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**Report Date**: 24 Oct 2025 1 of 26

Patient Name: 전관식 Gender: M Sample ID: N25-258 Primary Tumor Site: lung
Collection Date: 2025.09.18

# Sample Cancer Type: Lung Cancer

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# **Relevant Lung Cancer Findings**

Gene	Finding		Gene	Finding	
ALK	None detected		NTRK1	None detected	
BRAF	None detected		NTRK2	None detected	
EGFR	None detected		NTRK3	None detected	
ERBB2	None detected		RET	None detected	
KRAS	KRAS p.(G120	C) c.34G>T	ROS1	None detected	
MET	None detected				
Genomic Alt	eration	Finding			
Tumor Mu	ıtational Burden	6.63 Mut/Mb measured			

### **Relevant Biomarkers**

Tier	Genomic Alteration	Relevant Therapies (In this cancer type)	Relevant Therapies (In other cancer type)	Clinical Trials
IA	KRAS p.(G12C) c.34G>T  KRAS proto-oncogene, GTPase Allele Frequency: 34.94%  Locus: chr12:25398285  Transcript: NM_033360.4	adagrasib 1,2/II+ sotorasib 1,2/II+	adagrasib + cetuximab 1 / I, II+ panitumumab + sotorasib 1 / I, II+ adagrasib I, II+ adagrasib + panitumumab I, II+ cetuximab + sotorasib I, II+ sotorasib I, II+ bevacizumab + chemotherapy I	76
IIC	MTAP deletion methylthioadenosine phosphorylase Locus: chr9:21802646	None*	None*	10
IIC	CDKN2A deletion None* None*  cyclin dependent kinase inhibitor 2A  Locus: chr9:21968178		None*	3

 $<sup>\</sup>hbox{* \bf Public data sources included in relevant the rapies: FDA1, NCCN, EMA2, ESMO}$ 

**Line of therapy:** I: First-line therapy, II+: Other line of therapy

**Tier Reference:** Li et al. Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. J Mol Diagn. 2017 Jan;19(1):4-23.

<sup>\*</sup> Public data sources included in prognostic and diagnostic significance: NCCN, ESMO

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# **Relevant Biomarkers (continued)**

Tier	Genomic Alteration	Relevant Therapies (In this cancer type)	Relevant Therapies (In other cancer type)	Clinical Trials
IIC	CDKN2B deletion  cyclin dependent kinase inhibitor 2B  Locus: chr9:22005728	None*	None*	1
IIC	FANCM deletion  FA complementation group M  Locus: chr14:45605157	None*	None*	1

<sup>\*</sup> Public data sources included in relevant therapies: FDA1, NCCN, EMA2, ESMO

Line of therapy: I: First-line therapy, II+: Other line of therapy

Tier Reference: Li et al. Standards and Guidelines for the Interpretation and Reporting of Sequence Variants in Cancer: A Joint Consensus Recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. J Mol Diagn. 2017 Jan;19(1):4-23.

🔼 Alerts informed by public data sources: 🤣 Contraindicated, 🛡 Resistance, 🗳 Breakthrough, 🗚 Fast Track

KRAS p.(G12C) c.34G>T

A avutometinib + sotorasib 1, BBO-8520 1, BBP-398 + sotorasib 1

Public data sources included in alerts: FDA1, NCCN, EMA2, ESMO

### Prevalent cancer biomarkers without relevant evidence based on included data sources

ERCC2 deletion, MAP2K4 p.(I73Rfs\*9) c.217\_218delAT, MLH3 deletion, Microsatellite stable, RAD51B deletion, RAD52 p. (S346\*) c.1037C>A, XRCC3 deletion, NOTCH2 deletion, HLA-A deletion, HLA-B deletion, MAX p.(R60L) c.179G>T, DICER1 deletion, NCOR1 deletion, ARHGAP35 deletion, Tumor Mutational Burden

### **Variant Details**

DNA Sequence Variants							
Amino Acid Change	Coding	Variant ID	Locus	Allele Frequency	Transcript	Variant Effect	
p.(G12C)	c.34G>T	COSM516	chr12:25398285	34.94%	NM_033360.4	missense	
p.(I73Rfs*9)	c.217_218delAT		chr17:11958306	46.11%	NM_003010.4	frameshift Deletion	
p.(S346*)	c.1037C>A		chr12:1023218	54.69%	NM_134424.4	nonsense	
p.(R60L)	c.179G>T		chr14:65544747	42.80%	NM_002382.5	missense	
p.(I1154N)	c.3461_3462delTAinsA T	١.	chr2:50464011	27.83%	NM_004801.5	missense	
p.(Y695D)	c.2083T>G		chr3:121659747	48.70%	NM_021082.4	missense	
p.(A57_A62del)	c.162_179delTGCAGC GGCCGCAGCGGC		chr5:79950707	42.94%	NM_002439.5	nonframeshift Deletion	
p.(D365H)	c.1093G>C		chr8:104337427	33.75%	NM_001164615.2	missense	
p.(?)	c.3644-2A>G		chr9:139401427	29.34%	NM_017617.5	unknown	
p.(N1599K)	c.4797C>A		chr16:72831784	28.31%	NM_006885.4	missense	
p.(R1371L)	c.4112G>T		chr19:15288627	32.94%	NM_000435.3	missense	
	Amino Acid Change p.(G12C) p.(I73Rfs*9) p.(S346*) p.(R60L) p.(I1154N) p.(Y695D) p.(A57_A62del) p.(D365H) p.(?) p.(N1599K)	Amino Acid Change       Coding         p.(G12C)       c.34G>T         p.(I73Rfs*9)       c.217_218delAT         p.(S346*)       c.1037C>A         p.(R60L)       c.179G>T         p.(I1154N)       c.3461_3462delTAinsAT         p.(Y695D)       c.2083T>G         p.(A57_A62del)       c.162_179delTGCAGC         GGCCGCAGCGGC       c.1093G>C         p.(P)       c.3644-2A>G         p.(N1599K)       c.4797C>A	Amino Acid Change         Coding         Variant ID           p.(G12C)         c.34G>T         COSM516           p.(I73Rfs*9)         c.217_218delAT         .           p.(S346*)         c.1037C>A         .           p.(R60L)         c.179G>T         .           p.(I1154N)         c.3461_3462delTAinsA         .           p.(Y695D)         c.2083T>G         .           p.(A57_A62del)         c.162_179delTGCAGC         .           p.(D365H)         c.1093G>C         .           p.(?)         c.3644-2A>G         .           p.(N1599K)         c.4797C>A         .	Amino Acid Change         Coding         Variant ID         Locus           p.(G12C)         c.34G>T         COSM516         chr12:25398285           p.(I73Rfs*9)         c.217_218deIAT         chr17:11958306           p.(S346*)         c.1037C>A         chr12:1023218           p.(R60L)         c.179G>T         chr14:65544747           p.(I1154N)         c.3461_3462deITAinsA         chr2:50464011           p.(Y695D)         c.2083T>G         chr3:121659747           p.(A57_A62del)         c.162_179deITGCAGC GGCCGGC         chr5:79950707           p.(D365H)         c.1093G>C         chr8:104337427           p.(?)         c.3644-2A>G         chr9:139401427           p.(N1599K)         c.4797C>A         chr16:72831784	Amino Acid Change         Coding         Variant ID         Locus         Allele Frequency           p.(G12C)         c.34G>T         COSM516         chr12:25398285         34.94%           p.(I73Rfs*9)         c.217_218delAT         chr17:11958306         46.11%           p.(S346*)         c.1037C>A         chr12:1023218         54.69%           p.(R60L)         c.179G>T         chr14:65544747         42.80%           p.(I1154N)         c.3461_3462delTAinsA         chr2:50464011         27.83%           p.(Y695D)         c.2083T>G         chr3:121659747         48.70%           p.(A57_A62del)         c.162_179delTGCAGC GGCCGCAGCGGC         chr5:79950707         42.94%           p.(D365H)         c.1093G>C         chr8:104337427         33.75%           p.(?)         c.3644-2A>G         chr9:139401427         29.34%           p.(N1599K)         c.4797C>A         chr16:72831784         28.31%	Amino Acid Change         Coding         Variant ID         Locus         Frequency         Transcript           p.(G12C)         c.346>T         COSM516         chr12:25398285         34.94         NM_033360.4           p.(I73Rfs*9)         c.217_218delAT         chr17:11958306         46.11%         NM_003010.4           p.(S346*)         c.1037C>A         chr12:1023218         54.69%         NM_134424.4           p.(R60L)         c.179G>T         chr14:65544747         42.80%         NM_002382.5           p.(I1154N)         c.3461_3462delTAins → Chr2:50464011         27.83%         NM_004801.5           p.(Y695D)         c.2083T>G         chr3:121659747         48.70%         NM_021082.4           p.(A57_A62del)         c.162_179delTGCAGC GGCC         chr5:79950707         42.94%         NM_002439.5           p.(D365H)         c.1093G>C         chr8:104337427         33.75%         NM_001164615.2           p.(?)         c.3644-2A>G         chr9:139401427         29.34%         NM_017617.5           p.(N1599K)         c.4797C>A         chr16:72831784         28.31%         NM_006885.4	

<sup>\*</sup> Public data sources included in prognostic and diagnostic significance: NCCN, ESMO

### **Variant Details (continued)**

Copy Number Variations							
Gene	Locus	Copy Number	CNV Ratio				
MTAP	chr9:21802646	0.08	0.39				
CDKN2A	chr9:21968178	0	0.34				
CDKN2B	chr9:22005728	0	0.35				
FANCM	chr14:45605157	1.03	0.69				
ERCC2	chr19:45854865	0.88	0.64				
MLH3	chr14:75483761	0.95	0.67				
RAD51B	chr14:68290164	1	0.67				
XRCC3	chr14:104165043	0.73	0.6				
NOTCH2	chr1:120457903	0.94	0.66				
HLA-A	chr6:29910229	1.03	0.69				
HLA-B	chr6:31322252	0.38	0.48				
DICER1	chr14:95556791	0.92	0.66				
NCOR1	chr17:15935586	0.94	0.66				
ARHGAP35	chr19:47421913	0.91	0.65				
NRAS	chr1:115251152	1.05	0.69				
FOXA1	chr14:38060550	0.86	0.63				
AKT1	chr14:105236628	0.7	0.58				

# **Biomarker Descriptions**

KRAS p.(G12C) c.34G>T

KRAS proto-oncogene, GTPase

<u>Background:</u> The KRAS proto-oncogene encodes a GTPase that functions in signal transduction and is a member of the RAS superfamily which also includes NRAS and HRAS. RAS proteins mediate the transmission of growth signals from the cell surface to the nucleus via the PI3K/AKT/MTOR and RAS/RAF/MEK/ERK pathways, which regulate cell division, differentiation, and survival<sup>1,2,3</sup>.

Alterations and prevalence: Recurrent mutations in RAS oncogenes cause constitutive activation and are found in 20-30% of cancers. KRAS mutations are observed in up to 10-20% of uterine cancer, 30-35% of lung adenocarcinoma and colorectal cancer, and about 60% of pancreatic cancer<sup>4</sup>. The majority of KRAS mutations consist of point mutations occurring at G12, G13, and Q61<sup>4,5,6</sup>. Mutations at A59, K117, and A146 have also been observed but are less frequent<sup>7,8</sup>.

Potential relevance: The FDA has approved the small molecule inhibitors, sotorasib<sup>9</sup> (2021) and adagrasib<sup>10</sup> (2022), for the treatment of adult patients with KRAS G12C-mutated locally advanced or metastatic non-small cell lung cancer (NSCLC). Sotorasib and adagrasib are also useful in certain circumstances for KRAS G12C-mutated pancreatic adenocarcinoma<sup>11</sup>. The FDA has also granted breakthrough therapy designation (2022) to the KRAS G12C inhibitor, GDC-6036<sup>12</sup>, for KRAS G12C-mutated non-small cell lung cancer. The SHP2 inhibitor, BBP-398<sup>13</sup> was granted fast track designation (2022) in combination with sotorasib for previously treated patients with KRAS G12C-mutated metastatic NSCLC. The RAF/MEK clamp, avutometinib<sup>14</sup> was also granted fast track designation (2024) in combination with sotorasib for KRAS G12C-mutated metastatic NSCLC who have received at least one prior systemic therapy and have not been previously treated with a KRAS G12C inhibitor. The KRAS G12C inhibitor, BBO-8520<sup>15</sup>, was granted fast track designation in 2025 for previously treated KRAS G12C-mutated patients with metastatic NSCLC. The KRAS G12C inhibitor, D3S-001<sup>16</sup>, was granted fast track designation in 2024 for KRAS G12C-mutated patients with advanced unresectable or metastatic colorectal cancers. The PLK1 inhibitor, onvansertib<sup>17</sup>, was granted fast track designation (2020) in combination with bevacizumab and FOLFIRI for second-line treatment of patients with KRAS-mutated metastatic colorectal cancer (mCRC). The EGFR antagonists, cetuximab<sup>18</sup>

# **Biomarker Descriptions (continued)**

and panitumumab<sup>19</sup>, are contraindicated for treatment of colorectal cancer patients with KRAS mutations in exon 2 (codons 12 and 13), exon 3 (codons 59 and 61), and exon 4 (codons 117 and 146)<sup>8</sup>. Additionally, KRAS mutations are associated with poor prognosis in NSCLC<sup>20</sup>.

