 9. PMID: 25079552
178. Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. *Nature*. 2011 Jun 29;474(7353):609-15. PMID: 21720365
179. Petrelli et al. Clinical and pathological characterization of HER2 mutations in human breast cancer: a systematic review of the literature. *Breast Cancer Res. Treat.* 2017 Nov;166(2):339-349. PMID: 28762010
180. Bose et al. Activating HER2 mutations in HER2 gene amplification negative breast cancer. *Cancer Discov.* 2013 Feb;3(2):224-37. doi: 10.1158/2159-8290.CD-12-0349. Epub 2012 Dec 7. PMID: 23220880
181. Hudis. Trastuzumab--mechanism of action and use in clinical practice. *N. Engl. J. Med.* 2007 Jul 5;357(1):39-51. PMID: 17611206
182. Slamon et al. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science*. 1987 Jan 9;235(4785):177-82. PMID: 3798106
183. https://www.accessdata.fda.gov/drugsatfda_docs/label/2024/103792s5354lbl.pdf
184. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/125409s139lbl.pdf
185. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/125427s121lbl.pdf
186. NCCN Guidelines® - NCCN-Breast Cancer [Version 5.2025]
187. https://www.accessdata.fda.gov/drugsatfda_docs/label/2022/022059s031lbl.pdf
188. https://www.accessdata.fda.gov/drugsatfda_docs/label/2021/208051s009lbl.pdf
189. https://www.accessdata.fda.gov/drugsatfda_docs/label/2023/213411s004lbl.pdf
190. https://www.accessdata.fda.gov/drugsatfda_docs/nda/2024/761416Orig1s000Lbl.pdf
191. https://www.accessdata.fda.gov/drugsatfda_docs/label/2023/761150s005lbl.pdf
192. <https://www.targetedonc.com/view/her2targeted-antibody-zw25-earns-fda-fast-track-designation-in-gea>
193. <https://www.prnewswire.com/news-releases/remegen-announces-us-fda-has-granted-breakthrough-therapy-designation-for-disitamab-vedotin-rc48-in-urothelial-cancer-301138315.html>
194. <https://ir.ambrx.com/news/news-details/2023/ACE-Breast-02-Pivotal-Phase-3-Study-of-Ambrxs-ARX788-for-the-Treatment-of-HER2-Positive-Metastatic-Breast-Cancer-Achieves-Positive-Results/default.aspx>
195. Ma et al. Neratinib Efficacy and Circulating Tumor DNA Detection of HER2 Mutations in HER2 Nonamplified Metastatic Breast Cancer. *Clin. Cancer Res.* 2017 Oct 1;23(19):5687-5695. PMID: 28679771
196. De Grève et al. Clinical activity of afatinib (BIBW 2992) in patients with lung adenocarcinoma with mutations in the kinase domain of HER2/neu. *Lung Cancer*. 2012 Apr;76(1):123-7. PMID: 22325357
197. Kris et al. Targeting HER2 aberrations as actionable drivers in lung cancers: phase II trial of the pan-HER tyrosine kinase inhibitor dacomitinib in patients with HER2-mutant or amplified tumors. *Ann. Oncol.* 2015 Jul;26(7):1421-7. PMID: 25899785
198. Falchook et al. Non-small-cell lung cancer with HER2 exon 20 mutation: regression with dual HER2 inhibition and anti-VEGF combination treatment. *J Thorac Oncol.* 2013 Feb;8(2):e19-20. PMID: 23328556
199. David M et al. Neratinib in HER2- or HER3-mutant solid tumors: SUMMIT, a global, multi-histology, open-label, phase 2 'basket' study. *AACR 2017. Abstract CT001*
200. Lin et al. Response to Afatinib in a Patient with Non-Small Cell Lung Cancer Harboring HER2 R896G Mutation: A Case Report. *Onco Targets Ther.* 2019;12:10897-10902. PMID: 31849493
201. Chang et al. Sustained Partial Response to Afatinib in a Patient With Lung Adenocarcinoma Harboring HER2V659E Mutation. *JCO Precis Oncol.* 2020 Aug; 912-915. PMID: 35050762
202. Nayar et al. Acquired HER2 mutations in ER+ metastatic breast cancer confer resistance to estrogen receptor-directed therapies. *Nat. Genet.* 2019 Feb;51(2):207-216. PMID: 30531871
203. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/219042s000lbl.pdf
204. https://www.accessdata.fda.gov/drugsatfda_docs/label/2025/219972s000lblCorrected.pdf
205. <https://investor.greenwichlifesciences.com/news-events/press-releases/detail/102/us-fda-fast-track-designation>