#### MTAP deletion

methylthioadenosine phosphorylase

Background: The MTAP gene encodes methylthioadenosine phosphorylase<sup>21</sup>. Methylthioadenosine phosphorylase, a key enzyme in polyamine biosynthesis and methionine salvage pathways, catalyzes the reversible phosphorylation of S-methyl-5'-thioadenosine (MTA) to adenine and 5-methylthioribose-1-phosphate<sup>57,58</sup>. Loss of MTAP function is commonly observed in cancer due to deletion or promotor methylation which results in the loss of MTA phosphorylation and sensitivity of MTAP-deficient cells to purine synthesis inhibitors and to methionine deprivation<sup>58</sup>.

Alterations and prevalence: MTAP is flanked by CDKN2A tumor suppressor on chromosome 9p21 and is frequently found to be codeleted with CDKN2A in numerous solid and hematological cancers<sup>58,59</sup>. Consequently, biallelic loss of MTAP has been observed in 42% of glioblastoma multiforme, 32% of mesothelioma, 26% of bladder urothelial carcinoma, 22% of pancreatic adenocarcinoma, 21% of esophageal adenocarcinoma, 20% of lung squamous cell carcinoma and skin cutaneous melanoma, 15% of diffuse large B-cell lymphoma and head and neck squamous cell carcinoma, 12% of lung adenocarcinoma, 11% of cholangiocarcinoma, 9% of sarcoma, stomach adenocarcinoma and brain lower grade glioma, and 3% of ovarian serous cystadenocarcinoma, breast invasive carcinoma, adrenocortical carcinoma, thymoma and liver hepatocellular carcinoma<sup>4,7</sup>. Somatic mutations in MTAP have been found in 3% of uterine corpus endometrial carcinoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for MTAP aberrations.

#### **CDKN2A** deletion

cyclin dependent kinase inhibitor 2A

Background: CDKN2A encodes cyclin dependent kinase inhibitor 2A, a cell cycle regulator that controls G1/S progression<sup>21</sup>. CDKN2A, also known as p16/INK4A, belongs to a family of INK4 cyclin-dependent kinase inhibitors, which also includes CDKN2B (p15/INK4B), CDKN2C (p18/INK4C), and CDKN2D (p19/INK4D)<sup>66</sup>. The INK4 family regulates cell cycle progression by inhibiting CDK4 or CDK6, thereby preventing the phosphorylation of Rb<sup>67,68,69</sup>. CDKN2A encodes two alternative transcript variants, namely p16 and p14ARF, both of which exhibit differential tumor suppressor functions<sup>70</sup>. Specifically, the CDKN2A/p16 transcript inhibits cell cycle kinases CDK4 and CDK6, whereas the CDKN2A/p14ARF transcript stabilizes the tumor suppressor protein p53 to prevent its degradation<sup>21,70,71</sup>. CDKN2A aberrations commonly co-occur with CDKN2B<sup>66</sup>. Loss of CDKN2A/p16 results in downstream inactivation of the Rb and p53 pathways, leading to uncontrolled cell proliferation<sup>72</sup>. Germline mutations of CDKN2A are known to confer a predisposition to melanoma and pancreatic cancer<sup>73,74</sup>.

Alterations and prevalence: Somatic alterations in CDKN2A often result in loss of function (LOF) which is attributed to copy number loss, truncating, or missense mutations<sup>75</sup>. Somatic mutations in CDKN2A are observed in 20% of head and neck squamous cell carcinoma and pancreatic adenocarcinoma, 15% of lung squamous cell carcinoma, 13% of skin cutaneous melanoma, 8% of esophageal adenocarcinoma, 7% of bladder urothelial carcinoma, 6% of cholangiocarcinoma, 4% of lung adenocarcinoma and stomach adenocarcinoma, and 2% of liver hepatocellular carcinoma, uterine carcinosarcoma, and cervical squamous cell carcinoma<sup>4,7</sup>. Biallelic deletion of CDKN2A is observed in 56% of glioblastoma multiforme, 45% of mesothelioma, 39% of esophageal adenocarcinoma, 32% of bladder urothelial carcinoma, 31% of skin cutaneous melanoma and head and neck squamous cell carcinoma, 28% of pancreatic adenocarcinoma, 27% of diffuse large B-cell lymphoma, 26% of lung squamous cell carcinoma, 17% of lung adenocarcinoma and cholangiocarcinoma, 15% of sarcoma, 11% of stomach adenocarcinoma and of brain lower grade glioma, 7% of adrenocortical carcinoma, 6% of liver hepatocellular carcinoma, 4% of breast invasive carcinoma, kidney renal papillary cell carcinoma and thymoma, 3% of ovarian serous cystadenocarcinoma and kidney renal clear cell carcinoma, and 2% of uterine carcinosarcoma and kidney chromophobe<sup>4,7</sup>. Alterations in CDKN2A are also observed in pediatric cancers<sup>7</sup>. Biallelic deletion of CDKN2A is observed in 68% of T-lymphoblastic leukemia/lymphoma, 40% of B-lymphoblastic leukemia/lymphoma, 25% of glioma, 19% of bone cancer, and 6% of embryonal tumors<sup>7</sup>. Somatic mutations in CDKN2A are observed in less that 1.5% of bone cancer (5 in 327 cases), B-lymphoblastic leukemia/lymphoma (3 in 252 cases), and leukemia (1 in 354 cases)<sup>7</sup>.

Potential relevance: Loss of CDKN2A can be useful in the diagnosis of mesothelioma, and mutations in CDKN2A are ancillary diagnostic markers of malignant peripheral nerve sheath tumors<sup>76,77,78</sup>. Additionally, deletion of CDKN2B is a molecular marker used in staging Grade 4 pediatric IDH-mutant astrocytoma<sup>79</sup>. Currently, no therapies are approved for CDKN2A aberrations. However, CDKN2A LOF leading to CDK4/6 activation may confer sensitivity to CDK inhibitors such as palbociclib and abemaciclib<sup>80,81,82</sup>. Alternatively, CDKN2A expression and Rb inactivation demonstrate resistance to palbociclib in cases of glioblastoma multiforme<sup>83</sup>. CDKN2A (p16) expression is associated with a favorable prognosis for progression-free survival (PFS) and overall survival (OS) in p16/HPV positive head and neck cancer<sup>84,85,86,87</sup>.

# **Biomarker Descriptions (continued)**

#### **CDKN2B** deletion

cyclin dependent kinase inhibitor 2B

<u>Background</u>: CDKN2B encodes cyclin dependent kinase inhibitor 2B, a cell cycle regulator that controls G1/S progression<sup>21,66</sup>. CDKN2B, also known as p15/INK4B, belongs to a family of INK4 cyclin-dependent kinase inhibitors, which also includes CDKN2A (p16/INK4A), CDKN2C (p18/INK4C), and CDKN2D (p19/INK4D)<sup>66</sup>. The INK4 family regulates cell cycle progression by inhibiting CDK4 or CDK6, thereby preventing the phosphorylation of Rb<sup>67,68,69</sup>. CDKN2B is a tumor suppressor and aberrations in this gene commonly co-occur with CDKN2A<sup>66</sup>. Germline mutations in CDKN2B are linked to pancreatic cancer predisposition and familial renal cell carcinoma<sup>21,88,89</sup>.

Alterations and prevalence: CDKN2B copy number loss is a frequently occurring somatic aberration that is observed in 55% of glioblastoma multiforme, 43% of mesothelioma, 35% of esophageal adenocarcinoma, 31% of bladder urothelial carcinoma, 29% of skin cutaneous melanoma, 28% of head and neck squamous cell carcinoma, 27% of pancreatic adenocarcinoma, 26% of lung squamous cell carcinoma, 25% of diffuse large B -cell lymphoma, 16% of lung adenocarcinoma, 15% of sarcoma, 14% of cholangiocarcinoma, 11% of stomach adenocarcinoma and brain lower grade glioma, 5% of liver hepatocellular carcinoma, 4% of adrenocortical carcinoma, breast invasive carcinoma, thymoma, and kidney renal papillary cell carcinoma, 3% of kidney renal clear cell carcinoma and ovarian serous cystadenocarcinoma, and 2% of uterine carcinosarcoma and kidney chromophobe<sup>4,7</sup>. Somatic mutations in CDKN2B are observed in 2% of uterine carcinosarcoma<sup>4,7</sup>. CDKN2B copy number loss is also observed in pediatric cancers, including 64% of childhood T-lymphoblastic leukemia/lymphoma, 37% of pediatric B-lymphoblastic leukemia/lymphoma, 25% of pediatric gliomas, 14% of pediatric bone cancers, 6% of embryonal tumors, and 2% of peripheral nervous system cancers<sup>4,7</sup>. Somatic mutations in CDKN2B are observed in less than 1% of bone cancer (1 in 327 cases)<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for CDKN2B aberrations. Homozygous deletion of CDKN2B is a molecular marker used in staging grade 4 pediatric IDH-mutant astrocytoma<sup>79</sup>.

#### **FANCM** deletion

FA complementation group M

Background: The FANCM gene encodes the FA complementation group M protein, a member of the Fanconi Anemia (FA) family, which also includes FANCA, FANCB, FANCC, FANCD1 (BRCA2), FANCD2, FANCE, FANCF, FANCG, FANCI, FANCJ (BRIP1), FANCL, and FANCN (PALB2)<sup>21</sup>. FA genes are tumor suppressors that are responsible for the maintenance of replication fork stability, DNA damage repair through the removal of interstrand cross-links (ICL), and subsequent initiation of the homologous recombination repair (HRR) pathway<sup>22,23</sup>. In response to DNA damage, FANCA, FANCB, FANCC, FANCE, FANCF, FANCG, FANCL, and FANCM assemble to form the FA core complex which is responsible for the monoubiquitination of the FANCI-FANCD2 (ID2) complex<sup>22</sup>. Monoubiquitination of the ID2 complex promotes co-localization with BRCA1/2, which is critical in BRCA mediated DNA repair<sup>24,25</sup>. Loss of function mutations in the FA family and HRR pathway can result in the BRCAness phenotype, characterized by a defect in the HRR pathway, mimicking BRCA1 or BRCA2 loss<sup>26,27</sup>. Germline mutations in FA genes lead to Fanconi Anemia, a condition characterized by chromosomal instability and congenital abnormalities, including bone marrow failure and cancer predisposition<sup>28,29</sup>.

Alterations and prevalence: Somatic mutations in FANCM are observed in 11% of uterine corpus endometrial carcinoma, 8% of skin cutaneous melanoma, 7% of lung adenocarcinoma, 6% of stomach adenocarcinoma, 5% colorectal adenocarcinoma, uterine carcinosarcoma, and bladder urothelial carcinoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for FANCM aberrations. Consistent with other genes that contribute to the BRCAness phenotype, mutations in FANCM are shown to confer enhanced sensitivity in vitro to PARP inhibitors such as olaparib<sup>30</sup>.

### **ERCC2** deletion

ERCC excision repair 2, TFIIH core complex helicase subunit

Background: The ERCC2 gene encodes ERCC excision repair 2, TFIIH core complex helicase subunit, also known as XPD<sup>21</sup>. ERCC2 is a protein involved in the nucleotide excision repair (NER) pathway responsible for repairing bulky DNA lesions caused by UV radiation, environmental mutagens, chemical agents, and cyclopurines generated by reactive oxygen species<sup>114</sup>. ERCC2 functions as a helicase along with ERCC3/XPB in the TFIIH core complex<sup>114</sup>. During repair of bulky lesions by NER, the TFIIH core complex binds to the lesion, followed by DNA damage verification by ERCC2, which is essential for NER<sup>114</sup>. Following lesion binding and verification, ERCC2 unwinds DNA in the 5'-3' direction<sup>114</sup>. Mutations in ERCC2 lead to stalled RNA polymerase, resulting in persistent block of transcription<sup>114</sup>. Germline ERCC2 mutations can lead to hereditary disorders including: Cockayne syndrome, characterized by skin cancer susceptibility and neurodegerneration; xeroderma pigmentosum (XP), characterized by neurodegeneration and developmental defects; and trichothiodystrophy (TTD), characterized by brittle hair due to sulfur deficiency as well as other developmental defects<sup>114,115</sup>.

# **Biomarker Descriptions (continued)**

Alterations and prevalence: Somatic mutations in ERCC2 are predominantly missense and occur in 9% of bladder urothelial carcinoma, 4% of skin cutaneous melanoma, 3% of uterine corpus endometrial carcinoma, stomach adenocarcinoma, and cholangiocarcinoma, and 2% of lung squamous cell carcinoma<sup>4,7</sup>. The missense mutation, N238S, is observed to be recurrent in bladder urothelial carcinoma and is predicted to result in ERCC2 loss of function<sup>4,7,116</sup>. Biallelic loss of ERCC2 is observed in 2% of brain lower grade glioma and diffuse large B-cell lymphoma, as well as 1% of sarcoma and ovarian serous cystadenocarcinoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for ERCC2 aberrations. In one study, ERCC2 mutations correlated with enhanced response to cisplatin based chemotherapy compared to wild-type ERCC2 in patients with muscle-invasive urothelial carcinoma<sup>117</sup>.

#### MAP2K4 p.(I73Rfs\*9) c.217\_218delAT

mitogen-activated protein kinase kinase 4

Background: The MAP2K4 gene encodes the mitogen-activated protein kinase kinase 4, also known as MEK4<sup>21</sup>. MAP2K4 is a member of the mitogen-activated protein kinase 2 (MAP2K) subfamily which also includes MAP2K1, MAP2K2, MAP2K3, MAP2K5, and MAP2K6<sup>100</sup>. Activation of MAPK proteins occurs through a kinase signaling cascade<sup>100,101,102</sup>. Specifically, MAP3Ks are responsible for phosphorylation of MAP2K family members<sup>100,101,102</sup>. Once activated, MAP2Ks are responsible for the phosphorylation of various MAPK proteins whose signaling is involved in several cellular processes including cell proliferation, differentiation, and inflammation<sup>100,101,102</sup>. Mutations observed in MAP2K4 were have been observed to impair kinase activity and promote tumorigenesis in vitro, supporting a possible tumor suppressor role for MAP2K4<sup>103</sup>.

Alterations and prevalence: Somatic mutations in MAP2K4 have been observed in 5% of uterine carcinoma and colorectal cancer, and 4% of breast invasive carcinoma<sup>4,7</sup>. Biallelic deletions have been observed in 3% of stomach cancer, and 2% of breast invasive carcinoma, diffuse large B-cell lymphoma (DLBCL), colorectal, pancreatic, and ovarian cancer<sup>4,7</sup>. Nonsense, frameshift, and missense mutations in MAP2K4 generally inactivate the kinase activity, and lost expression has been identified in prostate, ovarian, brain, and pancreatic cancer models<sup>104,105</sup>.

Potential relevance: Currently, no therapies are approved for MA2PK4 aberrations.

#### MLH3 deletion

mutL homolog 3

Background: The MLH3 gene encodes the mutL homolog 3 protein<sup>21</sup>. MLH3 heterodimerizes with MLH1 to form the MutLγ complex which functions as an endonuclease during meiosis, specifically in meiotic recombination<sup>41</sup>. MLH3 is considered a mismatch repair (MMR) gene due to its functional role in yeast, however, its exact MMR role in humans is less clear<sup>41,42,43</sup>. Low expression of MMR genes, including MLH3, have been associated with high levels of microsatellite instability (MSI-H) in colorectal cancer<sup>44</sup>.

Alterations and prevalence: Somatic mutations in MLH3 are observed in 9% of uterine corpus endometrial carcinoma, 4% of colorectal adenocarcinoma, skin cutaneous melanoma, and stomach adenocarcinoma<sup>4,7</sup>. Biallelic deletions are observed in 2% of kidney chromophobe<sup>4,7</sup>.

<u>Potential relevance:</u> Currently, no therapies are approved for MLH3 aberrations.

### Microsatellite stable

Background: Microsatellites are short tandem repeats (STR) of 1 to 6 bases of DNA between 5 to 50 repeat units in length. There are approximately 0.5 million STRs that occupy 3% of the human genome<sup>118</sup>. Microsatellite instability (MSI) is defined as a change in the length of a microsatellite in a tumor as compared to normal tissue<sup>119,120</sup>. MSI is closely tied to the status of the mismatch repair (MMR) genes. In humans, the core MMR genes include MLH1, MSH2, MSH6, and PMS2<sup>121</sup>. Mutations and loss of expression in MMR genes, known as defective MMR (dMMR), lead to MSI. In contrast, when MMR genes lack alterations, they are referred to as MMR proficient (pMMR). Consensus criteria were first described in 1998 and defined MSI-high (MSI-H) as instability in two or more of the following five markers: BAT25, BAT26, D5S346, D2S123, and D17S250<sup>122</sup>. Tumors with instability in one of the five markers were defined as MSI-low (MSI-L) whereas, those with instability in zero markers were defined as MS-stable (MSS)<sup>122</sup>. Tumors classified as MSI-L are often phenotypically indistinguishable from MSS tumors and tend to be grouped with MSS<sup>123,124,125,126,127</sup>. MSI-H is a hallmark of Lynch syndrome (LS), also known as hereditary non-polyposis colorectal cancer, which is caused by germline mutations in the MMR genes<sup>120</sup>. LS is associated with an increased risk of developing colorectal cancer, as well as other cancers, including endometrial and stomach cancer<sup>119,120,124,128</sup>.

# **Biomarker Descriptions (continued)**

Alterations and prevalence: The MSI-H phenotype is observed in 30% of uterine corpus endothelial carcinoma, 20% of stomach adenocarcinoma, 15-20% of colon adenocarcinoma, and 5-10% of rectal adenocarcinoma<sup>119,120,129,130</sup>. MSI-H is also observed in 5% of adrenal cortical carcinoma and at lower frequencies in other cancers such as esophageal, liver, and ovarian cancers<sup>129,130</sup>.

Potential relevance: Anti-PD-1 immune checkpoint inhibitors including pembrolizumab<sup>131</sup> (2014) and nivolumab<sup>132</sup> (2015) are approved for patients with MSI-H or dMMR colorectal cancer who have progressed following chemotherapy. Pembrolizumab<sup>131</sup> is also approved as a single agent, for the treatment of patients with advanced endometrial carcinoma that is MSI-H or dMMR with disease progression on prior therapy who are not candidates for surgery or radiation. Importantly, pembrolizumab is approved for the treatment of MSI-H or dMMR solid tumors that have progressed following treatment, with no alternative option and is the first anti-PD-1 inhibitor to be approved with a tumor agnostic indication<sup>131</sup>. Dostarlimab<sup>133</sup> (2021) is also approved for dMMR recurrent or advanced endometrial carcinoma or solid tumors that have progressed on prior treatment and is recommended as a subsequent therapy option in dMMR/MSI-H advanced or metastatic colon or rectal cancer<sup>125,134</sup>. The cytotoxic T-lymphocyte antigen 4 (CTLA-4) blocking antibody, ipilimumab<sup>135</sup> (2011), is approved alone or in combination with nivolumab in MSI-H or dMMR colorectal cancer that has progressed following treatment with chemotherapy. MSI-H may confer a favorable prognosis in colorectal cancer although outcomes vary depending on stage and tumor location<sup>125,136,137</sup>. Specifically, MSI-H is a strong prognostic indicator of better overall survival (OS) and relapse free survival (RFS) in stage II as compared to stage III colorectal cancer patients<sup>137</sup>. The majority of patients with tumors classified as either MSS or pMMR do not benefit from treatment with single-agent immune checkpoint inhibitors as compared to those with MSI-H tumors<sup>138,139</sup>. However, checkpoint blockade with the addition of chemotherapy or targeted therapies have demonstrated response in MSS or pMMR cancers<sup>138,139</sup>.

#### **RAD51B deletion**

RAD51 paralog B

Background: The RAD51B gene encodes the RAD51 paralog B protein, a member of the RAD51 recombinase family that also includes RAD51, RAD51C (RAD51L2), RAD51D (RAD51L3), XRCC2, and XRCC3 paralogs. The RAD51 family of proteins are involved in homologous recombination repair (HRR) and DNA repair of double-strand breaks (DSB)<sup>106</sup>. RAD51B associates with other RAD51 paralogs to form RAD51B-RAD51C-RAD51D-XRCC2 (BCDX2) complex<sup>107</sup>. The BCDX2 complex binds single- and double-stranded DNA to hydrolyze ATP<sup>108</sup>. RAD51B is a tumor suppressor gene. Loss of function mutations in RAD51B are implicated in the BRCAness phenotype, which is characterized by a defect in HRR mimicking BRCA1 or BRCA2 loss<sup>26,109</sup>. Biallelic expression of RAD51B is required for chromosomal integrity and haploinsufficiency leads to aberrant HRR resulting in centrosome fragmentation, aneuploidy, and mild hypersensitivity to DNA-damaging agents<sup>110</sup>. Genetic variation within the RAD51B locus on 14q24.1 is significantly associated with familial breast cancer risk<sup>111</sup>.

Alterations and prevalence: Somatic mutations in RAD51B are observed in up to 3% of uterine cancer<sup>4,7</sup>. Loss of function mutations in RAD51B are rare, but variation within the RAD51B locus is significantly associated with familial breast cancer risk<sup>111</sup>.

Potential relevance: The PARP inhibitor, olaparib<sup>112</sup> is approved (2020) for metastatic castration-resistant prostate cancer (mCRPC) with deleterious or suspected deleterious, germline or somatic mutations in HRR genes that includes RAD51B. In 2022, the FDA granted fast track designation to the small molecule inhibitor, pidnarulex<sup>113</sup>, for BRCA1/2, PALB2, or other homologous recombination deficiency (HRD) mutations in breast and ovarian cancers.

### RAD52 p.(S346\*) c.1037C>A

RAD52 homolog, DNA repair protein

<u>Background:</u> The RAD52 gene encodes the RAD52 homolog, DNA repair protein<sup>21</sup>. RAD52 binds to single- and double-stranded DNA and enables strand exchange for double-strand break (DSB) repair by binding to RAD51<sup>31</sup>. RAD52 also promotes DSB repair through homologous recombination repair (HRR) by recruiting BRCA1 to sites of DSBs, which leads to the removal of TP53BP1 and prevents DSB repair by non-homologous end joining (NHEJ)<sup>32</sup>.

Alterations and prevalence: Somatic mutations in RAD52 are observed in 2% of uterine corpus endometrial carcinoma, uterine carcinosarcoma, and skin cutaneous melanoma<sup>4,7</sup>.

<u>Potential relevance:</u> Currently, no therapies are approved for RAD52 aberrations.

### XRCC3 deletion

X-ray repair cross complementing 3

Background: The XRCC3 gene encodes the X-ray cross complementing 3 protein, a member of the RAD51 recombinase family that also includes RAD51, RAD51C, RAD51D, and XRCC2 paralogs<sup>21,33</sup>. XRCC3 complexes with RAD51C to form the CX3 complex, which

# **Biomarker Descriptions (continued)**

functions in strand exchange and Holliday junction resolution during homologous recombination repair (HRR)<sup>33,34</sup>. XRCC3 may complex with BRCA2, FANCD2, and FANCG to maintain chromosome stability<sup>35</sup>.

Alterations and prevalence: Somatic mutations in XRCC3 are observed in 1% of uveal melanoma, colorectal adenocarcinoma, and cervical squamous cell carcinoma<sup>4,7</sup>. Biallelic deletions in XRCC3 are observed in 3% of cholangiocarcinoma and 2% of diffuse large B-cell lymphoma (DLBCL) and bladder urothelial carcinoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for XRCC3 aberrations. Pre-clinical evidence suggests that XRCC3 mutations may demonstrate sensitivity to cisplatin<sup>35</sup>.

#### **NOTCH2** deletion

notch 2

Background: The NOTCH2 gene encodes the notch receptor 2 protein, a type 1 transmembrane protein and member of the NOTCH family of genes, which also includes NOTCH1, NOTCH3, and NOTCH4. NOTCH proteins contain multiple epidermal growth factor (EGF)-like repeats in their extracellular domain, which are responsible for ligand binding and homodimerization, thereby promoting NOTCH signaling<sup>50</sup>. Following ligand binding, the NOTCH intracellular domain is released, which activates the transcription of several genes involved in regulation of cell proliferation, differentiation, growth, and metabolism<sup>51,52</sup>. In cancer, depending on the tumor type, aberrations in the NOTCH family can be gain of function or loss of function suggesting both oncogenic and tumor suppressor roles for NOTCH family members<sup>53,54,55,56</sup>.

Alterations and prevalence: Somatic mutations observed in NOTCH2 are primarily missense or truncating and are found in about 11% of uterine cancer, 6% of melanoma and stomach cancer, as well as 3-5% diffuse large B-cell lymphoma (DLBCL), lung, colorectal, bladder, cervical, and head and neck cancers<sup>4</sup>.

Potential relevance: Currently, no therapies are approved for NOTCH2 aberrations.

#### **HLA-A deletion**

major histocompatibility complex, class I, A

Background: The HLA-A gene encodes the major histocompatibility complex, class I,  $A^{21}$ . MHC (major histocompatibility complex) class I molecules are located on the cell surface of nucleated cells and present antigens from within the cell for recognition by cytotoxic T cells<sup>90</sup>. MHC class I molecules are heterodimers composed of two polypeptide chains, α and B2M<sup>91</sup>. The classical MHC class I genes include HLA-A, HLA-B, and HLA-C and encode the α polypeptide chains, which present short polypeptide chains, of 7 to 11 amino acids, to the immune system to distinguish self from non-self<sup>92,93,94</sup>. Downregulation of MHC class I promotes tumor evasion of the immune system, suggesting a tumor suppressor role for HLA-A<sup>95</sup>.

<u>Alterations and prevalence</u>: Somatic mutations in HLA-A are observed in 7% of diffuse large B-cell lymphoma (DLBCL), 4% of cervical squamous cell carcinoma and head and neck squamous cell carcinoma, 3% of colorectal adenocarcinoma, and 2% of uterine corpus endometrial carcinoma and stomach adenocarcinoma<sup>4,7</sup>. Biallelic loss of HLA-A is observed in 4% of DLBCL<sup>4,7</sup>.

<u>Potential relevance:</u> Currently, no therapies are approved for HLA-A aberrations.

#### **HLA-B** deletion

major histocompatibility complex, class I, B

Background: The HLA-B gene encodes the major histocompatibility complex, class I, B21. MHC (major histocompatibility complex) class I molecules are located on the cell surface of nucleated cells and present antigens from within the cell for recognition by cytotoxic T cells90. MHC class I molecules are heterodimers composed of two polypeptide chains, α and B2M91. The classical MHC class I genes include HLA-A, HLA-B, and HLA-C and encode the α polypeptide chains, which present short polypeptide chains, of 7 to 11 amino acids, to the immune system to distinguish self from non-self92,93,94. Downregulation of MHC class I promotes tumor evasion of the immune system, suggesting a tumor suppressor role for HLA-B95.

Alterations and prevalence: Somatic mutations in HLA-B are observed in 10% of diffuse large B-cell lymphoma (DLBCL), 5% of cervical squamous cell carcinoma and stomach adenocarcinoma, 4% of head and neck squamous cell carcinoma and colorectal adenocarcinoma, 3% of uterine cancer, and 2% of esophageal adenocarcinoma and skin cutaneous melanoma<sup>4,7</sup>. Biallelic loss of HLA-B is observed in 5% of DLBCL<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for HLA-B aberrations.

# **Biomarker Descriptions (continued)**

MAX p.(R60L) c.179G>T

MYC associated factor X

Background: The MAX gene encodes the MYC associated factor X protein, a member of the basic helix-loop-helix leucine zipper (bHLHZ) transcription factor family, which also includes MNT and MXD1-496. MAX is ubiquitously expressed as two common isoforms, p21 and p22, each of which have unique DNA binding and biological activities96. MAX heterodimerizes with bHLHZ transcription factors including MYC, MAD, MNT, and MGA to form complexes that act on DNA sequences to regulate the transcription of target genes involved in cell growth and proliferation96,97. Homozygous alterations involving MAX lead to a protein incapable of dimerization and repressing transcription98. Germline mutations in MAX are observed in hereditary pheochromocytoma, a rare neural crest cell tumor98.

Alterations and prevalence: Somatic mutations in MAX are observed in 4% of uterine cancer, and 1-1.5% of lung adenocarcinoma, colorectal cancer, and clear cell renal cell carcinoma<sup>4,7</sup>. The missense mutation R60Q has been observed to be recurrent in a variety of cancers<sup>4,7,99</sup>. This mutation falls within the bHLH domain and has been observed to alter the DNA binding properties of MAX<sup>99</sup>. Amplifications are observed 2% of diffuse large B-cell lymphoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for MAX aberrations.

#### **DICER1** deletion

dicer 1, ribonuclease III

Background: The DICER1 gene encodes the dicer 1, ribonuclease III protein<sup>21</sup>. DICER1 is a member of the ribonuclease (RNase) III family that also includes DROSHA<sup>60</sup>. Both DICER and DROSHA are responsible for the processing of precursor non-coding RNA (primary miRNA) into micro-RNA (miRNA)<sup>60,61</sup>. Following primary miRNA processing to hairpin precursor miRNA (pre-miRNA) by DROSHA and DGCR8, pre-miRNA is then cleaved by DICER1 resulting in the production of mature miRNA<sup>60</sup>. Once processed, mature miRNA is capable of post-transcriptional gene repression by recognizing complimentary target sites on messenger RNA (mRNA)<sup>60,61</sup>. miRNAs are frequently dysregulated in cancer, potentially through DGCR8, DICER1, or DROSHA aberrations that impact miRNA processing<sup>61,62,63,64</sup>. Germline DICER1 mutations result in DICER1 syndrome, a rare genetic disorder that predisposes affected individuals to tumor development<sup>65</sup>.

Alterations and prevalence: Somatic mutations in DICER1 are observed in 13% of uterine corpus endometrial carcinoma, 11% of skin cutaneous melanoma, and 4% of colorectal adenocarcinoma, bladder urothelial carcinoma, and uterine carcinosarcoma<sup>4,7</sup>. Biallelic loss of DICER1 is observed in 3% of cholangiocarcinoma and 2% kidney chromophobe<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for DICER1 aberrations.

#### **NCOR1** deletion

nuclear receptor corepressor 1

Background: NCOR1 encodes nuclear receptor corepressor 1, which serves as a scaffold protein for large corepressor including transducin beta like 1 X-linked (TBL1X/Y related 1 (TBL1XR1), the G-protein-pathway suppressor 2 (GPS2), and protein deacetylases such as histone deacetylase 3 (HDAC3)<sup>21,36,37</sup>. NCOR1 plays a key role in several processes including embryonal development, metabolism, glucose homeostasis, inflammation, cell fate, chromatin structure and genomic stability<sup>36,37,38,39</sup>. NCOR1 has been shown exhibit a tumor suppressor role by inhibiting invasion and metastasis in various cancer models<sup>37</sup>. Inactivation of NCOR1 through mutation or deletion is observed in several cancer types including colorectal cancer, bladder cancer, hepatocellular carcinomas, lung cancer, and breast cancer<sup>37,40</sup>.

Alterations and prevalence: Somatic mutations in NCOR1 are observed in 13% of uterine corpus endometrial carcinoma, 11% of skin cutaneous melanoma, 8% of bladder urothelial carcinoma, 7% of stomach adenocarcinoma, 6% of colorectal adenocarcinoma, 5% of lung squamous cell carcinoma and breast invasive carcinoma, 4% of cervical squamous cell carcinoma and lung adenocarcinoma, 3% of mesothelioma, head and neck squamous cell carcinoma, cholangiocarcinoma, and kidney renal papillary cell carcinoma, and 2% of esophageal adenocarcinoma, glioblastoma multiforme, and ovarian serous cystadenocarcinoma<sup>4,7</sup>. Biallelic loss of NCOR1 are observed in 3% of liver hepatocellular carcinoma, and 2% of uterine carcinosarcoma, stomach adenocarcinoma, diffuse large B-cell lymphoma, and bladder urothelial carcinoma<sup>4,7</sup>. Structural variants of NCOR1 are observed in 3% of cholangiocarcinoma and 2% of uterine carcinosarcoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for NCOR1 aberrations.

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# **Biomarker Descriptions (continued)**

#### **ARHGAP35** deletion

Rho GTPase activating protein 35

Background: ARHGAP35 encodes Rho GTPase activating protein 35, human glucocorticoid receptor DNA binding factor. ARHGAP35 functions as a repressor of glucocorticoid receptor transcription<sup>21</sup>. Rho GTPases regulate various cellular processes such as cell adhesion, cell migration and play a critical role in metastasis through the negative regulation of RhoA which is localized to the cell membrane<sup>45,46</sup>. Aberrations in ARHGAP35, including mutations, have been observed to result in both loss and gain of function thereby promoting tumor growth and metastasis<sup>47,48</sup>.

Alterations and prevalence: Somatic mutations of AHGAP35 are observed in 20% of uterine corpus endometrial carcinoma, 11% of uterine carcinosarcoma, 6% of skin cutaneous melanoma, bladder urothelial carcinoma, and lung squamous cell carcinoma, 5% of colorectal adenocarcinoma, and 4% of stomach adenocarcinoma and lung adenocarcinoma<sup>4,7</sup>. In endometrial cancer, R997\* has been observed to be recurrent and has been observed to confer loss of RhoGAP activity due to protein truncation and loss of its RhoGAP domain<sup>49</sup>. Amplification of AHGAP35 is observed in 4% of uterine carcinosarcoma, 2% of adrenocortical carcinoma, and diffuse large B-cell lymphoma<sup>4,7</sup>. Biallelic loss of AHGAP35 has been observed in 2% of sarcoma<sup>4,7</sup>.

Potential relevance: Currently, no therapies are approved for ARHGAP35 aberrations.

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### **Alerts Informed By Public Data Sources**

### **Current FDA Information**

Contraindicated

Not recommended



Resistance



Breakthrough



FDA information is current as of 2025-05-14. For the most up-to-date information, search www.fda.gov.

### KRAS p.(G12C) c.34G>T



### panitumumab, panitumumab + sotorasib

Cancer type: Colorectal Cancer

Label as of: 2025-01-16

Variant class: KRAS G12C mutation

#### Indications and usage:

VECTIBIX® is an epidermal growth factor receptor (EGFR) antagonist indicated for the treatment of:

Adult patients with wild-type RAS (defined as wild-type in both KRAS and NRAS as determined by an FDA-approved test) Metastatic Colorectal Cancer (mCRC)\*:

- In combination with FOLFOX for first-line treatment.
- As monotherapy following disease progression after prior treatment with fluoropyrimidine, oxaliplatin, and irinotecancontaining chemotherapy.

KRAS G12C-mutated Metastatic Colorectal Cancer (mCRC)\*

In combination with sotorasib, for the treatment of adult patients with KRAS G12C-mutated mCRC, as determined by an FDAapproved test, who have received prior treatment with fluoropyrimidine-, oxaliplatin-, and irinotecan-based chemotherapy.

\*Limitations of Use: VECTIBIX® is not indicated for the treatment of patients with RAS-mutant mCRC unless used in combination with sotorasib in KRAS G12C-mutated mCRC. VECTIBIX® is not indicated for the treatment of patients with mCRC for whom RAS mutation status is unknown.

#### Reference:

https://www.accessdata.fda.gov/drugsatfda\_docs/label/2025/125147s213lbl.pdf

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# KRAS p.(G12C) c.34G>T (continued)

### cetuximab

Cancer type: Colorectal Cancer Label as of: 2021-09-24 Variant class: KRAS G12 mutation

#### Indications and usage:

Erbitux® is an epidermal growth factor receptor (EGFR) antagonist indicated for treatment of:

Head and Neck Cancer

- Locally or regionally advanced squamous cell carcinoma of the head and neck in combination with radiation therapy.
- Recurrent locoregional disease or metastatic squamous cell carcinoma of the head and neck in combination with platinumbased therapy with fluorouracil.
- Recurrent or metastatic squamous cell carcinoma of the head and neck progressing after platinum-based therapy.

#### Colorectal Cancer

K-Ras wild-type, EGFR-expressing, metastatic colorectal cancer as determined by FDA-approved test

- in combination with FOLFIRI for first-line treatment,
- in combination with irinotecan in patients who are refractory to irinotecan-based chemotherapy,
- as a single agent in patients who have failed oxaliplatin- and irinotecan-based chemotherapy or who are intolerant to irinotecan.

Limitations of Use: Erbitux® is not indicated for treatment of Ras-mutant colorectal cancer or when the results of the Ras mutation tests are unknown.

BRAF V600E Mutation-Positive Metastatic Colorectal Cancer (CRC)

 in combination with encorafenib, for the treatment of adult patients with metastatic colorectal cancer (CRC) with a BRAF V600E mutation, as detected by an FDA-approved test, after prior therapy.

#### Reference:

https://www.accessdata.fda.gov/drugsatfda\_docs/label/2021/125084s279lbl.pdf

### 

Cancer type: Non-Small Cell Lung Cancer Variant class: KRAS G12C mutation

### **Supporting Statement:**

The FDA has granted Breakthrough Therapy designation to KRAS G12C inhibitor, GDC-6036, for KRAS G12C mutation in non-small cell lung cancer.

#### Reference:

https://assets.cwp.roche.com/f/126832/x/5738a7538b/irp230202.pdf

### avutometinib + sotorasib

Cancer type: Non-Small Cell Lung Cancer Variant class: KRAS G12C mutation

#### Supporting Statement:

The FDA has granted Fast Track designation to Verastem Oncology's investigational RAF/MEK clamp, avutometinib, in combination with Amgen's KRAS G12C inhibitor, LUMAKRASTM (sotorasib), for the treatment of patients with KRAS G12C-mutant metastatic non-small cell lung cancer (NSCLC) who have received at least one prior systemic therapy and have not been previously treated with a KRAS G12C inhibitor.

#### Reference:

https://investor.verastem.com/news-releases/news-release-details/verastem-oncology-granted-fast-track-designation-combination

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# KRAS p.(G12C) c.34G>T (continued)

### **ு №** BBO-8520

Cancer type: Non-Small Cell Lung Cancer Variant class: KRAS G12C mutation

### Supporting Statement:

The FDA has granted Fast Track designation to the KRAS G12C inhibitor, BBO-8520, for the treatment of adult patients with previously treated, KRAS<sup>G12C</sup>-mutated metastatic non-small cell lung cancer (NSCLC).

#### Reference:

https://www.businesswire.com/news/home/20250109170439/en/

#### ♣ BBP-398 + sotorasib

**Cancer type:** Non-Small Cell Lung Cancer, Variant class: KRAS G12C mutation Solid Tumor

#### Supporting Statement:

The FDA has granted Fast Track designation to a SHP2 inhibitor, BBP-398, in combination with LUMAKRAS® for adult patients with previously treated KRAS G12C-mutated metastatic NSCLC.

#### Reference:

https://bridgebio.com/news/bridgebio-pharma-announces-first-lung-cancer-patient-dosed-in-phase-1-2-trial-and-us-fda-fast-track-designation-for-shp2-inhibitor-bbp-398-in-combination-with-amgens-lumakras-sotorasib/

#### # D3S-001

Cancer type: Colorectal Cancer Variant class: KRAS G12C mutation

#### **Supporting Statement:**

The FDA has granted Fast Track designation to the KRAS G12C inhibitor, D3S-001, for the treatment of KRAS G12C mutated patients with advanced unresectable or metastatic colorectal cancers.

The FDA has also granted Fast Track designation to D3S-001, for the treatment of late-line non-small cell lung cancer (NSCLC) and colorectal cancer (CRC).

#### Reference:

https://www.d3bio.com/press-releases/d3-bios-d3s-001-receives-u-s-fda-fast-track-designation-for-the-treatment-of-colorectal-cancer-with-kras-g12c-mutation

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#### **Current NCCN Information**

Contraindicated

Not recommended

Resistance

Breakthrough

Fast Track

NCCN information is current as of 2025-05-01. To view the most recent and complete version of the guideline, go online to NCCN.org.

For NCCN International Adaptations & Translations, search www.nccn.org/global/what-we-do/international-adaptations.

Some variant specific evidence in this report may be associated with a broader set of alterations from the NCCN Guidelines. Specific variants listed in this report were sourced from approved therapies or scientific literature. These therapeutic options are appropriate for certain population segments with cancer. Refer to the NCCN Guidelines® for full recommendation.

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## KRAS p.(G12C) c.34G>T

### cetuximab

Cancer type: Colon Cancer Variant class: KRAS G12 mutation

#### Summary:

NCCN Guidelines® include the following supporting statement(s):

■ "Patients with any known KRAS mutation (exon 2, 3, 4) or NRAS mutation (exon 2, 3, 4) should not be treated with either cetuximab or panitumumab, unless given as part of a regimen targeting a KRAS G12C mutation."

Reference: NCCN Guidelines® - NCCN-Colon Cancer [Version 3.2025]

### cetuximab

Cancer type: Rectal Cancer Variant class: KRAS G12 mutation

### Summary:

NCCN Guidelines® include the following supporting statement(s):

"Patients with any known KRAS mutation (exons 2, 3, and 4) or NRAS mutation (exons 2, 3, and 4) should not be treated with either cetuximab or panitumumab, unless given as part of a regimen targeting a KRAS G12C mutation."

Reference: NCCN Guidelines® - NCCN-Rectal Cancer [Version 2.2025]

### panitumumab

Cancer type: Colon Cancer Variant class: KRAS G12 mutation

### Summary:

NCCN Guidelines® include the following supporting statement(s):

■ "Patients with any known KRAS mutation (exon 2, 3, 4) or NRAS mutation (exon 2, 3, 4) should not be treated with either cetuximab or panitumumab, unless given as part of a regimen targeting a KRAS G12C mutation."

Reference: NCCN Guidelines® - NCCN-Colon Cancer [Version 3.2025]

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# KRAS p.(G12C) c.34G>T (continued)

### panitumumab

Cancer type: Rectal Cancer Variant class: KRAS G12 mutation

Summary:

NCCN Guidelines® include the following supporting statement(s):

■ "Patients with any known KRAS mutation (exons 2, 3, and 4) or NRAS mutation (exons 2, 3, and 4) should not be treated with either cetuximab or panitumumab, unless given as part of a regimen targeting a KRAS G12C mutation."

Reference: NCCN Guidelines® - NCCN-Rectal Cancer [Version 2.2025]

#### **Current EMA Information**

EMA information is current as of 2025-05-14. For the most up-to-date information, search www.ema.europa.eu.

### KRAS p.(G12C) c.34G>T

cetuximab, cetuximab + oxaliplatin

Cancer type: Colorectal Cancer Label as of: 2025-01-16 Variant class: KRAS G12 mutation

Reference:

https://www.ema.europa.eu/en/documents/product-information/erbitux-epar-product-information\_en.pdf

panitumumab + oxaliplatin

Cancer type: Colorectal Cancer Label as of: 2025-05-07 Variant class: KRAS G12 mutation

Reference:

https://www.ema.europa.eu/en/documents/product-information/vectibix-epar-product-information\_en.pdf

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#### **Current ESMO Information**

Contraindicated

Not recommended

Resistance

Breakthrough

Fast Track

ESMO information is current as of 2025-05-01. For the most up-to-date information, search www.esmo.org.

### KRAS p.(G12C) c.34G>T

### cetuximab

Cancer type: Colorectal Cancer Variant class: KRAS G12 mutation

#### Summary:

ESMO Clinical Practice Guidelines include the following supporting statement:

- "The presence of RAS mutations is associated with resistance to anti-EGFR mAbs and knowing the expanded RAS mutational status is mandatory for use of both cetuximab and panitumumab, avoiding anti-EGFR mAb treatment when a RAS mutation is confirmed."
- "RAS testing is mandatory before treatment with anti-EGFR mAbs and can be carried out on either the primary tumor or other metastatic sites [III, A]".

Reference: ESMO Clinical Practice Guidelines - ESMO-Metastatic Colorectal Cancer [Ann Oncol (2023); https://doi.org/10.1016/j.annonc.2022.10.003 (published)]

### panitumumab

Cancer type: Colorectal Cancer Variant class: KRAS G12 mutation

#### Summary:

ESMO Clinical Practice Guidelines include the following supporting statement:

- "The presence of RAS mutations is associated with resistance to anti-EGFR mAbs and knowing the expanded RAS mutational status is mandatory for use of both cetuximab and panitumumab, avoiding anti-EGFR mAb treatment when a RAS mutation is confirmed"
- "RAS testing is mandatory before treatment with anti-EGFR mAbs and can be carried out on either the primary tumor or other metastatic sites [III, A]".

Reference: ESMO Clinical Practice Guidelines - ESMO-Metastatic Colorectal Cancer [Ann Oncol (2023); https://doi.org/10.1016/j.annonc.2022.10.003 (published)]

### **Genes Assayed**

### Genes Assayed for the Detection of DNA Sequence Variants

ABL1, ABL2, ACVR1, AKT1, AKT2, AKT3, ALK, AR, ARAF, ATP1A1, AURKA, AURKB, AURKC, AXL, BCL2, BCL2L12, BCL6, BCR, BMP5, BRAF, BTK, CACNA1D, CARD11, CBL, CCND1, CCND2, CCND3, CCNE1, CD79B, CDK4, CDK6, CHD4, CSF1R, CTNNB1, CUL1, CYSLTR2, DDR2, DGCR8, DROSHA, E2F1, EGFR, EIF1AX, EPAS1, ERBB2, ERBB3, ERBB4, ESR1, EZH2, FAM135B, FGF7, FGFR1, FGFR2, FGFR3, FGFR4, FLT3, FLT4, FOXA1, FOXL2, FOXO1, GATA2, GLI1, GNA11, GNAQ, GNAS, HIF1A, HRAS, IDH1, IDH2, IKBKB, IL6ST, IL7R, IRF4, IRS4, KCNJ5, KDR, KIT, KLF4, KLF5, KNSTRN, KRAS, MAGOH, MAP2K1, MAP2K2, MAPK1, MAX, MDM4, MECOM, MED12, MEF2B, MET, MITF, MPL, MTOR, MYC, MYCN, MYD88, MYOD1, NFE2L2, NRAS, NSD2, NT5C2, NTRK1, NTRK2, NTRK3, NUP93, PAX5, PCBP1, PDGFRA, PDGFRB, PIK3C2B, PIK3CA, PIK3CB, PICTOR, RIT1, ROS1, RPL10, SETBP1, SF3B1, SIX1, SIX2, SLCO1B3, SMC1A, SMO, SNCAIP, SOS1, SOX2, SPOP, SRC, SRSF2, STAT3, STAT5B, STAT6, TAF1, TERT, TGFBR1, TOP1, TOP2A, TPMT, TRRAP, TSHR, U2AF1, USP8, WAS, XPO1, ZNF217, ZNF429

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### **Genes Assayed (continued)**

### Genes Assayed for the Detection of Copy Number Variations

ABCB1, ABL1, ABL2, ABRAXAS1, ACVR1B, ACVR2A, ADAMTS12, ADAMTS2, AKT1, AKT2, AKT3, ALK, AMER1, APC, AR, ARAF, ARHGAP35, ARID1A, ARID1B, ARID2, ARID5B, ASXL1, ASXL2, ATM, ATR, ATRX, AURKA, AURKC, AXIN1, AXIN2, AXL, B2M, BAP1, BARD1, BCL2, BCL2L12, BCL6, BCOR, BLM, BMPR2, BRAF, BRCA1, BRCA2, BRIP1, CARD11, CASP8, CBFB, CBL, CCND1, CCND2, CCND3, CCNE1, CD274, CD276, CDC73, CDH1, CDH10, CDK12, CDK4, CDK6, CDKN1A, CDKN1B, CDKN2A, CDKN2B, CDKN2C, CHD4, CHEK1, CHEK2, CIC, CREBBP, CSMD3, CTCF, CTLA4, CTNND2, CUL3, CUL4A, CUL4B, CYLD, CYP2C9, DAXX, DDR1, DDR2, DDX3X, DICER1, DNMT3A, DOCK3, DPYD, DSC1, DSC3, EGFR, EIF1AX, ELF3, EMSY, ENO1, EP300, EPCAM, EPHA2, ERAP1, ERAP2, ERBB2, ERBB3, ERBB4, ERCC2, ERCC4, ERRFI1, ESR1, ETV6, EZH2, FAM135B, FANCA, FANCC, FANCD2, FANCE, FANCF, FANCG, FANCI, FANCI, FANCM, FAT1, FBXW7, FGF19, FGF23, FGF4, FGF9, FGFR1, FGFR2, FGFR3, FGFR4, FLT3, FLT4, FOXA1, FUBP1, FYN, GATA2, GATA3, GLI3, GNA13, GNAS, GPS2, HDAC2, HDAC9, HLA-A, HLA-B, HNF1A, IDH2, IGF1R, IKBKB, IL7R, INPP4B, JAK1, JAK2, JAK3, KDM5C, KDM6A, KDR, KEAP1, KIT, KLF5, KMT2A, KMT2B, KMT2C, KMT2D, KRAS, LARP4B, LATS1, LATS2, MAGOH, MAP2K1, MAP2K4, MAP2K7, MAP3K1, MAP3K4, MAPK1, MAPK8, MAX, MCL1, MDM2, MDM4, MECOM, MEF2B, MEN1, MET, MGA, MITF, MLH1, MLH3, MPL, MRE11, MSH2, MSH3, MSH6, MTAP, MTOR, MUTYH, MYC, MYCL, MYCN, MYD88, NBN, NCOR1, NF1, NF2, NFE2L2, NOTCH1, NOTCH2, NOTCH3, NOTCH4, NRAS, NTRK1, NTRK3, PALB2, PARP1, PARP2, PARP3, PARP4, PBRM1, PCBP1, PDCD1, PDCD1LG2, PDGFRA, PDGFRB, PDIA3, PGD, PHF6, PIK3C2B, PIK3CA, PIK3CB, PIK3R1, PIK3R2, PIM1, PLCG1, PMS1, PMS2, POLD1, POLE, POT1, PPM1D, PPP2R1A, PPP2R2A, PPP6C, PRDM1, PRDM9, PRKACA, PRKAR1A, PTCH1, PTEN, PTPN11, PTPRT, PXDNL, RAC1, RAD50, RAD51, RAD51B, RAD51C, RAD51D, RAD52, RAD54L, RAF1, RARA, RASA1, RASA2, RB1, RBM10, RECQL4, RET, RHEB, RICTOR, RIT1, RNASEH2A, RNASEH2B, RNF43, ROS1, RPA1, RPS6KB1, RPTOR, RUNX1, SDHA, SDHB, SDHD, SETBP1, SETD2, SF3B1, SLCO1B3, SLX4, SMAD2, SMAD4, SMARCA4, SMARCB1, SMC1A, SMO, SOX9, SPEN, SPOP, SRC, STAG2, STAT3, STAT6, STK11, SUFU, TAP1, TAP2, TBX3, TCF7L2, TERT, TET2, TGFBR2, TNFAIP3, TNFRSF14, TOP1, TP53, TP63, TPMT, TPP2, TSC1, TSC2, U2AF1, USP8, USP9X, VHL, WT1, XPO1, XRCC2, XRCC3, YAP1, YES1, ZFHX3, ZMYM3, ZNF217, ZNF429, ZRSR2

### Genes Assayed for the Detection of Fusions

AKT2, ALK, AR, AXL, BRAF, BRCA1, BRCA2, CDKN2A, EGFR, ERBB2, ERBB4, ERG, ESR1, ETV1, ETV4, ETV5, FGFR1, FGFR2, FGR3, FGR, FLT3, JAK2, KRAS, MDM4, MET, MYB, MYBL1, NF1, NOTCH1, NOTCH4, NRG1, NTRK1, NTRK2, NTRK3, NUTM1, PDGFRA, PDGFRB, PIK3CA, PPARG, PRKACA, PRKACB, PTEN, RAD51B, RAF1, RB1, RELA, RET, ROS1, RSPO2, RSPO3, TERT

### Genes Assayed with Full Exon Coverage

ABRAXAS1, ACVR1B, ACVR2A, ADAMTS12, ADAMTS2, AMER1, APC, ARHGAP35, ARID1A, ARID1B, ARID2, ARID5B, ASXL1, ASXL2, ATM, ATR, ATRX, AXIN1, AXIN2, B2M, BAP1, BARD1, BCOR, BLM, BMPR2, BRCA1, BRCA2, BRIP1, CALR, CASP8, CBFB, CD274, CD276, CDC73, CDH1, CDH10, CDK12, CDKN1A, CDKN1B, CDKN2A, CDKN2B, CDKN2C, CHEK1, CHEK2, CIC, CIITA, CREBBP, CSMD3, CTCF, CTLA4, CUL3, CUL4A, CUL4B, CYLD, CYP2C9, CYP2D6, DAXX, DDX3X, DICER1, DNMT3A, DOCK3, DPYD, DSC1, DSC3, ELF3, ENO1, EP300, EPCAM, EPHA2, ERAP1, ERAP2, ERCC2, ERCC4, ERCC5, ERRF11, ETV6, FANCA, FANCC, FANCD2, FANCE, FANCF, FANCG, FANCI, FANCI, FANCM, FAS, FAT1, FBXW7, FUBP1, GATA3, GNA13, GPS2, HDAC2, HDAC9, HLA-A, HLA-B, HNF1A, ID3, INPP4B, JAK1, JAK2, JAK3, KDM5C, KDM6A, KEAP1, KLHL13, KMT2A, KMT2B, KMT2C, KMT2D, LARP4B, LATS1, LATS2, MAP2K4, MAP2K7, MAP3K1, MAP3K4, MAPK8, MEN1, MGA, MLH1, MLH3, MRE11, MSH2, MSH3, MSH6, MTAP, MTUS2, MUTYH, NBN, NCOR1, NF1, NF2, NOTCH1, NOTCH2, NOTCH3, NOTCH4, PALB2, PARP1, PARP2, PARP3, PARP4, PBRM1, PDCD1, PDCD1LG2, PDIA3, PGD, PHF6, PIK3R1, PMS1, PMS2, POLD1, POLE, POT1, PPM1D, PPP2R2A, PRDM1, PRDM9, PRKAR1A, PSMB10, PSMB8, PSMB9, PTCH1, PTEN, PTPRT, RAD50, RAD51, RAD51B, RAD51C, RAD51D, RAD52, RAD54L, RASA1, RASA2, RB1, RBM10, RECQL4, RNASEH2A, RNASEH2B, RNASEH2C, RNF43, RPA1, RPL22, RPL5, RUNX1, RUNX1T1, SDHA, SDHB, SDHC, SDHD, SETD2, SLX4, SMAD2, SMAD4, SMARCA4, SMARCB1, SOCS1, SOX9, SPEN, STAG2, STAT1, STK11, SUFU, TAP1, TAP2, TBX3, TCF7L2, TET2, TGFBR2, TMEM132D, TNFAIP3, TNFRSF14, TP53, TP63, TPP2, TSC1, TSC2, UGT1A1, USP9X, VHL, WT1, XRCC2, XRCC3, ZBTB20, ZFHX3, ZMYM3, ZRSR2

# **Relevant Therapy Summary**

■ In this cancer type
O In other cancer type
O In this cancer type and other cancer types
X No evidence

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
adagrasib	•	0	•		<b>(II)</b>
sotorasib	•	0	•	•	<b>(II)</b>
adagrasib + cetuximab	0	0	×	×	×
panitumumab + sotorasib	A	0	×	×	×
panitumumab	A	×	×	×	×
adagrasib + panitumumab	×	0	×	×	×
cetuximab + sotorasib	×	0	×	×	×
bevacizumab + CAPOX	×	×	×	0	×
bevacizumab + FOLFIRI	×	×	×	0	×
bevacizumab + FOLFOX	×	×	×	0	×
bevacizumab + FOLFOXIRI	×	×	×	0	×
adagrasib, pembrolizumab, chemotherapy	×	×	×	×	<b>(III)</b>
D-1553	×	×	×	×	<b>(III)</b>
divarasib, sotorasib, adagrasib	×	×	×	×	<b>(III)</b>
glecirasib, JAB-3312, tislelizumab, chemotherapy	×	×	×	×	<b>(III)</b>
MK-1084, pembrolizumab	×	×	×	×	<b>(III)</b>
olomorasib, durvalumab	×	×	×	×	<b>(III)</b>
olomorasib, pembrolizumab, chemotherapy	×	×	×	×	<b>(III)</b>
RMC-6236	×	×	×	×	<b>(III)</b>
sotorasib, pembrolizumab, chemotherapy	×	×	×	×	<b>(III)</b>
adagrasib, pembrolizumab	×	×	×	×	(II/III)
adagrasib, radiation therapy	×	×	×	×	(II)
daratumumab, TG-01 (Targovax), QS-21 Stimulon, nivolumab	×	×	×	×	<b>(II)</b>
divarasib	×	×	×	×	(II)
glecirasib	×	×	×	×	<b>(II)</b>
regorafenib	×	×	×	×	<b>(II)</b>
sintilimab, catequentinib	×	×	×	×	<b>(II)</b>
sotorasib, chemotherapy	×	×	×	×	<b>(II)</b>
sotorasib, chemotherapy, bevacizumab (Allergan)	×	×	×	×	(II)

<sup>\*</sup> Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

# **Relevant Therapy Summary (continued)**

■ In this cancer type
O In other cancer type
O In this cancer type and other cancer types
X No evidence

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials <sup>3</sup>
sotorasib, durvalumab	×	×	×	×	<b>(II)</b>
afatinib, selumetinib	×	×	×	×	<b>(</b>  /  )
avutometinib, sotorasib, defactinib	×	×	×	×	(I/II)
D-1553, ifebemtinib	×	×	×	×	<b>(</b> 1/11)
DCC-3116, sotorasib	×	×	×	×	<b>(</b> I/II)
divarasib, pembrolizumab, chemotherapy	×	×	×	×	<b>(</b> 1/11)
ERAS-0015	×	×	×	×	<b>(</b>  /  )
FMC-376	×	×	×	×	<b>(</b> I/II)
glecirasib, JAB-3312	×	×	×	×	<b>(</b> I/II)
HBI 2376, D-1553	×	×	×	×	<b>(</b> I/II)
HS-10370	×	×	×	×	<b>(</b>  /  )
HYP-2090PTSA	×	×	×	×	<b>●</b> (I/II)
IMM-1-104	×	×	×	×	(I/II)
MRTX0902, adagrasib	×	×	×	×	<b>(</b> I/II)
RMC-6291, pembrolizumab, chemotherapy, RMC-6236	×	×	×	×	<b>(</b> I/II)
YL-15293	×	×	×	×	<b>(</b> I/II)
ZG-19018	×	×	×	×	(I/II)
zotatifin, sotorasib	×	×	×	×	<b>(</b> I/II)
adagrasib, olaparib	×	×	×	×	(I)
AMG 193, sotorasib	×	×	×	×	<b>(</b> l)
BAY-3498264, sotorasib	×	×	×	×	(I)
BBO-8520, pembrolizumab	×	×	×	×	(I)
BEBT-607	×	×	×	×	(I)
BMS-986488, adagrasib	×	×	×	×	<b>(</b> I)
BPI-421286	×	×	×	×	<b>(</b> I)
carfilzomib, sotorasib	×	×	×	×	<b>(</b> I)
divarasib, bevacizumab, RLY-1971, inavolisib	×	×	×	×	<b>(</b> l)
GEC-255	×	×	×	×	(I)
HBI-2438	×	×	×	×	<b>(</b> I)

<sup>\*</sup> Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

# **Relevant Therapy Summary (continued)**

In this cancer type

O In other cancer type

• In this cancer type and other cancer types

× No evidence

# KRAS p.(G12C) c.34G>T (continued)

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
HMPL-415	×	×	×	×	<b>(</b> 1)
HRS-7058	×	×	×	×	<b>(</b> 1)
JAB-3312	×	×	×	×	<b>(</b> 1)
JSKN-016	×	×	×	×	<b>(</b> 1)
KO-2806, adagrasib	×	×	×	×	<b>(</b> I)
KQB-365	×	×	×	×	<b>(</b> 1)
KRAS peptide vaccine, poly-ICLC, nivolumab, ipilimumab	×	×	×	×	<b>(</b> 1)
KRAS-EphA-2-CAR-DC, anti-PD-1, ipilimumab	×	×	×	×	<b>(</b> I)
ladarixin, sotorasib	×	×	×	×	<b>(</b> I)
MK-0472, MK-1084	×	×	×	×	<b>(</b> 1)
MK-1084	×	×	×	×	<b>(</b> 1)
Nest-1	×	×	×	×	<b>(</b> 1)
patritumab deruxtecan	×	×	×	×	<b>(</b> 1)
RMC-6291, RMC-6236	×	×	×	×	<b>(</b> 1)
sotorasib, afatinib, pembrolizumab, atezolizumab, chemotherapy, BI-1701963	×	×	×	×	<b>(</b> I)
sotorasib, radiation therapy	×	×	×	×	<b>(</b> I)
SY-5933	×	×	×	×	<b>(</b> I)
ZEN-3694, binimetinib	×	×	×	×	<b>(</b> I)

# **MTAP** deletion

Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
AMG 193	×	×	×	×	<b>(</b> I/II)
TNG-456, abemaciclib	×	×	×	×	<b>(</b> I/II)
TNG-462	×	×	×	×	<b>(</b> I/II)
AMG 193, sotorasib	×	×	×	×	(I)
GTA-182	×	×	×	×	(I)
ISM-3412	×	×	×	×	(I)
MRTX-1719	×	×	×	×	(I)

 $<sup>\</sup>star$  Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

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# **Relevant Therapy Summary (continued)**

■ In this cancer type
O In other cancer type
In this cancer type and other cancer types
X No evidence

MTAP deletion (continued)					
Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
PH020-803	×	×	×	×	<b>(</b> l)
S-095035	×	×	×	×	<b>(</b> 1)
SYH-2039	×	×	×	×	(I)

#### FDA Clinical Trials\* NCCN **EMA ESMO** Relevant Therapy palbociclib × × × × (II) palbociclib, abemaciclib × × × × (II) **AMG 193** × × × × **(**|/||)

CDKN2B deletion					
Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
palbociclib, abemaciclib	×	×	×	×	<b>(II)</b>

FANCIVI deletion					
Relevant Therapy	FDA	NCCN	EMA	ESMO	Clinical Trials*
pamiparib, tislelizumab	×	×	×	×	<b>(II)</b>

<sup>\*</sup> Most advanced phase (IV, III, II/III, II, I/II, I) is shown and multiple clinical trials may be available.

#### **HRR Details**

EANCM deletion

**CDKN2A** deletion

Gene/Genomic Alteration	Finding
LOH percentage	5.6%
RAD51B	CNV, CN:1.0
RAD51B	LOH, 14q24.1(68290164-69061406)x1

Homologous recombination repair (HRR) genes were defined from published evidence in relevant therapies, clinical guidelines, as well as clinical trials, and include - BRCA1, BRCA2, ATM, BARD1, BRIP1, CDK12, CHEK1, CHEK2, FANCL, PALB2, RAD51B, RAD51C, RAD51D, and RAD54L.

Thermo Fisher Scientific's Ion Torrent Oncomine Reporter software was used in generation of this report. Software was developed and designed internally by Thermo Fisher Scientific. The analysis was based on Oncomine Reporter (6.1.1 data version 2025.06(006)). The data presented here are from a curated knowledge base of publicly available information, but may not be exhaustive. FDA information was sourced from www.fda.gov and is current as of 2025-05-14. NCCN information was sourced from www.nccn.org and is current as of 2025-05-01. EMA information was sourced from www.ema.europa.eu and is current as of 2025-05-14. ESMO information was sourced from www.esmo.org and is current as of 2025-05-01. Clinical Trials information is current as of 2025-05-01. For the most upto-date information regarding a particular trial, search www.clinicaltrials.gov by NCT ID or search local clinical trials authority website by local identifier listed in 'Other identifiers.' Variants are reported according to HGVS nomenclature and classified following AMP/ ASCO/CAP guidelines (Li et al. 2017). Based on the data sources selected, variants, therapies, and trials listed in this report are listed in order of potential clinical significance but not for predicted efficacy of the therapies.

### References

- 1. Pylayeva-Gupta et al. RAS oncogenes: weaving a tumorigenic web. Nat. Rev. Cancer. 2011 Oct 13;11(11):761-74. PMID: 21993244
- 2. Karnoub et al. Ras oncogenes: split personalities. Nat. Rev. Mol. Cell Biol. 2008 Jul;9(7):517-31. PMID: 18568040
- Scott et al. Therapeutic Approaches to RAS Mutation. Cancer J. 2016 May-Jun;22(3):165-74. doi: 10.1097/ PP0.0000000000187. PMID: 27341593
- 4. Weinstein et al. The Cancer Genome Atlas Pan-Cancer analysis project. Nat. Genet. 2013 Oct;45(10):1113-20. PMID: 24071849
- 5. Román et al. KRAS oncogene in non-small cell lung cancer: clinical perspectives on the treatment of an old target. Mol Cancer. 2018 Feb 19;17(1):33. doi: 10.1186/s12943-018-0789-x. PMID: 29455666
- Dinu et al. Prognostic significance of KRAS gene mutations in colorectal cancer--preliminary study. J Med Life. 2014 Oct-Dec;7(4):581-7. PMID: 25713627
- 7. Cerami et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov. 2012 May;2(5):401-4. PMID: 22588877
- 8. Allegra et al. Extended RAS Gene Mutation Testing in Metastatic Colorectal Carcinoma to Predict Response to Anti-Epidermal Growth Factor Receptor Monoclonal Antibody Therapy: American Society of Clinical Oncology Provisional Clinical Opinion Update 2015. J. Clin. Oncol. 2016 Jan 10;34(2):179-85. PMID: 26438111
- https://www.accessdata.fda.gov/drugsatfda\_docs/label/2025/2146650rig1s009correctedlbl.pdf
- 10. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2024/216340s005lbl.pdf
- 11. NCCN Guidelines® NCCN-Pancreatic Adenocarcinoma [Version 2.2025]
- 12. https://assets.cwp.roche.com/f/126832/x/5738a7538b/irp230202.pdf
- 13. https://bridgebio.com/news/bridgebio-pharma-announces-first-lung-cancer-patient-dosed-in-phase-1-2-trial-and-us-fda-fast-track-designation-for-shp2-inhibitor-bbp-398-in-combination-with-amgens-lumakras-sotorasib/
- 14. https://investor.verastem.com/news-releases/news-release-details/verastem-oncology-granted-fast-track-designation-combination
- 15. https://www.businesswire.com/news/home/20250109170439/en/
- 16. https://www.d3bio.com/press-releases/d3-bios-d3s-001-receives-u-s-fda-fast-track-designation-for-the-treatment-of-colorectal-cancer-with-kras-g12c-mutation
- 17. https://cardiffoncology.com/wp-content/uploads/2021/07/Cardiff\_Oncology\_Investor\_Presentation-\_July\_2021.pdf
- 18. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2021/125084s279lbl.pdf
- 19. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2025/125147s213lbl.pdf
- 20. Slebos et al. K-ras oncogene activation as a prognostic marker in adenocarcinoma of the lung. N. Engl. J. Med. 1990 Aug 30;323(9):561-5. PMID: 2199829
- 21. O'Leary et al. Reference sequence (RefSeq) database at NCBI: current status, taxonomic expansion, and functional annotation. Nucleic Acids Res. 2016 Jan 4;44(D1):D733-45. PMID: 26553804
- 22. Niraj et al. The Fanconi Anemia Pathway in Cancer. Annu Rev Cancer Biol. 2019 Mar;3:457-478. PMID: 30882047
- 23. Rodríguez et al. Fanconi anemia pathway. Curr Biol. 2017 Sep 25;27(18):R986-R988. PMID: 28950089
- 24. Garcia-Higuera et al. Interaction of the Fanconi anemia proteins and BRCA1 in a common pathway. Mol. Cell. 2001 Feb;7(2):249-62. PMID: 11239454
- 25. Hussain et al. Direct interaction of FANCD2 with BRCA2 in DNA damage response pathways. Hum. Mol. Genet. 2004 Jun 15;13(12):1241-8. PMID: 15115758
- 26. Lord et al. BRCAness revisited. Nat. Rev. Cancer. 2016 Feb;16(2):110-20. PMID: 26775620
- 27. Byrum et al. Defining and Modulating 'BRCAness'. Trends Cell Biol. 2019 Sep;29(9):740-751. PMID: 31362850
- 28. Michl et al. Interplay between Fanconi anemia and homologous recombination pathways in genome integrity. EMBO J. 2016 May 2;35(9):909-23. PMID: 27037238
- 29. Abbasi et al. A rare FANCA gene variation as a breast cancer susceptibility allele in an Iranian population. Mol Med Rep. 2017 Jun;15(6):3983-3988. PMID: 28440412
- 30. Stoepker et al. DNA helicases FANCM and DDX11 are determinants of PARP inhibitor sensitivity. DNA Repair (Amst). 2015 Feb;26:54-64. PMID: 25583207
- 31. Jalan et al. Emerging Roles of RAD52 in Genome Maintenance. Cancers (Basel). 2019 Jul 23;11(7). PMID: 31340507
- 32. Yasuhara et al. Human Rad52 Promotes XPG-Mediated R-loop Processing to Initiate Transcription-Associated Homologous Recombination Repair. Cell. 2018 Oct 4;175(2):558-570.e11. PMID: 30245011

- 33. Prakash et al. Homologous recombination and human health: the roles of BRCA1, BRCA2, and associated proteins. Cold Spring Harb Perspect Biol. 2015 Apr 1;7(4):a016600. PMID: 25833843
- 34. Liu et al. Role of RAD51C and XRCC3 in genetic recombination and DNA repair. J Biol Chem. 2007 Jan 19;282(3):1973-9. PMID: 17114795
- 35. Wilson et al. FANCG promotes formation of a newly identified protein complex containing BRCA2, FANCD2 and XRCC3. Oncogene. 2008 Jun 12;27(26):3641-52. PMID: 18212739
- 36. Geiger et al. Role of the Nuclear Receptor Corepressor 1 (NCOR1) in Atherosclerosis and Associated Immunometabolic Diseases. Front Immunol. 2020;11:569358. PMID: 33117357
- 37. Martínez-Iglesias et al. Tumor suppressive actions of the nuclear receptor corepressor 1. Pharmacol Res. 2016 Jun;108:75-79. PMID: 27149915
- 38. Bhaskara et al. Hdac3 is essential for the maintenance of chromatin structure and genome stability. Cancer Cell. 2010 Nov 16;18(5):436-47. PMID: 21075309
- 39. Mottis et al. Emerging roles of the corepressors NCoR1 and SMRT in homeostasis. Genes Dev. 2013 Apr 15;27(8):819-35. PMID: 23630073
- 40. Noblejas-López et al. Evaluation of transcriptionally regulated genes identifies NCOR1 in hormone receptor negative breast tumors and lung adenocarcinomas as a potential tumor suppressor gene. PLoS One. 2018;13(11):e0207776. PMID: 30485330
- 41. Li. Mechanisms and functions of DNA mismatch repair. Cell Res. 2008 Jan;18(1):85-98. PMID: 18157157
- 42. Kadyrova et al. Human MutLγ, the MLH1-MLH3 heterodimer, is an endonuclease that promotes DNA expansion. Proc Natl Acad Sci U S A. 2020 Feb 18;117(7):3535-3542. PMID: 32015124
- 43. Al-Sweel et al. mlh3 mutations in baker's yeast alter meiotic recombination outcomes by increasing noncrossover events genome-wide. PLoS Genet. 2017 Aug;13(8):e1006974. PMID: 28827832
- 44. Narayanan et al. Tumor Infiltrating Lymphocytes and Macrophages Improve Survival in Microsatellite Unstable Colorectal Cancer. Sci Rep. 2019 Sep 17;9(1):13455. PMID: 31530839
- 45. Croft et al. Regulating the conversion between rounded and elongated modes of cancer cell movement. Cancer Cell. 2008 Nov 4;14(5):349-51. PMID: 18977323
- 46. Garrett et al. Reoperative median sternotomy. Ann Thorac Surg. 1989 Aug;48(2):305. PMID: 2764627
- 47. Héraud et al. Cells. 2019 Apr 12;8(4). PMID: 31013840
- 48. Zhao et al. Glucocorticoid receptor DNA binding factor 1 expression and osteosarcoma prognosis. Tumour Biol. 2014 Dec;35(12):12449-58. PMID: 25185653
- 49. Jonsson et al. Fresh gas flow in coaxial Mapleson A and D circuits during spontaneous breathing. Acta Anaesthesiol Scand. 1986 Oct;30(7):588-93. PMID: 3101384
- 50. Sakamoto et al. Distinct roles of EGF repeats for the Notch signaling system. Exp. Cell Res. 2005 Jan 15;302(2):281-91. PMID: 15561108
- 51. Bray. Notch signalling in context. Nat. Rev. Mol. Cell Biol. 2016 Nov;17(11):722-735. PMID: 27507209
- 52. Kopan et al. The canonical Notch signaling pathway: unfolding the activation mechanism. Cell. 2009 Apr 17;137(2):216-33. PMID: 19379690
- 53. Lobry et al. Oncogenic and tumor suppressor functions of Notch in cancer: it's NOTCH what you think. J. Exp. Med. 2011 Sep 26;208(10):1931-5. PMID: 21948802
- 54. Goriki et al. Unravelling disparate roles of NOTCH in bladder cancer. Nat Rev Urol. 2018 Jun;15(6):345-357. PMID: 29643502
- 55. Wang et al. Loss-of-function mutations in Notch receptors in cutaneous and lung squamous cell carcinoma. Proc. Natl. Acad. Sci. U.S.A. 2011 Oct 25;108(43):17761-6. PMID: 22006338
- 56. Xiu et al. The role of oncogenic Notch2 signaling in cancer: a novel therapeutic target. Am J Cancer Res. 2019;9(5):837-854. PMID: 31218097
- 57. Harasawa et al. Chemotherapy targeting methylthioadenosine phosphorylase (MTAP) deficiency in adult T cell leukemia (ATL). Leukemia. 2002 Sep;16(9):1799-807. PMID: 12200696
- 58. Bertino et al. Targeting tumors that lack methylthioadenosine phosphorylase (MTAP) activity: current strategies. Cancer Biol Ther. 2011 Apr 1;11(7):627-32. PMID: 21301207
- Katya et al. Cancer Dependencies: PRMT5 and MAT2A in MTAP/p16-Deleted Cancers. 10.1146/annurevcancerbio-030419-033444
- 60. Aharoni et al. Dynamical comparison between Drosha and Dicer reveals functional motion similarities and dissimilarities. PLoS One. 2019;14(12):e0226147. PMID: 31821368

- 61. Lee et al. MicroRNAs in cancer. Annu Rev Pathol. 2009;4:199-227. PMID: 18817506
- 62. Hammond. An overview of microRNAs. Adv Drug Deliv Rev. 2015 Jun 29;87:3-14. PMID: 25979468
- 63. Wen et al. Biosci Rep. 2018 Jun 29;38(3). PMID: 29654164
- 64. Kumar et al. Impaired microRNA processing enhances cellular transformation and tumorigenesis. Nat Genet. 2007 May;39(5):673-7. PMID: 17401365
- 65. Robertson et al. DICER1 Syndrome: DICER1 Mutations in Rare Cancers. Cancers (Basel). 2018 May 15;10(5). PMID: 29762508
- 66. Xia et al. Dominant role of CDKN2B/p15INK4B of 9p21.3 tumor suppressor hub in inhibition of cell-cycle and glycolysis. Nat Commun. 2021 Apr 6;12(1):2047. PMID: 33824349
- 67. Scruggs et al. Loss of CDKN2B Promotes Fibrosis via Increased Fibroblast Differentiation Rather Than Proliferation. Am. J. Respir. Cell Mol. Biol. 2018 Aug;59(2):200-214. PMID: 29420051
- 68. Roussel. The INK4 family of cell cycle inhibitors in cancer. Oncogene. 1999 Sep 20;18(38):5311-7. PMID: 10498883
- 69. Aytac et al. Rb independent inhibition of cell growth by p15(INK4B). Biochem. Biophys. Res. Commun. 1999 Aug 27;262(2):534-8. PMID: 10462509
- 70. Hill et al. The genetics of melanoma: recent advances. Annu Rev Genomics Hum Genet. 2013;14:257-79. PMID: 23875803
- 71. Kim et al. The regulation of INK4/ARF in cancer and aging. Cell. 2006 Oct 20;127(2):265-75. PMID: 17055429
- 72. Sekulic et al. Malignant melanoma in the 21st century: the emerging molecular landscape. Mayo Clin. Proc. 2008 Jul;83(7):825-46. PMID: 18613999
- 73. Orlow et al. CDKN2A germline mutations in individuals with cutaneous malignant melanoma. J. Invest. Dermatol. 2007 May;127(5):1234-43. PMID: 17218939
- 74. Bartsch et al. CDKN2A germline mutations in familial pancreatic cancer. Ann. Surg. 2002 Dec;236(6):730-7. PMID: 12454511
- 75. Adib et al. CDKN2A Alterations and Response to Immunotherapy in Solid Tumors. Clin Cancer Res. 2021 Jul 15;27(14):4025-4035. PMID: 34074656
- 76. NCCN Guidelines® NCCN-Mesothelioma: Peritoneal [Version 2.2025]
- 77. NCCN Guidelines® NCCN-Mesothelioma: Pleural [Version 2.2025]
- 78. NCCN Guidelines® NCCN-Soft Tissue Sarcoma [Version 5.2024]
- 79. Louis et al. cIMPACT-NOW update 6: new entity and diagnostic principle recommendations of the cIMPACT-Utrecht meeting on future CNS tumor classification and grading. Brain Pathol. 2020 Jul;30(4):844-856. PMID: 32307792
- 80. Longwen et al. Frequent genetic aberrations in the cell cycle related genes in mucosal melanoma indicate the potential for targeted therapy. J Transl Med. 2019 Jul 29;17(1):245. PMID: 31358010
- 81. Logan et al. PD-0332991, a potent and selective inhibitor of cyclin-dependent kinase 4/6, demonstrates inhibition of proliferation in renal cell carcinoma at nanomolar concentrations and molecular markers predict for sensitivity. Anticancer Res. 2013 Aug;33(8):2997-3004. PMID: 23898052
- 82. von et al. Preclinical Characterization of Novel Chordoma Cell Systems and Their Targeting by Pharmocological Inhibitors of the CDK4/6 Cell-Cycle Pathway. Cancer Res. 2015 Sep 15;75(18):3823-31. PMID: 26183925
- 83. Cen et al. p16-Cdk4-Rb axis controls sensitivity to a cyclin-dependent kinase inhibitor PD0332991 in glioblastoma xenograft cells. Neuro-oncology. 2012 Jul;14(7):870-81. PMID: 22711607
- 84. Vitzthum et al. The role of p16 as a biomarker in nonoropharyngeal head and neck cancer. Oncotarget. 2018 Sep 7;9(70):33247-33248. PMID: 30279955
- 85. Chung et al. p16 protein expression and human papillomavirus status as prognostic biomarkers of nonoropharyngeal head and neck squamous cell carcinoma. J. Clin. Oncol. 2014 Dec 10;32(35):3930-8. PMID: 25267748
- 86. Bryant et al. Prognostic Role of p16 in Nonoropharyngeal Head and Neck Cancer. J. Natl. Cancer Inst. 2018 Dec 1;110(12):1393-1399. PMID: 29878161
- 87. Stephen et al. Significance of p16 in Site-specific HPV Positive and HPV Negative Head and Neck Squamous Cell Carcinoma. Cancer Clin Oncol. 2013;2(1):51-61. PMID: 23935769
- 88. Jafri et al. Germline Mutations in the CDKN2B Tumor Suppressor Gene Predispose to Renal Cell Carcinoma. . Cancer Discov.2015 Jul;5(7):723-9. PMID: 25873077
- 89. Tu et al. CDKN2B deletion is essential for pancreatic cancer development instead of unmeaningful co-deletion due to juxtaposition to CDKN2A. Oncogene. 2018 Jan 4;37(1):128-138. PMID: 28892048
- 90. Hulpke et al. The MHC I loading complex: a multitasking machinery in adaptive immunity. Trends Biochem Sci. PMID: 23849087

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Report Date: 24 Oct 2025

- 91. Adams et al. The adaptable major histocompatibility complex (MHC) fold: structure and function of nonclassical and MHC class I-like molecules. Annu Rev Immunol. 2013;31:529-61. PMID: 23298204
- 92. Rossjohn et al. T cell antigen receptor recognition of antigen-presenting molecules. Annu Rev Immunol. 2015;33:169-200. PMID: 25493333
- 93. Parham. MHC class I molecules and KIRs in human history, health and survival. Nat Rev Immunol. 2005 Mar;5(3):201-14. PMID: 15719024
- 94. Sidney et al. HLA class I supertypes: a revised and updated classification. BMC Immunol. 2008 Jan 22;9:1. PMID: 18211710
- 95. Cornel et al. MHC Class I Downregulation in Cancer: Underlying Mechanisms and Potential Targets for Cancer Immunotherapy. Cancers (Basel). 2020 Jul 2;12(7). PMID: 32630675
- 96. Hurlin et al. The MAX-interacting transcription factor network. Semin. Cancer Biol. 2006 Aug;16(4):265-74. PMID: 16908182
- 97. Susan. An Overview of the Basic Helix-Loop-Helix Proteins. Genome Biol. 2004;5(6):226. PMID: 15186484
- 98. Cascón et al. MAX and MYC: a heritable breakup. Cancer Res. 2012 Jul 1;72(13):3119-24. PMID: 22706201
- 99. Wang et al. MAX is an epigenetic sensor of 5-carboxylcytosine and is altered in multiple myeloma. Nucleic Acids Res. 2017 Mar 17;45(5):2396-2407. PMID: 27903915
- 100. Pritchard et al. Molecular pathways: mitogen-activated protein kinase pathway mutations and drug resistance. Clin. Cancer Res. 2013 May 1;19(9):2301-9. PMID: 23406774
- 101. Lee et al. Targeting MAPK Signaling in Cancer: Mechanisms of Drug Resistance and Sensitivity. Int J Mol Sci. 2020 Feb 7;21(3). PMID: 32046099
- 102. Bubici et al. JNK signalling in cancer: in need of new, smarter therapeutic targets. Br J Pharmacol. 2014 Jan;171(1):24-37. PMID: 24117156
- 103. Ahn et al. Map2k4 functions as a tumor suppressor in lung adenocarcinoma and inhibits tumor cell invasion by decreasing peroxisome proliferator-activated receptor γ2 expression. Mol. Cell. Biol. 2011 Nov;31(21):4270-85. PMID: 21896780
- 104. Robinson et al. Mitogen-activated protein kinase kinase 4/c-Jun NH2-terminal kinase kinase 1 protein expression is subject to translational regulation in prostate cancer cell lines. Mol. Cancer Res. 2008 Mar;6(3):501-8. PMID: 18337456
- 105. Xue et al. MAP3K1 and MAP2K4 mutations are associated with sensitivity to MEK inhibitors in multiple cancer models. Cell Res. 2018 Jul;28(7):719-729. PMID: 29795445
- 106. Sullivan et al. RAD-ical New Insights into RAD51 Regulation. Genes (Basel). 2018 Dec 13;9(12). PMID: 30551670
- 107. Suwaki et al. RAD51 paralogs: roles in DNA damage signalling, recombinational repair and tumorigenesis. Semin. Cell Dev. Biol. 2011 Oct;22(8):898-905. PMID: 21821141
- 108. Chun et al. Rad51 paralog complexes BCDX2 and CX3 act at different stages in the BRCA1-BRCA2-dependent homologous recombination pathway. Mol. Cell. Biol. 2013 Jan;33(2):387-95. PMID: 23149936
- 109. Lim et al. Evaluation of the methods to identify patients who may benefit from PARP inhibitor use. Endocr. Relat. Cancer. 2016 Jun;23(6):R267-85. PMID: 27226207
- 110. Date et al. Haploinsufficiency of RAD51B causes centrosome fragmentation and aneuploidy in human cells. Cancer Res. 2006 Jun 15;66(12):6018-24. PMID: 16778173
- 111. Pelttari et al. RAD51B in Familial Breast Cancer. PLoS ONE. 2016;11(5):e0153788. PMID: 27149063
- 112. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2023/208558s028lbl.pdf
- 113. https://www.senhwabio.com//en/news/20220125
- 114. Marteijn et al. Understanding nucleotide excision repair and its roles in cancer and ageing. Nat Rev Mol Cell Biol. 2014 Jul;15(7):465-81. PMID: 24954209
- 115. Cleaver et al. Disorders of nucleotide excision repair: the genetic and molecular basis of heterogeneity. Nat Rev Genet. 2009 Nov;10(11):756-68. PMID: 19809470
- 116. Hirotsu et al. Genomic Profiling Identified ERCC2 E606Q Mutation in Helicase Domain Respond to Platinum-Based Neoadjuvant Therapy in Urothelial Bladder Cancer. Front Oncol. 2020;10:1643. PMID: 32984035
- 117. Van et al. Somatic ERCC2 mutations correlate with cisplatin sensitivity in muscle-invasive urothelial carcinoma. Cancer Discov. 2014 Oct;4(10):1140-53. PMID: 25096233
- 118. Lander et al. Initial sequencing and analysis of the human genome. Nature. 2001 Feb 15;409(6822):860-921. PMID: 11237011
- 119. Baudrin et al. Molecular and Computational Methods for the Detection of Microsatellite Instability in Cancer. Front Oncol. 2018 Dec 12;8:621. doi: 10.3389/fonc.2018.00621. eCollection 2018. PMID: 30631754
- 120. Nojadeh et al. Microsatellite instability in colorectal cancer. EXCLI J. 2018;17:159-168. PMID: 29743854

Report Date: 24 Oct 2025 26 of 26

- 121. Saeed et al. Microsatellites in Pursuit of Microbial Genome Evolution. Front Microbiol. 2016 Jan 5;6:1462. doi: 10.3389/fmicb.2015.01462. eCollection 2015. PMID: 26779133
- 122. Boland et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. Cancer Res. 1998 Nov 15;58(22):5248-57. PMID: 9823339
- 123. Halford et al. Low-level microsatellite instability occurs in most colorectal cancers and is a nonrandomly distributed quantitative trait. Cancer Res. 2002 Jan 1;62(1):53-7. PMID: 11782358
- 124. Imai et al. Carcinogenesis and microsatellite instability: the interrelationship between genetics and epigenetics. Carcinogenesis. 2008 Apr;29(4):673-80. PMID: 17942460
- 125. NCCN Guidelines® NCCN-Colon Cancer [Version 3.2025]
- 126. Pawlik et al. Colorectal carcinogenesis: MSI-H versus MSI-L. Dis. Markers. 2004;20(4-5):199-206. PMID: 15528785
- 127. Lee et al. Low-Level Microsatellite Instability as a Potential Prognostic Factor in Sporadic Colorectal Cancer. Medicine (Baltimore). 2015 Dec;94(50):e2260. PMID: 26683947
- 128. Latham et al. Microsatellite Instability Is Associated With the Presence of Lynch Syndrome Pan-Cancer. J. Clin. Oncol. 2019 Feb 1;37(4):286-295. PMID: 30376427
- 129. Cortes-Ciriano et al. A molecular portrait of microsatellite instability across multiple cancers. Nat Commun. 2017 Jun 6;8:15180. doi: 10.1038/ncomms15180. PMID: 28585546
- 130. Bonneville et al. Landscape of Microsatellite Instability Across 39 Cancer Types. JCO Precis Oncol. 2017;2017. PMID: 29850653
- 131. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2025/125514s174lbl.pdf
- 132. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2025/125554s129lbl.pdf
- 133. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2024/761174s009lbl.pdf
- 134. NCCN Guidelines® NCCN-Rectal Cancer [Version 2.2025]
- 135. https://www.accessdata.fda.gov/drugsatfda\_docs/label/2025/125377s133lbl.pdf
- 136. Ribic et al. Tumor microsatellite-instability status as a predictor of benefit from fluorouracil-based adjuvant chemotherapy for colon cancer. N. Engl. J. Med. 2003 Jul 17;349(3):247-57. PMID: 12867608
- 137. Klingbiel et al. Prognosis of stage II and III colon cancer treated with adjuvant 5-fluorouracil or FOLFIRI in relation to microsatellite status: results of the PETACC-3 trial. Ann. Oncol. 2015 Jan;26(1):126-32. PMID: 25361982
- 138. Hermel et al. The Emerging Role of Checkpoint Inhibition in Microsatellite Stable Colorectal Cancer. J Pers Med. 2019 Jan 16;9(1). PMID: 30654522
- 139. Ciardiello et al. Immunotherapy of colorectal cancer: Challenges for therapeutic efficacy. Cancer Treat. Rev. 2019 Jun;76:22-32. PMID: 31079031